



Gdański Uniwersytet Medyczny

Rozprawa doktorska

„Niskie stężenie katestatyny jako czynnik ryzyka rozwoju chorób sercowo-naczyniowych – ocena u pacjentów z incydentaloma nadnercza”.

Ewa Zalewska

Promotor: Prof. dr hab. n. med. Krzysztof Sworczak

Promotor pomocniczy: dr Piotr Kmiec

Katedra i Klinika Endokrynologii
i Chorób Wewnętrznych GUMed

Gdańsk 2023

Składam podziękowania wszystkim współautorom i osobom zaangażowanym w uzyskanie materiału do badań będących podstawą publikacji składających się na niniejszą rozprawę doktorską.

Dziękuję moim Promotorom, Panu Profesorowi Krzysztofowi Sworczakowi i Doktorowi Piotrowi Kmeciowi za pomoc i życzliwość przy powstawaniu rozprawy doktorskiej.

Serdecznie dziękuję Rodzinie za wsparcie.

Praca została zrealizowana dzięki wsparciu Gdańskiego Uniwersytetu Medycznego (grant MB 2022: 01-10022 i MN 01-350/08/126) oraz z projektu POWR.03.05.00-00-z082/18 współfinansowanego przez Europejski Fundusz Społeczny w ramach „Operational Programme Knowledge Education Development 2014–2020”.

SPIS TREŚCI

I.	WYKAZ PRAC WCHODZĄCYCH W SKŁAD ROZPRAWY DOKTORSKIEJ.....	4
II.	WYKAZ STOSOWANYCH SKRÓTÓW	5
III.	STRESZCZENIE W JĘZYKU POLSKIM	6
IV.	STRESZCZENIE W JĘZYKU ANGIELSKIM.....	7
V.	WPROWADZENIE	8
VI.	CELE PRACY.....	11
VII.	OMÓWIENIE PUBLIKACJI WCHODZĄCYCH W SKŁAD ROZPRAWY DOKTORSKIEJ.....	11
	A. ARTYKUŁ ORYGINALNY	11
	B. PRACA POGLĄDOWA	16
VIII.	PODSUMOWANIE	18
IX.	PIŚMIENNICTWO	19
X.	PUBLIKACJE WCHODZĄCE W SKŁAD ROZPRAWY DOKTORSKIEJ.....	28
XI.	OŚWIADCZENIA WSPÓŁAUTORÓW	29

I. WYKAZ PRAC WCHODZĄCYCH W SKŁAD ROZPRAWY DOKTORSKIEJ

Praca oryginalna:

Autorzy: Zalewska Ewa, Kmiec Piotr, Sobolewski Jakub, Koprowski Andrzej, Sworczak Krzysztof;

tytuł artykułu: „Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas.”;

Frontiers in Endocrinology, 2023, doi: 10.3389/fendo.2023.1198911.

IF 5.2 | MNiSW 100 | Q1

Praca poglądowa:

Autorzy: Zalewska Ewa, Kmiec Piotr, Sworczak Krzysztof;

tytuł artykułu: “Role of Catestatin in the Cardiovascular System and Metabolic Disorders.”;

Frontiers in Cardiovascular Medicine, 2022, doi:10.3389/fcvm.2022.909480.

IF 3.6 | MNiSW 40 | Q2

Łączna wartość wskaźnika oddziaływania (IF): 8.8

Łączna punktacja MNiSW: 140

II. WYKAZ STOSOWANYCH SKRÓTÓW

ABPM – 24-hour ambulatory blood pressure monitoring; 24 godzinny pomiar ciśnienia tętniczego metodą holter,

AI – adrenal incidentaloma; incydentaloma nadnercza,

BMI – body mass index; wskaźnik masy ciała,

BP – blood pressure; ciśnienie tętnicze,

Cts – catestatin; katestatyna,

CgA – chromogranin A; chromogranina A,

CVDs – cardiovascular diseases; choroby sercowo - naczyniowe,

FRS - Framingham risk score; skala Framingham oceniająca 10-letnie ryzyko miażdżycowej choroby sercowo-naczyniowej oszacowane za pomocą kalkulatora udostępnionego online w 2018r. przez American Heart Association i American College of Cardiology na podstawie Framingham Heart Study,

HDL-C – high density lipoprotein cholesterol; frakcja lipoprotein o wysokiej gęstości,

hs-CRP – high sensitive C-reactive protein; białko C-reaktywne wysokiej czułości,

HT – hypertension; nadciśnienie tętnicze,

LDL-C – low density lipoprotein cholesterol; frakcja lipoprotein o niskiej gęstości,

MACS – mild autonomous cortisol secretion; łagodne autonomiczne wydzielanie kortyzolu,

MetS – metabolic syndrome; zespół metaboliczny,

RVLM - rostral ventrolateral medulla; dogłowy brzuszno-boczny obszar rdzenia przedłużonego,

SCORE2/-OP - Systematic Coronary Risk Estimation 2 /-Older People; kalkulator ryzyka-naczyniowego według Europejskiego Towarzystwa Kardiologicznego 2021,

TTE – transthoracic echocardiography; echokardiografia przezklatkowa.

III. STRESZCZENIE W JĘZYKU POLSKIM

Zwiększoną aktywność współczulnego układu nerwowego odgrywa istotną rolę w patofizjologii nadciśnienia tętniczego (HT) i chorób układu krożenja (CVDs), które są główną przyczyną zgonów na świecie. Katestatyna (Cts) ze względu na swoją zdolność do hamowania dalszego uwalniania katecholamin w mechanizmie ujemnego sprzężenia zwrotnego przeciwdziała nadmiernej aktywacji współczulnej. Na podstawie badań eksperymentalnych wykazano na modelach zwierzęcych, że Cts może prowadzić do normalizacji ciśnienia tętniczego, hamowania ekspresji cytokin prozapalnych, poprawy wrażliwości na insulinę oraz zmniejszenia nasilenia miażdżycy, masy lewej komory, a także grubości kompleksu intima-media. Korzystny wpływ Cts na układ sercowo-naczyniowy i homeostazę metaboliczną został szczegółowo omówiony w mojej pracy poglądowej, będącej częścią rozprawy doktorskiej. Badania kliniczne wskazują, że Cts może stać się nowym biomarkerem, służącym do oceny ryzyka rozwoju i monitorowania przebiegu CVDs. W pracy oryginalnej, zawartej w niniejszej rozprawie doktorskiej, badam związek między Cts a modyfikatorami ryzyka sercowo-naczyniowego, w tym: HT, dyslipidemią, hiperurykemią, a także stężeniem wysokoczułego białka-C-reaktywnego w populacji pacjentów z incydentaloma nadnerczy, którzy są uznawani za grupę osób o podwyższonym ryzyku sercowo-naczyniowym. Ponadto badam związek Cts z bezobjawowym uszkodzeniem narządowym wywołanym przez HT przy pomocy echokardiografii i oceny kompleksu intima-media tętnicy szyjnej wspólnej. Jako pierwsza wykazałam poniższe zależności: 1) wśród osób bez jawnej CVDs innej niż pierwotne HT stężenia osoczowe Cts są porównywalne u pacjentów z gruczolakami nadnercza i tymi z prawidłową morfologią nadnerczy; 2) stężenia Cts są niższe u dorosłych pacjentów z zespołem metabolicznym w porównaniu do osób bez niego; 3) zachodzi ujemna korelacja pomiędzy Cts a ryzykiem sercowo-naczyniowym wyliczonym za pomocą skali Framingham. Konieczne są dalsze, długofalowe badania na większej populacji, aby zastosować Cts jako biomarker CVDs w praktyce klinicznej.

IV. STRESZCZENIE W JĘZYKU ANGIELSKIM

Dissertation topic: "Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas"

Dysregulation of the sympathetic nervous system plays an important role in the pathophysiology of hypertension (HT) and cardiovascular diseases (CVDs), which are the leading cause of death globally. Catestatin (Cts) due to its high capacity to inhibit further release of catecholamines in a negative feedback mechanism counteracts the sympathetic outflow. In the field of experimental research, in animal models, it has been demonstrated that Cts can lead to normalization of blood pressure, inhibition of the expression of pro-inflammatory cytokines, improvement of insulin sensitivity, and reduction of: the severity of atherosclerosis, the left ventricular mass, as well as the thickness of the intima-media complex. The beneficial effects of Cts on the cardiovascular system and the metabolic homeostasis have been discussed in detail in my review paper. Clinical studies point to Cts as a novel biomarker for assessing the risk of development as well as monitoring the course of CVDs. In my original paper I investigated the association between Cts and cardiovascular risk modifiers including HT, dyslipidemia, hyperuricemia, as well as high sensitive-C-reactive protein level in the population of adrenal incidentaloma patients, who are considered to be at an increased cardiovascular risk. Moreover, I examined associations between Cts and asymptomatic HT-mediated organ damage based on echocardiography and examination of the intima-media complex of common carotid artery. I was the first to report: 1) comparable plasma Cts concentrations in adrenal adenoma patients and those of matched controls with normal adrenal morphology among persons without overt CVD other than primary HT, 2) lower Cts in adult patients with metabolic syndrome than those without it; 3) a correlation between Cts and Framingham risk score. Still, longitudinal assessment of larger populations is required to apply serum Cts as a biomarker in clinical practice.

V. WPROWADZENIE

W ciągu ostatnich trzech dekad nastąpił wzrost liczby zgonów z powodu chorób sercowo-naczyniowych (CVDs), które odpowiadają obecnie za blisko jedną trzecią wszystkich zgonów na świecie [World Heart Report 2023]. Szacuje się, że na całym świecie około 1,28 miliarda dorosłych w wieku od 30 do 79 lat cierpi na nadciśnienie tętnicze (HT), które leży u podłożu sztywnienia tętnic prowadząc do: zmniejszenia dopływu krwi i tlenu do serca, uszkodzenia nerek oraz pęknięcia lub zablokowania tętnic zaopatrujących mózg [World Heart Report 2021].

W patofizjologii HT i CVDs zwiększoną aktywność układu współczulnego odgrywa niezaprzeczalnie istotną rolę [de Lucia et al. 2019; Valensi 2021]. Układ współczulny wraz z układem przywspółczulnym należą do układu autonomicznego, który unerwia narządy wewnętrzne i reguluje procesy niezależne od woli. Harmonijne współdziałanie układu współczulnego, odpowiadającego za mobilizację organizmu, i przywspółczulnego, którego działanie przejawia się w spoczynku, jest niezbędne do prawidłowej pracy układu sercowo-naczyniowego. Regulacja aktywności współczulnej odbywa się głównie w mechanizmie odruchów. Ewolucyjnie większe znaczenie dla przycięcia osobnika miało odpowiedź na spadek ciśnienia tętniczego (BP) w wyniku utraty objętości krwi krążącej lub utrudniony dostęp do wody i soli niż przed nadmiernym wzrostem BP. Stąd wynika asymetria odruchu i skuteczniejsza ochrona przed obniżeniem niż nadmiernym podwyższeniem BP [Traczyk 2015].

Wzrost BP pobudza mechanoreceptory zlokalizowane w zatokach szyjnych i w łuku aorty, tzw. baroreceptory tętnicze, co przekazywane jest za pośrednictwem jądra pasma samotnego do doogonowego obszaru brzuszno-bocznego rdzenia przedłużonego w którym zgrupowane są neurony hamujące dogłowy brzuszno-boczny obszar rdzenia przedłużonego (RVLM). W RVLM zlokalizowane są neurony przedwspółczulne, podtrzymujące toniczną aktywność współczulną skierowaną głównie do układu krążenia. W konsekwencji hamowania RVLM dochodzi do odruchowego obniżenia BP. Z kolei obniżenie BP powoduje odbarczenie baroreceptorów tętniczych, co skutkuje odhamowaniem tonicznej aktywności współczulnej, prowadząc do zwiększenia całkowitego oporu obwodowego i pojemności minutowej serca a w konsekwencji powrót prawidłowego BP [Traczyk 2015].

Neurony przedwspółczulne RVLM pobudzają współczulne neurony przedzwojowe rdzenia kręgowego do produkcji acetylocholiny, która działa na receptory cholinergiczne typu N zlokalizowane w neuronach zazwojowych i komórkach chromochłonnych rdzenia nadnerczy prowadząc do ich aktywacji i produkcji katecholamin (CAs): noradrenaliny i adrenaliny [Gaede et al. 2010; Callingham 1965]. CAs łączą się z receptorami α1-adrenergicznymi naczyń prowadząc do ich skurcza, co powoduje wzrost BP, oraz β1-adrenergicznymi serca, skutkiem czego jest: zwiększenie siły skurcza (tzw. dodatni efekt inotropowy), częstości akcji serca (tzw. dodatni efekt chronotropowy), tępia relaksacji (tzw. dodatni efekt luzytropowy) oraz prędkości przewodzenia (tzw. dodatni efekt dromotropowy) [Motiejunaite et al. 2021]. Ponadto pobudzenie receptorów β1-adrenergicznych w nerkach powoduje uwolnienie reniny, która stymuluje produkcję angiotensyny II, ta z kolei uwalnianie aldosteronu przez korę nadnerczy. Stymulacja osi renina-angiotensyna-aldosteron przyczynia się do dalszego wzrostu BP przez zwiększenie całkowitego oporu obwodowego i reabsorpcji wody w nerkach [Traczyk 2015].

CAs są uwalniane przez egzocytozę z pęcherzyków ziarnistych wraz z innymi białkami, w tym chromograniną A (CgA), która do niedawna uznawana była za białko nieaktywne. W 1988 po raz pierwszy wykazano, że CgA podlega proteolitycznej hydrolizie i jest prekursorem dla peptydu mającego zdolność regulacji wydzielania CAs [Simon et al. 1988]. W 1997 Mahata i in. udało się zidentyfikować ten peptyd i nadali mu nazwę "katestatyna" (Cts) ze względu na wysoką zdolność do hamowania dalszego uwalniania CAs w mechanizmie ujemnego sprzężenia zwrotnego [Mahata et al. 1997]. Cts wiąże się z receptorem cholinergicznym typu N, blokując wychwyt jonów sodu, co hamuje depolaryzację błony komórkowej i napływ jonów wapnia przez bramkowane napięciem kanały wapniowe [Taupenot et al. 2000; Herrero et al. 2002]. W konsekwencji dochodzi do zahamowania zarówno dalszego uwalniania CAs przez egzocytozę, jak i transkrypcji genu CgA. Kolejne badania wykazały, iż Cts jest zaangażowana nie tylko w regulację układu sercowo-naczyniowego, ale również immunologicznego, jak i w homeostazę metaboliczną [Bozic et al. 2021; Pasqua et al. 2019; Muntjewerff et al. 2018; Bourebaba et al. 2021]. Dotychczas nie jest znany optymalny zakres stężeń Cts, zapewniający jej plejotropowe działanie biologiczne.

Postulowano, iż Cts może stać się nowym biomarkerem służącym do monitorowania przebiegu i oceny ryzyka rozwoju CVDs [Bozic et al. 2021; O'Connor et al. 2002; 2008; Durakoğlugil et al. 2015; Zhao et al. 2016]. Klasyczne czynniki ryzyka rozwoju CVDs określone zostały na podstawie „Framingham Heart Study” i są nimi: palenie wyrobów tytoniowych, nieaktywny tryb życia, otyłość, wysokie BP, obniżone stężenie frakcji cholesterolu o wysokiej gęstości (HDL-C) i podwyższone stężenie frakcji cholesterolu o niskiej gęstości (LDL-C) [Jahangiry et al. 2017; Mahmood et al. 2014]. Przykładami innych nowych biomarkerów, służących do wyróżnienia osób, które szczególnie skorzystałby z interwencji medycznych (np. obniżenia LDL-C) są kwas moczowy [Yu et al. 2020] i wysokoczułe białko C-reaktywne (hs-CRP) [Koosha et al. 2020]. Wysokie stężenie kwasu moczowego sprzyja rozwojowi CVDs poprzez regulację sygnałów molekularnych takich jak odpowiedź zapalna, stres oksydacyjny, insulinooporność, dysfunkcja śródblonka oraz stres retikulum endoplazmatycznego [Yu et al. 2020]. Rola hs-CRP w patogenezie CVD polega głównie na zapoczątkowaniu i rozwoju zmian miażdżycowych, zwiększeniu prawdopodobieństwa pęknięcia blaszek miażdżycowych oraz promowaniu insulinooporności i cukrzycy typu 2 [Koosha et al. 2020].

Obecność incidentaloma nadnercza o radiologicznym charakterze gruczołaka lub przerostu kory nadnerczy (AI), a zwłaszcza łagodne autonomiczne wydzielanie kortyzolu (MACS) w jej przebiegu, wiązane jest z zaburzeniami metabolicznymi i podwyższonym ryzykiem sercowo-naczyniowym [YAĞI et al. 2022]. Częstość występowania AI wahana się od 1,05% do 8,7% (w zależności od źródła) i rośnie z wiekiem, osiągając szczyt w piątej i szóstej dekadzie życia [Sherlock et al. 2020]. Te dwa fakty – wysoka częstość występowania oraz możliwa podkliniczna aktywność hormonalna – sprawiają, że pacjenci z AI stanowią szczególniewartą zbadania populację w kontekście ryzyka CVDs. Co istotne, stężenie Cts u pacjentów z AI nie było do tej pory oceniane.

VI. CELE PRACY

Obrałam następujące cele:

- 1) określenie stężenia Cts u pacjentów z przypadkowo wykrytą zmianą ogniskową nadnercza o radiologicznym charakterze gruczolaka lub rozrostu kory nadnerczy bez jawnej CVD innej niż HT,
- 2) zbadanie zależności pomiędzy stężeniem Cts a zaburzeniami metabolicznymi, BP oraz wykładnikami podklinicznego uszkodzenia narządowego w przebiegu HT u tych chorych.

VII. OMÓWIENIE PUBLIKACJI WCHODZĄCYCH W SKŁAD ROZPRAWY DOKTORSKIEJ

Na niniejszą rozprawę składa się jedna publikacja oryginalna i jedna poglądowa. W artykule pogladowym przeanalizowana została rola Cts w układzie sercowo-naczyniowym i zaburzeniach metabolicznych na podstawie aktualnego piśmiennictwa [Zalewska et al. 2022]. W ramach pracy oryginalnej zrealizowane zostały zaplanowane cele rozprawy doktorskiej [Zalewska et al. 2023].

A. ARTYKUŁ ORYGINALNY

Artykuł oryginalny „Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas”, (tłumaczenie własne: “Niskie stężenie katesatyny jako czynnik ryzyka rozwoju chorób sercowo – naczyniowych – ocena u pacjentów z incydentaloma nadnercza” przedstawia wyniki przekrojowego, obserwacyjnego badania klinicznego, które zostało przeprowadzone za zgodą Niezależnej Komisji Bioetycznej do spraw Badań Naukowych przy Gdańskim Uniwersytecie Medycznym (NKBB/659/2019).

Podstawowym celem badania była ocena związku między stężeniem Cts a czynnikami ryzyka rozwoju CVD jak i podklinicznym uszkodzeniem narządowym w przebiegu HT u pacjentów z AI bez jawnej CVD innej niż HT.

Uczestników badania rekrutowano kolejno spośród pacjentów z AI hospitalizowanych w Klinice Endokrynologii i Chorób Wewnętrznych Uniwersyteckiego Centrum Klinicznego Gdańskiego Uniwersytetu Medycznego

w okresie od listopada 2018 do lutego 2020 roku celem oceny funkcji wydzielniczej nadnerczy. Do badania włączono 64 pacjentów z radiologicznymi cechami gruczolaka/rozrostu nadnerczy ujawnionymi w tomografii komputerowej lub rezonansie magnetycznym, którzy zgodzili się na udział w nim i nie spełniali żadnego z poniższych kryteriów wykluczenia, którymi były m.in.:

- utrwalona i/lub jawną CVD inną niż pierwotna HT w tym: przewlekła choroba wieńcowa, ostry zespół wieńcowy, udar, przemijający atak niedokrwienienny, choroba tętnic obwodowych, patologiczne arytmie, istotne wady zastawkowe, tamponada serca, kardiomiopatia, wrodzone wady serca, niewydolność serca, żylna choroba zakrzepowo-zatorowa i zapalenie naczyń;
- wiek powyżej 75. roku życia lub poniżej 40. roku życia;
- choroba nerek z eGFR <60 ml/min/1,73m² i/lub białkomoczem >0,25 g/24h);
- otyłość III stopnia (body mass index [BMI] >40 kg/m²);
- czynna choroba nowotworowa i/lub immunologiczna i/lub zakaźna;
- okres przedmenopauzalny;
- terapia antagonistami receptora mineralokortykoidowego;
- występowanie nieprawidłowej funkcji wydzielniczej nadnerczy innej niż MACS, zdefiniowanego jako stężenie kortyzolu w tescie nocnym („overnight”) hamowania z 1 mg deksametazonu między 50 a 138 nmol/l przy niewystępowaniu cech fenotypowych zespołu Cushinga (tzw. MACS); wykluczono zatem osoby z:
 - autonomicznym wydzielaniem kortyzolu charakteryzującym się kortyzolemią ponad 138 nmol/l w tescie nocnym z 1 mg deksametazonu oraz z jawnym klinicznie zespołem Cushinga,
 - pierwotnym aldosteronizmem oraz te z dodatnim wskaźnikiem aldosterononowo-reninowym (powyżej 2 ng/dL:mIU/mL) i jednocześnie występującym HT,
 - osoby z hiperandrogenizmem i tych z
 - guzem chromochłonnym czy nieprawidłowym dobowym wydalaniem pochodnych metoksykatecholamin z moczem.

Na podstawie dokumentacji medycznej naszego szpitala, obejmującej badania zlecone w przychodniach specjalistycznych i na oddziale ratunkowym, zidentyfikowano 24 osoby z prawidłową morfologią nadnerczy w badaniu CT/MRI wykonanym w ciągu 5 lat poprzedzających niniejsze badanie, którzy nie spełnili

żadnego z ustalonych kryteriów wykluczenia i zgodzili się wziąć udział w naszym badaniu jako grupa kontrolna.

Zarówno pacjenci z AI, jak i z grupy kontrolnej przeszli następującą ocenę:

- wywiad lekarski i badanie fizyczne;
- ocenę laboratoryjną stężenia surowiczego/osoczowego: Cts, kreatyniny, elektrolitów (sód, potas), aldosteronu, reniny, lipidogramu (cholesterol całkowity, HDL-C, LDL-C, triglicerydów), kwasu moczowego oraz hs-CRP;
- spoczynkową, 12-odporwadzeniową elektrokardiografię;
- echokardiografię przezklatkową (TTE);
- ocenę maksymalnego wymiaru kompleksu intima-media tętnicy szyjnej wspólnej;
- 24-godzinny, ambulatoryjny pomiar ciśnienia tętniczego metodą holter (ABPM).

Ponadto u pacjentów z AI oceniono stężenie porannego kortyzolu w surowicy, siarczanu dehydroepiandrosteronu i 24-godz. wydalanie kortyzolu, białka i albuminy z moczem, a także wykonano nocny test hamowania kortyzolu z 1 mg deksametazonu (test „overnight”). W teście tym o godz. 23:00 pacjent przyjmuje doustnie 1 mg deksametazonu, a kolejnego dnia o godz. 8:00 ocenia się kortysolemię.

Na podstawie uzyskanych danych wyliczone zostało:

- 10-letnie ryzyko rozwoju CVD przy użyciu kalkulatora z 2018r. udostępnionego online przez American Heart Association and the American College of Cardiology na podstawie Framingham Heart Study (FRS) – <https://www.framinghamheartstudy.org/fhs-risk-functions/cardiovascular-disease-10-year-risk/>;
- dla osób bez cukrzycy 10-letnie ryzyko rozwoju CVD obliczone za pomocą Systematic Coronary Risk Estimation 2 (SCORE2) dla osób w wieku 40-69 lat i SCORE2-Older People (SCORE2-OP) dla osób w wieku 70-75 lat dla krajów o wysokim ryzyku sercowo-naczyniowym. Byliśmy świadomi, że SCORE2/-OP został opracowany w celu oszacowania ryzyka u osób nieotrzymujących dotychczas leków obniżających BP i poziom lipidów, a znaczna część naszych pacjentów stosowała już przynajmniej jedną z tych terapii. Niemniej jednak doszliśmy do wniosku, że zastosowanie tego najpopularniejszego w Polsce narzędzia do oszacowania ryzyka sercowo-naczyniowego będzie wartościowe.

Grupy pacjentów z AI i kontrolna były porównywalne pod względem wieku, płci, BMI, częstości występowania nikotynizmu i HT, cukrzycy typu 2., blaszek miażdżycowych i dyslipidemii. FRS było również porównywalne między pacjentami z AI a grupą kontrolną.

Rozkład Cts był bimodalny zarówno u pacjentów z AI jak i w grupie kontrolnej. W obu grupach u ok. 75% chorych wykazano Cts poniżej 45,2 ng/ml, nie obserwowano pacjentów z Cts w zakresie 45,2-99, a u ok. 25% badanych Cts była w zakresie 100-133 ng/ml. Przed uwzględnieniem potencjalnych czynników zakłócających (płeć, wiek, BMI) stężenie Cts było nieco wyższe u pacjentów z AI: 6,45 (zakres międzykwartylowy: 4,9-37) vs. 4,5 (3,5-28) ng/ml, $p=0,047$. Jednak po przeprowadzeniu analizy regresji wielorakiej obecność AI nie była istotnie związana ze stężeniem Cts ($\beta=-8,1$, $p=0,44$). Ponieważ CgA nie ulega ekspresji w tkance gruczołowej kory nadnerczy, uzyskany wynik porównywalnych stężeń Cts między pacjentami z AI i grupą kontrolną po uwzględnieniu potencjalnych czynników zakłócających był przeze mnie oczekiwany.

Wśród pacjentów z AI, stężenie Cts było wyższe:

- u kobiet niż u mężczyzn: 7,3 (5,5-103) vs. 6 (4,26-7,6) ng/ml, $p=0,03$,
 - u osób z HT w porównaniu do osób z prawidłowym BP: 5,6 (4,36-6,82) vs. 21,7 (6,85-107), $p < 0,001$,
 - oraz u osób z zespołem metabolicznym (MetS) w porównaniu do osób bez MetS: 26 (6,4-116) vs. 5,6 (4,4-7,5) ng/ml, $p < 0,01$,
- niezależnie od potencjalnych czynników zakłócających.

Ponadto u pacjentów z AI stwierdzono ujemne, słabe korelacje między Cts a: BMI ($r=-0,31$) i FRS ($r=-0,42$) oraz dodatnie z HDL-C ($r=0,32$) niezależnie od leczenia statynami. Co ciekawe, wśród uczestników bez terapii obniżającej stężenie cholesterolu wystąpiły również dodatnie korelacje między Cts a: cholesterolom całkowitym i LDL-C ($r=0,36$ dla obu zależności). Ponadto, analizując uczestników badania z grupy badanej i kontrolnej jako całość, zaobserwowałam ujemną korelację między Cts a kwasem moczowym ($r=-0,27$, $p=0,01$), podczas gdy dla każdej grupy analizowanej osobno nie osiągnięto istotności statystycznej dla tej zależności prawdopodobnie ze względu na wielkość próby.

W dalszej analizie wykazałem, że wśród pacjentów z AI ze stężeniem Cts poniżej mediany wynoszącej 6,5 ng/ml (4,9-37 ng/ml) w porównaniu z pozostałymi częściej występowało HT (iloraz szans (OR) 0,17, przedział ufności (CI) 0,05-5,37, $p=0,003$)

i MetS (OR 0,21, CI 0,06-7,51, p=0,018). Co więcej BMI, 24-godzinne średnie skurczowe BP i FRS były również wyższe u tych ze stężeniami Cts poniżej mediany (odpowiednio $30,1 \pm 4$ vs. $27,2 \pm 3,6$ kg/m², p=0,004; $123 \pm 7,4$ vs. $117 \pm 9,5$ mmHg, p=0,022; 13,2% (8,9-19,2) vs. 6,3% (4,2-10,8), p=0,002).

Ponadto podzieliłam uczestników AI na cztery podgrupy w oparciu o rozkład Cts, tj. na tych z: „bardzo niskim” ($<4,9$ ng/ml, n=17), „niskim” ($\geq 4,9$ i $<6,5$ ng/ml, n=15), „pośrednim” ($\geq 6,5$ i $\leq 45,2$ ng/ml, n=17) i „wysokim” (≥ 100 ng/ml, n=15) stężeniem Cts. Analiza wykazała, że płeć męska występowała częściej w podgrupie z „bardzo niskim” w porównaniu do tej z „wysokim” stężeniem Cts (53% vs. 6,7%). Odsetek osób z HT oraz tych z MetS był taki sam w czterech podgrupach stężeń Cts, a istotność statystyczna dla różnic w częstości występowania obu schorzeń uzyskana została dla porównań między osobami z „bardzo niskim” a: „pośrednim” i „wysokim” stężeniem Cts (odpowiednio 82,4% wobec 35,3% i 33,3%, p=0,04 w obu przypadkach). Co więcej, HDL-C było niższe w podgrupie z „bardzo niskim” w porównaniu do trzech pozostałych podgrup Cts ($42,9 \pm 42,9$ wobec $61,6 \pm 17,8$, $56,7 \pm 13,7$ i $55,8 \pm 9,5$ mg/dl, odpowiednio p=0,001, 0,01 i 0,03); a 24-godzinne średnie skurczowe BP było wyższe w podgrupie „niskiego” Cts niż „pośredniego” ($124,7 \pm 6,7$ vs. $114,5 \pm 8,5$ mmHg, p=0,008). FRS w podgrupie „bardzo niskiego” Cts było wyższe niż w podgrupie „pośredniego” i „wysokiego” Cts: 14,3% (19,2-24,5) vs 7,1% (4,7-14,2) i 5,6% (3,8-9,8), odpowiednio p=0,014 i 0,005, zgodnie z różnicami w proporcjach płci, częstości zaburzeń metabolicznych i HT między podgrupami. Nie odnotowano istotnych statystycznie zależności między Cts a parametrami z TTE oraz ultrasonografii tętnicy szyjnej wspólnej.

Najistotniejszym ograniczeniem danych jest niska liczebność badanej populacji w kontekście istotnej heterogenności uczestników (np. w zakresie rozkładu wieku badanych osób czy BMI) i szczególnie niska liczba mężczyzn. Inne wady to brak oznaczania CgA (iloraz Cts do CgA mógłby dostarczyć dodatkowych danych) i badań hormonalnych w grupie kontrolnej. Ponadto bardziej czułe metody oceny śródblonka i serca mogłyby wykazać zależność ze stężeniem Cts, których nie udało się zaobserwować przy zastosowaniu klasycznych parametrów TTE i ultrasonografii tętnicy szyjnej wspólnej.

Należy jednak podkreślić mocne strony pracy. Co najważniejsze, przeprowadziłam dogłębną analizę związku między Cts a czynnikami ryzyka sercowo-naczyniowego i po raz pierwszy wykazałam, że:

- wśród osób bez jawnej CVD innej niż pierwotne HT stężenia Cts w osoczu pacjentów z gruczolakami nadnercza są porównywalne z osobami o prawidłowej morfologii nadnerczy;

- u dorosłych pacjentów z MetS Cts jest niższa niż u osób bez niego;

- zachodzi ujemna korelacja między Cts a FRS.

Konieczne są dalsze, długofalowe badania na większej populacji w celu potwierdzenia, czy niskie stężenie Cts jest czynnikiem ryzyka rozwoju CVD.

B. PRACA POGLĄDOWA

Niniejsza rozprawa doktorska zawiera artykuł przeglądowy „Role of Catestatin in the Cardiovascular System and Metabolic Disorders” (tłumaczenie własne: „Rola katestatyny w układzie sercowo-naczyniowym i zaburzeniach metabolicznych”) [Zalewska et al. 2022]. Artykuł został sporządzony na podstawie przeglądu piśmiennictwa przeprowadzonego w kwietniu 2022r. za pomocą bazy Pubmed z użyciem kluczowego terminu „catestatin”. Na podstawie tytułów i streszczeń wybrałam artykuły kwalifikujące się do omówienia w kolejnych częściach pracy przeglądowej, dotyczących roli Cts w regulacji BP, funkcji i chorób serca oraz zaburzeń metabolicznych, jak i miażdżycy. W każdej z części omówiono badania eksperymentalne *in vitro* i *in vivo* na modelach zwierzęcych, a także badania kliniczne.

W kontekście regulacji BP badania eksperymentalne wskazują, iż Cts oprócz hamowania uwalniania CAz z komórek chromochłonnych i neuronów noradrenergicznych, o czym wspomniałam wcześniej, może wykazywać silne działanie rozszerzające naczynia krwionośne, w którym pośredniczy – przynajmniej częściowo – uwalnianie histaminy [Kennedy et al. 1998; Krüger et al. 2003]. Ujemne efekty inotropowe i luzytropowe Cts mogą również prowadzić do obniżenia BP [Tommaso Angelone et al. 2008]. Ponadto Cts odgrywa rolę w centralnej kontroli krążeniowo-oddechowej [Gaede et al. 2009; 2010]. Na podstawie badań klinicznych wykazano niskie stężenie Cts w stanie przednadciśnieniowym [O'Connor et al. 2002], jednakże wciąż istnieją kontrowersje dotyczące wysokości stężenia Cts u osób z pierwotnym HT. Niektóre badania wykazały porównywalne stężenie Cts u chorych z pierwotnym HT i osób z prawidłowym BP [Durakoğlugil et al. 2015;

O'Connor et al. 2002], inne – niższe w HT [O'Connor et al. 2008; Zalewska et al. 2023], kolejne – wyższe [Meng et al. 2011; Kumric et al. 2022]. Wysunięto hipotezę, że niskie stężenie Cts może predysponować do rozwoju HT, jednakże wyższa aktywność współczulnego układu nerwowego, która wzrasta wraz z postępem HT, przyczynia się do kompensacyjnego wzrostu stężenia Cts [Meng et al. 2011]. Rozwój HT może być również związany ze zmniejszoną konwersją CgA do Cts [Fung et al. 2011; O'Connor et al. 2008; Biswas et al. 2008] i zależeć od wariantów proteolitycznych Cts, uwarunkowanych genetycznie i wywierających różny efekt na BP [Rao et al. 2007].

W kontekście wpływu Cts na funkcję serca *in vitro* i *in vivo* z udziałem modeli zwierzęcych wykazano, że Cts ma potencjalne działanie kardioprotekcyjne, działając jako peptyd kardiodepresyjny [Mazza et al. 2012; Tommaso Angelone et al. 2008; 2011; Imbrogno et al. 2010; T. Angelone et al. 2015], a także zmniejszając apoptozę kardiomiocytów indukowaną stresem oksydacyjnym [D. Wang et al. 2016; Penna et al. 2010; Perrelli et al. 2013; Liao et al. 2015; Chu et al. 2020; Tamm et al. 1994; Brar et al. 2010]. Badania kliniczne sugerują patogenną rolę niskiego stężenia Cts w niedokrwieniu mięśnia sercowego [Xu et al. 2017] oraz na jej udział w przebiegu choroby wieńcowej i niewydolności serca [H. Chen et al. 2019; Zhu et al. 2011; O'Connor et al. 2002]. Podwyższone wartości Cts były obserwowane w momencie dekompensacji niewydolności serca [Liu et al. 2013] i w 3. dobie od wystąpienia ostrego zespołu wieńcowego [X. Wang et al. 2011; Meng et al. 2013; Zhu et al. 2015], co wydaje się potwierdzać kompensacyjny wzrost stężenia Cts do wyższej aktywności współczulnego układu nerwowego. Na podstawie wyników powyższych badań postulowano, iż Cts może znaleźć zastosowanie w monitorowaniu przebiegu CAD i HF [H. Chen et al. 2019; Zhu et al. 2011].

Jeśli chodzi o udział Cts w rozwoju miażdżycy i zaburzeń metabolicznych, badania eksperymentalne wskazują, że Cts może hamować odpowiedź zapalną i interakcje leukocytów z komórkami śródbłonka [Y. Chen et al. 2019; Ying et al. 2021; Kojima et al. 2018; Muntjewerff et al. 2022; Egger et al. 2008], zapobiegać miażdżycy wywołanej przez makrofagi [Y. Chen et al. 2019; Kojima et al. 2018], regulować migrację monocytów [Egger et al. 2008] oraz wytwarzanie i uwalnianie cytokin [Muntjewerff et al. 2022; Egger et al. 2008; Aung et al. 2011]. Wykazano, że Cts jako nowy regulator metabolizmu u gryzoni pomaga w osiągnięciu wrażliwości na insulinę, przewyściężeniu stresu retikulum endoplazmatycznego [Dasgupta et al.

2020], redukcji tkanki tłuszczowej poprzez zwiększenie lipolizy, nasilenie utleniania kwasów tłuszczowych i ich asymilacji do lipidów [Bandyopadhyay et al. 2012]. Na podstawie badań klinicznych wysunięto podejrzenie, iż niskie stężenie Cts związane jest z niekorzystnym profilem metabolicznym, w tym z otyłością, nieprawidłowym stężeniem lipidów i insulinoopornością [Durakoğlugil et al. 2015; O'Connor et al. 2002; Sahu et al. 2012; Simunovic et al. 2019; Kim et al. 2010; Borovac et al. 2019]. Powyższe obserwacje, wraz z wykazanym obniżonym stężeniem Cts w surowicy osób z dodatnim wywiadem rodzinnym w kierunku HT [O'Connor et al. 2002], skłoniły do wysunięcia hipotezy, iż Cts może znaleźć zastosowanie jako nowy czynnik ryzyka rozwoju CVDs [O'Connor et al. 2002; Durakoğlugil et al. 2015].

VIII. PODSUMOWANIE

Aktualne dane wskazują, że endogenny, bioaktywny peptyd Cts jest czynnikiem regulującym homeostazę sercowo-naczyniową i immunometaboliczną, który być może w przyszłości znajdzie zastosowanie jako nowy biomarker CVDs i zaburzeń metabolicznych. Wyniki przeprowadzonej przeze mnie pracy oryginalnej wskazują na związek niskiego stężenia Cts z ryzykiem sercowo-naczyniowym w populacji osób z przypadkowo wykrytymi gruczolakami nadnercza bez jawnej CVDs innej niż HT. Niewystępowanie istotnych różnic w rozkładzie i stężeniu Cts w grupie pacjentów z AI i kontrolnej sugerują, iż uzyskane wyniki badań mogą być odniesione do populacji z prawidłową morfologią nadnerczy. Należy podkreślić, że dotychczas badania nad Cts prowadzono głównie na modelach zwierzęcych i potrzebne są kolejne prace, aby zastosować oznaczenie Cts w praktyce klinicznej czy – w dalszej perspektywie – wykorzystać korzystne działanie Cts w terapii.

IX. PIŚMIENNICTWO

- Angelone, T., A. M. Quintieri, T. Pasqua, E. Filice, P. Cantafio, F. Scavello, C. Rocca, S. K. Mahata, A. Gattuso, and M. C. Cerra. 2015. "The NO Stimulator, Catestatin, Improves the Frank-Starling Response in Normotensive and Hypertensive Rat Hearts." *Nitric Oxide - Biology and Chemistry* 50: 10–19. <https://doi.org/10.1016/j.niox.2015.07.004>.
- Angelone, Tommaso, Anna Maria Quintieri, Bhawanjit K. Brar, Pauline T. Limchaiyawat, Bruno Tota, Sushil K. Mahata, and Maria Carmela Cerra. 2008. "The Antihypertensive Chromogranin A Peptide Catestatin Acts as a Novel Endocrine/Paracrine Modulator of Cardiac Inotropism and Lusitropism." *Endocrinology* 149 (10): 4780–93. <https://doi.org/10.1210/en.2008-0318>.
- Angelone, Tommaso, Anna Maria Quintieri, Teresa Pasqua, Stefano Gentile, Bruno Tota, Sushil K Mahata, and Maria Carmela Cerra. 2011. "Phosphodiesterase Type-2 and NO-Dependent S -Nitrosylation Mediate the Cardioinhibition of the Antihypertensive Catestatin." *American Journal of Physiology-Heart and Circulatory Physiology* 302 (2): H431–H44. <https://doi.org/10.1152/ajpheart.00491.2011>.
- Aung, Gyi, François Niyonsaba, Hiroko Ushio, Naoki Kojiwara, Hirohisa Saito, Shigaku Ikeda, Hideki Ogawa, and Ko Okumura. 2011. "Catestatin, a Neuroendocrine Antimicrobial Peptide, Induces Human Mast Cell Migration, Degranulation and Production of Cytokines and Chemokines." *Immunology* 132 (4): 527–39. <https://doi.org/10.1111/j.1365-2567.2010.03395.x>.
- Bandyopadhyay, Gautam K., Christine U. Vu, Stefano Gentile, Howon Lee, Nilima Biswas, Nai Wen Chi, Daniel T. O'Connor, and Sushil K. Mahata. 2012. "Catestatin (Chromogranin A352-372) and Novel Effects on Mobilization of Fat from Adipose Tissue through Regulation of Adrenergic and Leptin Signaling." *Journal of Biological Chemistry* 287 (27): 23141–51. <https://doi.org/10.1074/jbc.M111.335877>.
- Biswas, Nilima, Sucheta M. Vaingankar, Manjula Mahata, Madhusudan Das, Jiaur R. Gayen, Laurent Taupenot, Justin W. Torpey, Daniel T. O'Connor, and Sushil K. Mahata. 2008. "Proteolytic Cleavage of Human Chromogranin A Containing Naturally Occurring Catestatin Variants: Differential Processing at Catestatin Region by Plasmin." *Endocrinology* 149 (2): 749–57. <https://doi.org/10.1210/en.2007-0838>.

- Borovac, Josip A., Zoran Dogas, Daniela Supe-Domic, Tea Galic, and Josko Bozic. 2019. "Catestatin Serum Levels Are Increased in Male Patients with Obstructive Sleep Apnea." *Sleep and Breathing* 23 (2): 473–81. <https://doi.org/10.1007/s11325-018-1703-x>.
- Bourebaba, Yasmina, Malwina Mularczyk, Krzysztof Marycz, and Lynda Bourebaba. 2021. "Catestatin Peptide of Chromogranin A as a Potential New Target for Several Risk Factors Management in the Course of Metabolic Syndrome." *Biomedicine & Pharmacotherapy* 134 (February): 111113. <https://doi.org/10.1016/J.BIOPHA.2020.111113>.
- Bozic, Josko, Marko Kumric, Tina Ticinovic Kurir, Hrvoje Urlic, Dinko Martinovic, Marino Vilovic, Nada Tomasovic Mrcela, and Josip A. Borovac. 2021. "Catestatin as a Biomarker of Cardiovascular Diseases: A Clinical Perspective." *Biomedicines* 9 (12): 1757. <https://doi.org/10.3390/biomedicines9121757>.
- Brar, Bhawanjit K., Erik Helgeland, Sushil K. Mahata, Kuixing Zhang, Daniel T. O'Connor, Karen B. Helle, and Anne K. Jonassen. 2010. "Human Catestatin Peptides Differentially Regulate Infarct Size in the Ischemic-Reperfused Rat Heart." *Regulatory Peptides* 165 (1): 63–70. <https://doi.org/10.1016/j.regpep.2010.07.153>.
- Callingham, B A. 1965. "The Chromaffin Cell." *Nature* 208 (5005): 6. <https://doi.org/10.1038/208006a0>.
- Chen, Hua, Dongxia Liu, Lan Ge, Tao Wang, Zhenzhen Ma, Yuping Han, Yawei Duan, et al. 2019. "Catestatin Prevents Endothelial Inflammation and Promotes Thrombus Resolution in Acute Pulmonary Embolism in Mice." *Bioscience Reports* 39 (11): 1–10. <https://doi.org/10.1042/BSR20192236>.
- Chen, Yanjia, Xiaoqun Wang, Chendie Yang, Xiuxiu Su, Wenbo Yang, Yang Dai, Hui Han, et al. 2019. "Decreased Circulating Catestatin Levels Are Associated with Coronary Artery Disease: The Emerging Anti-Inflammatory Role." *Atherosclerosis* 281: 78–88. <https://doi.org/10.1016/j.atherosclerosis.2018.12.025>.
- Chu, Song Yun, Fen Peng, Jie Wang, Lin Liu, Lei Meng, Jing Zhao, Xiao Ning Han, and Wen Hui Ding. 2020. "Catestatin in Defense of Oxidative-Stress-Induced Apoptosis: A Novel Mechanism by Activating the Beta2 Adrenergic Receptor and PKB/Akt Pathway in Ischemic-Reperfused Myocardium." *Peptides* 123: 170200. <https://doi.org/10.1016/j.peptides.2019.170200>.
- Dasgupta, Abhijit, Gautam K. Bandyopadhyay, Indrani Ray, Keya Bandyopadhyay,

- Nirmalya Chowdhury, Rajat K. De, and Sushil K. Mahata. 2020. "Catestatin Improves Insulin Sensitivity by Attenuating Endoplasmic Reticulum Stress: In Vivo and in Silico Validation." *Computational and Structural Biotechnology Journal* 18: 464–81. <https://doi.org/10.1016/j.csbj.2020.02.005>.
- Durakoğlugil, M.E., T. Ayaz, S.A. Kocaman, A. Kıbaş, T. Durakoğlugil, T. Erdoğan, M. Çetin, O.Z. Şahin, and Y. Çiçek. 2015. "The Relationship of Plasma Catestatin Concentrations with Metabolic and Vascular Parameters in Untreated Hypertensive Patients: Influence on High-Density Lipoprotein Cholesterol." *Anatolian Journal of Cardiology* 15 (7): 577–85. <https://doi.org/10.5152/akd.2014.5536>.
- Egger, Margot, Arno G.E. Beer, Markus Theurl, Wilfried Schgoer, Benjamin Hotter, Tobias Tatarczyk, Danijela Vasiljevic, et al. 2008. "Monocyte Migration: A Novel Effect and Signaling Pathways of Catestatin." *European Journal of Pharmacology* 598 (1–3): 104–11. <https://doi.org/10.1016/j.ejphar.2008.09.016>.
- Fung, Maple M., Rany M. Salem, Parag Mehtani, Brenda Thomas, Christine F. Lu, Brandon Perez, Fangwen Rao, et al. 2011. "Direct Vasoactive Effects of the Chromogranin A (CHGA) Peptide Catestatin in Humans In Vivo." *Clinical and Experimental Hypertension* 32 (5): 278–87. <https://doi.org/10.1161/CIRCULATIONAHA.110.956839>.
- Gaede, Andrea H., Mandy S.Y. Lung, and Paul M. Pilowsky. 2009. "Catestatin Attenuates the Effects of Intrathecal Nicotine and Isoproterenol." *Brain Research* 1305: 86–95. <https://doi.org/10.1016/j.brainres.2009.09.088>.
- Gaede, Andrea H., and Paul M. Pilowsky. 2010. "Catestatin in Rat RVLM Is Sympathoexcitatory, Increases Barosensitivity, and Attenuates Chemosensitivity and the Somatosympathetic Reflex." *American Journal of Physiology - Regulatory Integrative and Comparative Physiology* 299 (6): 1538–45. <https://doi.org/10.1152/ajpregu.00335.2010>.
- Herrero, Carlos J., Eva Alés, Antonio J. Pintado, Manuela G. López, Esther García-Palomero, Sushil K. Mahata, Daniel T. O'Connor, Antonio G. García, and Carmen Montiel. 2002. "Modulatory Mechanism of the Endogenous Peptide Catestatin on Neuronal Nicotinic Acetylcholine Receptors and Exocytosis." *Journal of Neuroscience* 22 (2): 377–88. <https://doi.org/10.1523/jneurosci.22-02-00377.2002>.
- Imbrogno, Sandra, Filippo Garofalo, Maria Carmela Cerra, Sushil K. Mahata, and

- Bruno Tota. 2010. "The Catecholamine Release-Inhibitory Peptide Catestatin (Chromogranin A344-364) Modulates Myocardial Function in Fish." *Journal of Experimental Biology* 213 (21): 3636–43. <https://doi.org/10.1242/jeb.045567>.
- Jahangiry, Leila, Mahdieh Abbasalizad Farhangi, and Fatemeh Rezaei. 2017. "Framingham Risk Score for Estimation of 10-Years of Cardiovascular Diseases Risk in Patients with Metabolic Syndrome." *Journal of Health, Population and Nutrition* 36 (1): 1–6. <https://doi.org/10.1186/s41043-017-0114-0>.
- Kennedy, Brian P., Sushil K. Mahata, Daniel T. O'Connor, and Michael G. Ziegler. 1998. "Mechanism of Cardiovascular Actions of the Chromogranin A Fragment Catestatin in Vivo." *Peptides* 19 (7): 1241–48. [https://doi.org/10.1016/S0196-9781\(98\)00086-2](https://doi.org/10.1016/S0196-9781(98)00086-2).
- Kim, Jinkwan, Seungkwan Lee, Rakesh Bhattacharjee, Abdelnaby Khalyfa, Leila Kheirandish-Gozal, and David Gozal. 2010. "Leukocyte Telomere Length and Plasma Catestatin and Myeloid-Related Protein 8/14 Concentrations in Children with Obstructive Sleep Apnea." *Chest* 138 (1): 91–99. <https://doi.org/10.1378/chest.09-2832>.
- Kojima, Miho, Nana Ozawa, Yusaku Mori, Yui Takahashi, Kaho Watanabe-Kominato, Remina Shirai, Rena Watanabe, et al. 2018. "Catestatin Prevents Macrophage-Driven Atherosclerosis but Not Arterial Injury-Induced Neointimal Hyperplasia." *Thrombosis and Haemostasis* 118 (1): 182–94. <https://doi.org/10.1160/TH17-05-0349>.
- Koosha, Pooya, Hamidreza Roohafza, Nizal Sarrafzadegan, Mehrbod Vakhshoori, Mohammad Talaei, Erfan Sheikhbahaei, and Masoumeh Sadeghi. 2020. "High Sensitivity C-Reactive Protein Predictive Value for Cardiovascular Disease: A Nested Case Control from Isfahan Cohort Study (ICS)." *Global Heart* 15 (1): 1–13. <https://doi.org/10.5334/GH.367>.
- Krüger, Per Gøran, Sushil K. Mahata, and Karen B. Helle. 2003. "Catestatin (CgA344-364) Stimulates Rat Mast Cell Release of Histamine in a Manner Comparable to Mastoparan and Other Cationic Charged Neuropeptides." *Regulatory Peptides* 114 (1): 29–35. [https://doi.org/10.1016/S0167-0115\(03\)00069-7](https://doi.org/10.1016/S0167-0115(03)00069-7).
- Kumric, Marko, Josip Vrdoljak, Goran Dujic, Daniela Supe-Domic, Tina Ticinovic Kurir, Zeljko Dujic, and Josko Bozic. 2022. "Serum Catestatin Levels Correlate with Ambulatory Blood Pressure and Indices of Arterial Stiffness in Patients with Primary Hypertension." *Biomolecules* 12 (9).

<https://doi.org/10.3390/biom12091204>.

- Liao, Feng, Yang Zheng, Junyan Cai, Jinghui Fan, Jing Wang, Jichun Yang, Qinghua Cui, Guoheng Xu, Chaoshu Tang, and Bin Geng. 2015. "Catestatin Attenuates Endoplasmic Reticulum Induced Cell Apoptosis by Activation Type 2 Muscarinic Acetylcholine Receptor in Cardiac Ischemia/Reperfusion." *Scientific Reports* 5 (1): 16590. <https://doi.org/10.1038/srep16590>.
- Liu, Lin, Wenhui Ding, Renxu Li, Xiaojin Ye, Jing Zhao, Jie Jiang, Lei Meng, et al. 2013. "Plasma Levels and Diagnostic Value of Catestatin in Patients with Heart Failure." *Peptides* 46: 20–25. <https://doi.org/10.1016/j.peptides.2013.05.003>.
- Lucia, Claudio de, Michela Piedepalumbo, Giuseppe Paolisso, and Walter J. Koch. 2019. "Sympathetic Nervous System in Age-Related Cardiovascular Dysfunction: Pathophysiology and Therapeutic Perspective." *International Journal of Biochemistry and Cell Biology* 108: 29–33. <https://doi.org/10.1016/j.biocel.2019.01.004>.
- Mahata, Sushil K., Daniel T. O'Connor, Manjula Mahata, Seung Hyun Yoo, Laurent Taupenot, Hongjiang Wu, Bruce M. Gill, and Robert J. Palmer. 1997. "Novel Autocrine Feedback Control of Catecholamine Release: A Discrete Chromogranin A Fragment Is a Noncompetitive Nicotinic Cholinergic Antagonist." *Journal of Clinical Investigation* 100 (6): 1623–33. <https://doi.org/10.1172/JCI119686>.
- Mahmood, Syed S., Daniel Levy, Ramachandran S. Vasan, and Thomas J. Wang. 2014. "The Framingham Heart Study and the Epidemiology of Cardiovascular Diseases: A Historical Perspective." *Lancet* 383 (9921): 999–1008. [https://doi.org/10.1016/S0140-6736\(13\)61752-3](https://doi.org/10.1016/S0140-6736(13)61752-3).The.
- Mazza, Rosa, Teresa Pasqua, and Alfonsina Gattuso. 2012. "Cardiac Heterometric Response: The Interplay between Catestatin and Nitric Oxide Deciphered by the Frog Heart." *Nitric Oxide - Biology and Chemistry* 27 (1): 40–49. <https://doi.org/10.1016/j.niox.2012.04.003>.
- Meng, Lei, Jie Wang, Wen Hui Ding, Ping Han, Ying Yang, Li Tong Qi, and Bao Wei Zhang. 2013. "Plasma Catestatin Level in Patients with Acute Myocardial Infarction and Its Correlation with Ventricular Remodelling." *Postgraduate Medical Journal* 89 (1050): 193–96. <https://doi.org/10.1136/postgradmedj-2012-131060>.
- Meng, Lei, Xiao Jin Ye, Wen Hui Ding, Ying Yang, Bei Bing Di, Lin Liu, and Yong Huo.

2011. "Plasma Catecholamine Release-Inhibitory Peptide Catestatin in Patients with Essential Hypertension." *Journal of Cardiovascular Medicine* 12 (9): 643–47. <https://doi.org/10.2459/JCM.0b013e328346c142>.
- Motiejunaite, Justina, Laurence Amar, and Emmanuelle Vidal-Petiot. 2021. "Adrenergic Receptors and Cardiovascular Effects of Catecholamines." *Annales d'Endocrinologie* 82 (3–4): 193–97. <https://doi.org/10.1016/j.ando.2020.03.012>.
- Muntjewerff, Elke M., Gina Dunkel, Mara J.T. Nicolaisen, Sushil K. Mahata, and Geert Van Den Bogaart. 2018. "Catestatin as a Target for Treatment of Inflammatory Diseases." *Frontiers in Immunology* 9 (OCT): 1–10. <https://doi.org/10.3389/fimmu.2018.02199>.
- Muntjewerff, Elke M, Gustaf Christoffersson, Sushil K Mahata, and Geert van den Bogaart. 2022. "Putative Regulation of Macrophage-Mediated Inflammation by Catestatin." *Trends in Immunology* 43 (1): 41–50. <https://doi.org/10.1016/j.it.2021.11.002>.
- O'Connor, Daniel T., Mala T. Kailasam, Brian P. Kennedy, Michael G. Ziegler, Noboru Yanaihara, and Robert J. Parmer. 2002. "Early Decline in the Catecholamine Release-Inhibitory Peptide Catestatin in Humans at Genetic Risk of Hypertension." *Journal of Hypertension* 20 (7): 1335–45. <https://doi.org/10.1097/00004872-200207000-00020>.
- O'Connor, Daniel T., Gu Zhu, Fangwen Rao, Laurent Taupenot, Maple M. Fung, Madhusudan Das, Sushil K. Mahata, et al. 2008. "Heritability and Genome-Wide Linkage in US and Australian Twins Identify Novel Genomic Regions Controlling Chromogranin a: Implications for Secretion and Blood Pressure." *Circulation* 118 (3): 247–57. <https://doi.org/10.1161/CIRCULATIONAHA.107.709105>.
- Pasqua, Teresa, Carmine Rocca, Angela Spena, Tommaso Angelone, and Maria Carmela Cerra. 2019. "Modulation of the Coronary Tone in the Expanding Scenario of Chromogranin-A and Its Derived Peptides." *Future Medicinal Chemistry* 11 (12): 1501–11. <https://doi.org/10.4155/fmc-2018-0585>.
- Penna, Claudia, Giuseppe Alloatti, Maria Pia Gallo, Maria Carmela Cerra, Renzo Levi, Francesca Tullio, Eleonora Bassino, et al. 2010. "Catestatin Improves Post-Ischemic Left Ventricular Function and Decreases Ischemia/Reperfusion Injury in Heart." *Cellular and Molecular Neurobiology* 30 (8): 1171–79. <https://doi.org/10.1007/s10571-010-9598-5>.

- Perrelli, Maria Giulia, Francesca Tullio, Carmelina Angotti, Maria Carmela Cerra, Tommaso Angelone, Bruno Tota, Giuseppe Alloatti, Claudia Penna, and Pasquale Pagliaro. 2013. "Catestatin Reduces Myocardial Ischaemia/Reperfusion Injury: Involvement of PI3K/Akt, PKCs, Mitochondrial KATP Channels and ROS Signalling." *Pflugers Archiv European Journal of Physiology* 465 (7): 1031–40. <https://doi.org/10.1007/s00424-013-1217-0>.
- Rao, Fangwen, Gen Wen, Jiaur R. Gayen, Madhusudan Das, Sucheta M. Vaingankar, Brinda K. Rana, Manjula Mahata, et al. 2007. "Catecholamine Release-Inhibitory Peptide Catestatin (Chromogranin A352-372): Naturally Occurring Amino Acid Variant Gly364Ser Causes Profound Changes in Human Autonomic Activity and Alters Risk for Hypertension." *Circulation* 115 (17): 2271–81. <https://doi.org/10.1161/CIRCULATIONAHA.106.628859>.
- Sahu, Bhavani S., Jagan M. Obbineni, Giriraj Sahu, Prasanna K.R. Allu, Lakshmi Subramanian, Parshuram J. Sonawane, Pradeep K. Singh, et al. 2012. "Functional Genetic Variants of the Catecholamine-Release-Inhibitory Peptide Catestatin in an Indian Population: Allele-Specific Effects on Metabolic Traits." *Journal of Biological Chemistry* 287 (52): 43840–52. <https://doi.org/10.1074/jbc.M112.407916>.
- Sherlock, Mark, Andrew Scarsbrook, Afroze Abbas, Sheila Fraser, Padiporn Limumpornpatch, Rosemary Dineen, and Paul M Stewart. 2020. "Adrenal Incidentaloma." *Endocrine Reviews* 41 (6): 775–820. <https://doi.org/10.1210/endrev/bnaa008>.
- Simon, J. P., M. F. Bader, and D. Aunis. 1988. "Secretion from Chromaffin Cells Is Controlled by Chromogranin A-Derived Peptides." *Proceedings of the National Academy of Sciences of the United States of America* 85 (5): 1712–16. <https://doi.org/10.1073/pnas.85.5.1712>.
- Simunovic, Marko, Daniela Supe-Domic, Zeljka Karin, Marina Degoricija, Martina Paradzik, Josko Bozic, Ivana Unic, and Veselin Skrabic. 2019. "Serum Catestatin Concentrations Are Decreased in Obese Children and Adolescents." *Pediatric Diabetes* 20 (5): 549–55. <https://doi.org/10.1111/pedi.12825>.
- Tamm, Christian, Richard Benzi, Irène Papageorgiou, Isabelle Tardy, and René Lerch. 1994. "Substrate Competition in Postischemic Myocardium: Effect of Substrate Availability during Reperfusion on Metabolic and Contractile Recovery in Isolated Rat Hearts." *Circulation Research* 75 (6): 1103–12.

[https://doi.org/10.1161/01.RES.75.6.1103.](https://doi.org/10.1161/01.RES.75.6.1103)

Taupenot, Laurent, Sushil K. Mahata, Manjula Mahata, Robert J. Parmer, and Daniel T. O'Connor. 2000. "Interaction of the Catecholamine Release-Inhibitory Peptide Catestatin (Human Chromogranin A352-372) with the Chromaffin Cell Surface and Torpedo Electrophax: Implications for Nicotinic Cholinergic Antagonism." *Regulatory Peptides* 95 (1–3): 9–17. [https://doi.org/10.1016/S0167-0115\(00\)00135-X](https://doi.org/10.1016/S0167-0115(00)00135-X).

Traczyk, Władysław Z. 2015. *Fizjologia Człowieka z Elementami Fizjologii Stosowanej i Klinicznej*. PZWL Wydawnictwo Lekarskie.

Valensi, Paul. 2021. "Autonomic Nervous System Activity Changes in Patients with Hypertension and Overweight: Role and Therapeutic Implications." *Cardiovascular Diabetology* 20 (1): 1–12. <https://doi.org/10.1186/s12933-021-01356-w>.

Wang, Dandan, Tao Liu, Shaobo Shi, Ran Li, Yingguang Shan, Yan Huang, Dan Hu, and Congxin Huang. 2016. "Chronic Administration of Catestatin Improves Autonomic Function and Exerts Cardioprotective Effects in Myocardial Infarction Rats." *Journal of Cardiovascular Pharmacology and Therapeutics* 21 (6): 526–35. <https://doi.org/10.1177/1074248416628676>.

Wang, Xinyu, Shunlin Xu, Ying Liang, Dan Zhu, Lin Mi, Guisong Wang, and Wei Gao. 2011. "Dramatic Changes in Catestatin Are Associated with Hemodynamics in Acute Myocardial Infarction." *Biomarkers* 16 (4): 372–77. <https://doi.org/10.3109/1354750X.2011.578260>.

World Heart Report. 2021. "Guideline for the Pharmacological Treatment of Hypertension in Adults." Geneva: World Health Organization. <https://apps.who.int/iris/bitstream/handle/10665/344424/9789240033986-eng.pdf>.

—. 2023. "World Heart Report 2023: Confronting the World's Number One Killer." Geneva, Switzerland. World Heart Federation. 2023. <https://world-heart-federation.org/wp-content/uploads/World-Heart-Report-2023.pdf>.

Xu, Weixian, Haiyi Yu, Hubing Wu, Siheng Li, Baoxia Chen, and Wei Gao. 2017. "Plasma Catestatin in Patients with Acute Coronary Syndrome." *Cardiology (Switzerland)* 136 (3): 164–69. <https://doi.org/10.1159/000448987>.

YAĞIZ, Burcu, Aysen AKALIN, Göknur YORULMAZ, Aslı Ceren MACUNLUOĞLU, and Onur YAĞIZ. 2022. "Can Non-Functional Adrenal Incidentaloma Be Ranked

among Cardiovascular Risk Factors?" *The European Research Journal* 8 (6): 747–54. <https://doi.org/10.18621/eurj.872835>.

Ying, Wei, Kechun Tang, Ennio Avolio, Jan M Schilling, Teresa Pasqua, Matthew A Liu, Hongqiang Cheng, et al. 2021. "The Immunosuppression of Macrophages Underlies the Cardioprotective Effects of Catestatin (CST)." *BioRxiv*, 2020.05.12.092254.

Yu, Wei, and Ji Dong Cheng. 2020. "Uric Acid and Cardiovascular Disease: An Update From Molecular Mechanism to Clinical Perspective." *Frontiers in Pharmacology* 11. <https://doi.org/10.3389/fphar.2020.582680>.

Zalewska, Ewa, Piotr Kmiec, Jakub Sobolewski, Andrzej Koprowski, and Krzysztof Sworczak. 2023. "Low Catestatin as a Risk Factor for Cardiovascular Disease – Assessment in Patients with Adrenal Incidentalomas." *Frontiers in Endocrinology* 14 (July): 1–12. <https://doi.org/https://doi.org/10.3389/fendo.2023.1198911>.

Zalewska, Ewa, Piotr Kmiec, and Krzysztof Sworczak. 2022. "Role of Catestatin in the Cardiovascular System and Metabolic Disorders." *Frontiers in Cardiovascular Medicine* 9. <https://doi.org/10.3389/fcvm.2022.909480>.

Zhao, Yilin, and Dan Zhu. 2016. "Potential Applications of Catestatin in Cardiovascular Diseases." *Biomarkers in Medicine* 10 (8): 877–88. <https://doi.org/10.2217/bmm-2016-0086>.

Zhu, Dan, Fangfang Wang, Haiyi Yu, Lin Mi, and Wei Gao. 2011. "Catestatin Is Useful in Detecting Patients with Stage B Heart Failure." *Biomarkers* 16 (8): 691–97. <https://doi.org/10.3109/1354750X.2011.629058>.

Zhu, Dan, Hong Xie, Xinyu Wang, Ying Liang, Haiyi Yu, and Wei Gao. 2015. "Correlation of Plasma Catestatin Level and the Prognosis of Patients with Acute Myocardial Infarction." *PLoS ONE* 10 (4): 1–14. <https://doi.org/10.1371/journal.pone.0122993>.

X. PUBLIKACJE WCHODZĄCE W SKŁAD ROZPRAWY DOKTORSKIEJ

XI. OŚWIADCZENIA WSPÓŁAUTORÓW



OPEN ACCESS

EDITED BY

Carmine Izzo,
University of Salerno, Italy

REVIEWED BY

Elke Marjolein Muntjewerff,
Uppsala University, Sweden
Yalin Wu,
University of Alabama at Birmingham,
United States

*CORRESPONDENCE

Piotr Kmiec
piotrkmiec@gumed.edu.pl

RECEIVED 02 April 2023

ACCEPTED 20 June 2023

PUBLISHED 14 July 2023

CITATION

Zalewska E, Kmiec P, Sobolewski J,
Koprowski A and Sworczak K (2023) Low
catestatin as a risk factor for cardiovascular
disease – assessment in patients with
adrenal incidentalomas.

Front. Endocrinol. 14:1198911.
doi: 10.3389/fendo.2023.1198911

COPYRIGHT

© 2023 Zalewska, Kmiec, Sobolewski,
Koprowski and Sworczak. This is an open-
access article distributed under the terms of
the [Creative Commons Attribution License
\(CC BY\)](#). The use, distribution or
reproduction in other forums is permitted,
provided the original author(s) and the
copyright owner(s) are credited and that
the original publication in this journal is
cited, in accordance with accepted
academic practice. No use, distribution or
reproduction is permitted which does not
comply with these terms.

Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas

Ewa Zalewska¹, Piotr Kmiec^{1*}, Jakub Sobolewski²,
Andrzej Koprowski² and Krzysztof Sworczak¹

¹Department of Endocrinology and Internal Medicine, Medical University of Gdańsk, Gdańsk, Poland

²First Department of Cardiology, Medical University of Gdańsk, Gdańsk, Poland

Background: Catestatin (Cts) is a peptide derived from proteolytic cleavage of chromogranin A, which exhibits cardioprotective and anti-inflammatory properties. Cts has been proposed as a potential biomarker for cardiovascular (CV) disease.

Objectives: examining Cts in patients with incidentally discovered adrenocortical adenomas (AI), and its associations with CV risk factors and blood pressure (BP).

Materials and methods: In this cross-sectional study, 64 AI patients without overt CV disease other than primary hypertension were recruited along with 24 age-, sex-, and body-mass-index (BMI)-matched controls with normal adrenal morphology. Laboratory, 24-h ambulatory BP monitoring, echocardiography, and common carotid artery sonography examinations were performed.

Results: Unadjusted Cts was higher in AI patients (median 6.5, interquartile range: 4.9–37 ng/ml) versus controls (4.5 (3.5 – 28)), $p=0.048$, however, the difference was insignificant after adjusting for confounding variables. Cts was lower in subjects with metabolic syndrome than in those without it (5.2 (3.9– 6.9) vs. 25.7 (5.8–115) ng/ml, $p<0.01$), and in men compared to women (4.9 (4–7.4) ng/ml vs. 7 (4.8–100), $p=0.015$). AI patients in the lower half of Cts levels compared to those in the upper had a higher prevalence of hypertension (OR 0.15, 95% CI: 0.041–0.5, $p<0.001$) and metabolic syndrome (OR 0.15, 95% CI 0.041–0.5, $p<0.001$). In AI patients Cts correlated positively with high-density lipoprotein cholesterol (Spearman's $r=0.31$), negatively with BMI ($r=-0.31$), and 10-year atherosclerotic CV disease risk ($r=-0.42$).

Conclusions: Our data indicate associations between CV risk factors and Cts. More clinical research is needed to apply serum Cts as a biomarker.

KEYWORDS

catestatin, adrenal incidentaloma (AI), cardiovascular disease(s), risk predictor, metabolic syndrome

1 Introduction

Risk factors for atherosclerotic cardiovascular disease (ASCVD) can be divided into nonmodifiable (e.g. age or sex) and modifiable (smoking, elevated blood pressure (BP), dyslipidemia, diabetes (DM), and obesity). Apart from established risk factors, new are sought (e.g. uric acid (UA) (1) and high-sensitivity C-reactive protein [hs-CRP] (2)) to help distinguish persons at higher risk of developing cardiovascular disease (CVD), who would benefit more from medical interventions such as low-density lipoprotein cholesterol (LDL-C) reduction (3).

The sympathetic nervous system plays a pivotal role in CVD development. Chromogranin A (CgA) is co-stored and co-released with catecholaminec from sympathetic neuronal vesicles and the adrenall medulla. One of CgA's proteolytic cleavage products is catestatin (Cts), a cardioprotective, anti-hypertensive, and anti-inflammatory peptide (4, 5). *In vitro*, Cts was shown to bind to nicotinic acetylcholine receptors, which inhibits membrane depolarization and blocks calcium influx, and, consequently, suppresses catecholamine release and activation of the sympathetic nervous system (6). Studies with animal models demonstrated Cts exerts anti-inflammatory effects, cardioprotection, and reduces obesity and insulin resistance (7–9). Clinical studies indicate Cts is involved in the course of hypertension (HT), coronary artery diseases (CAD), and heart failure (HF) (10–12). Adolescents with metabolic syndrome (MetS) had decreased Cts, which was postulated as a novel CVD risk factor (12–14).

In the current study we aimed at 1) determining Cts levels in patients with an incidentally-discovered adrenocortical adenoma/hyperplasia (AI) and without overt CVD other than HT, as well as 2) investigating associations between Cts and ASCVD risk modifiers, and asymptomatic HT-mediated organ damage (15). The presence of an AI *per se*, and particularly mild autonomous cortisol secretion (MACS) in its course, have been associated with metabolic disorders, elevated CV risk and mortality (16). So far, Cts has not been investigated in this patient population.

2 Subjects and methods

2.1 Study population

Study participants with an AI were recruited among 376 consecutive adult patients hospitalized in the Department of Endocrinology and Internal Medicine of the University Clinical Center of the Medical University of Gdańsk between November 2018 through February 2020 due to an adrenal lesion. We included 64 patients with radiological features of an adrenal adenoma/hyperplasia revealed by computed tomography (CT) or magnetic resonance (MR), who agreed to participate in the study, and met none of the following exclusion criteria: 1) age over 75 or under 40; 2) obesity grade III ($BMI >40 \text{ kg/m}^2$); 3) premenopausal period; 4) adrenal hormone excess other than MACS, i.e. cortisolemia between 50 and 138 nmol/l in the overnight 1-mg dexamethasone suppression test (DST) and no phenotypic features of Cushing's syndrome (17); 5) kidney disease with eGFR<60 ml/min/

1.73m^2 and/or proteinuria $>0.25 \text{ g/24h}$; 6) treatment with a mineralocorticoid receptor antagonist; 7) established and/or overt CVD other than primary HT, including: a) ASCVD (CAD, stroke, transient ischemic attack, peripheral artery disease), b) significant cardiac disease (e.g. pathological arrhythmia, severe valvular heart disease, cardiac tamponade, cardiomyopathy, congenital heart disease, HF), c) vascular diseases (among others venous thromboembolism and vasculitis); 8) active malignancy; 9) decompensated autoimmune disease or immune disease associated with CV and/or renal complications; 10) infectious diseases; 11) current or past addiction to alcohol and/or illicit drugs. Study participants were recruited based on anamnesis, physical examination, additional examinations available for review prior to enrollment and performed in the course of the study. Initially, 73 patients were included, however, three withdrew their consent to participate due to the COVID-19 pandemic, in four patients transthoracic echocardiography (TTE) revealed cardiac post-ischemic lesions, and two were diagnosed with primary aldosteronism.

Based on medical records of our hospital, which included examinations ordered in outpatient clinics and the emergency department, we identified 153 persons with normal adrenal morphology in a CT/MRI scan performed within five years preceding this study. There were 129 who met at least one of the above-listed exclusion criteria, declined participation or were unreachable, therefore, 24 subjects without an AI were enrolled as controls.

The research complied with the Declaration of Helsinki and was approved by the Independent Bioethics Committee for Research of our University. Informed consent for inclusion in the study was obtained in writing from each participant.

2.2 Study design

Both AI patients and controls underwent the following evaluation: 1) medical interview; 2) physical examination; 3) antecubital venous blood sampling for laboratory analyses; 4) resting 12-lead electrocardiography (ECG); 5) TTE; 6) common carotid artery (CCA) ultrasonography (USG) including CIMT determination, 7) 24-hour ambulatory blood pressure monitoring (ABPM).

Body-mass-index (BMI) was calculated by dividing body weight (W) in kg by the square of height (H) in meters. 2009 International Diabetes Federation criteria were used to diagnose MetS (18). Subjects with HT received hypotensive medications at the time of enrollment or were diagnosed by ABPM based on mean systolic and diastolic BP (SBP and DBP, respectively) of at least 135/85 mmHg for daytime, 120/75 mmHg for nighttime, and/or 130/80 mmHg for the 24-h period (15). Atherogenic dyslipidemia was defined as triglycerides (TGL) $\geq 150 \text{ mg/dL}$ and serum high-density lipoprotein cholesterol (HDL-C) $<40 \text{ mg/dL}$ for men and $<45 \text{ mg/dL}$ for women.

In all study participants, 10-year ACSVD risk was estimated using the 2018 calculator provided online by the American Heart Association and the American College of Cardiology based on Framingham Heart Study (FHS-ASCVD Risk) (19, 20). The calculator estimates 10-year risk of developing ASCVD including coronary death, myocardial infarction, coronary insufficiency, angina, ischemic stroke,

hemorrhagic stroke, transient ischemic attack, peripheral artery disease, and HF for individuals aged 30 to 74 and without CVD at baseline based on the following predictors: age, type 2 DM (DMt2), smoking, treated and untreated SBP, total cholesterol (TC), HDL-C, and LDL-C.

For nondiabetic subjects, 10-year CVD risk was also calculated using Systematic Coronary Risk Estimation 2 (SCORE2) for subjects aged 40-69 and SCORE2-Older People (SCORE2-OP) for those aged 70-75 for high CV risk countries, which include Poland (21). Predictors used in SCORE2-OP are: age, sex, smoking, SBP, and non-HDL-C. We are aware SCORE2-OP was developed to estimate risk in treatment-naïve persons, and that a significant portion of our subjects received lipid- and BP-lowering therapy. Nevertheless, we concluded applying this estimation tool along with FHS-ASCVD Risk calculation is of value.

2.3 Laboratory examinations

Blood was drawn between 8 and 10 a.m. after a fasting period of at least 8 hours from an antecubital vein, and used for regular examinations in the laboratory of our hospital apart from samples preserved for the determination of plasma Cts in all subjects, serum aldosterone and plasma direct renin concentration (DRC) in controls. These were centrifuged at 2,000 rpm for 20 minutes at 4 degrees C, aliquoted and stored at -80 degrees C until analysis.

Samples were analyzed in Central Clinical Laboratory in Gdańsk using standard laboratory methods (with a Siemens IMMULITE 1000 Immunoassay System for most biochemical tests, and an Abbott Architect analyzer, which applies the spectrophotometric method). Serum Cts was determined by an enzyme-linked immunosorbent assay (ELISA) by using a commercially-available diagnostic kit (SunRedBio, catalogue no: 201-12-8276; sensitivity: 0.268 ng/mL; assay range: 0.3-90 ng/mL). Cts concentrations above 90 ng/ml ($n=15$) were extrapolated based on ELISA standard curve.

Serum Cts, creatinine, sodium, potassium, aldosterone, renin, lipid profile (TC, HDL-C, LDL-C, and TGL), UA, hs-CRP, 24-h urinary protein and albumin excretion were determined both in AI patients and controls. Morning serum cortisol, dehydroepiandrosterone sulphate (DHEA-S), overnight 1 mg-DST cortisol and 24-h urinary cortisol were determined in AI patients. In most ($n=50$) AI patients, 24-h urinary meta- and normetanephrine excretion was determined, in others it had been performed prior to hospitalization. Screening for primary hyperaldosteronism based on aldosterone-to-renin ratio (ADRR) was performed without modifying antihypertensive medications in both AI patients and controls; there were no study participants with both HT and an ADRR above 2 ng/dL: μ IU/mL.

2.4 Ambulatory blood pressure monitoring

24h ABPM was conducted using a Spacelabs Ontrak 90227 monitor on the non-dominant arm. During the day BP was recorded every 20 minutes, while during nighttime rest every 30 minutes. ABPM was repeated or not considered in the analysis if more than 30% of measurements were invalid. Normal results were adopted according to the European Society of Cardiology/European

Society of Hypertension 2018 guideline: <130/80 mmHg for the 24-h period, <135/85 mmHg for daytime, and <120/70 mmHg for nighttime (22). Patients were classified as ‘non-dippers’ if their mean diurnal SBP and DBP were not at least 10% higher than nocturnal (22).

2.5 Transthoracic echocardiography

All measurements were performed in accordance with the recommendations endorsed by the American Society of Echocardiography and the European Association of Cardiovascular Imaging (23). Three on-site cardiology consultants with an expertise in ultrasonography performed TTE using the GE Vivid E9/E95 ultrasound system.

Measurements included left-ventricular (LV) internal dimension in diastole (LVIDd) and systole (LVIDs), LV ejection fraction (LVEF) according to modified Simpson’s rule (24), posterior LV wall thickness (LVPWd), and interventricular septal thickness (IVSd). LV mass (LVM) was calculated with the cube formula: $LVM(g)=0.8 \times 1.04 \times [(LVEDd+IVSd+LVPWd)^3 - LVEDd^3] + 0.6$. LVM was indexed to body surface area (BSA) calculated using the DuBois formula ($BSA=0.007184 \times H^{0.725} \times W^{0.425}$): LVM index (LVMI)=LVM/BSA. Relative wall thickness (RWT) was calculated with the formula: $RWT=(2 \times LVPWd)/LVEDd$. Left ventricular hypertrophy (LVH) was defined as $LVMI > 95 \text{ g/m}^2$ for females and $> 115 \text{ g/m}^2$ for males. RWT was used to further classify LVH as either concentric ($RWT > 0.42$) or eccentric ($RWT \leq 0.42$).

Disk summation technique from apical four and two-chamber views was used to determine left atrial volume (LAV), which was indexed to BSA: LAV index (LAVI) (ml/m^2) = LAV/BSA (15). Apical four-chamber view was used to record peak blood flow velocity from LV relaxation in early diastole (E) and peak velocity flow in late diastole (A). Since LVEF was normal in all study participants, four criteria were applied to assess diastolic function: (1) $LAVI \geq 34 \text{ ml/m}^2$, (2) tricuspid regurgitation velocity (TR) $\geq 2.8 \text{ m/s}$, (3) ratio of E to average early mitral annular velocity (e') ≥ 14 , (4) septal $e' < 7 \text{ cm/s}$ or lateral $e' < 10 \text{ cm/s}$. Indeterminate diastolic function was stated if two criteria were met, and dysfunction if three or four (15).

2.6 Common carotid artery USG

Maximum carotid intima-media thickness (CIMT) measurements were recorded using echo-tracking technology on the distal wall of the right carotid artery, 1 to 3 cm below the carotid artery bifurcation. The presence of atherosclerotic plaques (ASP) defined as a CIMT $\geq 1.5 \text{ mm}$, or by a focal increase in thickness of 0.5 mm or 50% of the surrounding CIMT value was also recorded.

2.7 Statistical analysis

Data were analyzed using R-studio. Discrete variables were presented as number (n) or n (percentage). Continuous

quantitative data with a normal distribution were presented as mean \pm standard deviation (SD), and in the case of a non-normal distribution as median (interquartile range, IQR). We used the Shapiro-Wilk test to determine if a data set was well-modeled by a normal distribution. To compare differences between two independent groups Welch's t-test was used when variables were normally distributed or the Mann-Whitney U test in the case of non-normal distribution. One-way ANOVA and Tukey's honestly significant difference (HSD) tests were used to compare three or more independent groups. Simple (bivariate) correlations were computed with the non-parametric Spearman rank-order method (correlation coefficient r is given). Associations between dichotomous categorical variables were examined with Fisher's exact test, and Benjamini-Hochberg Method was applied to correct for multiple testing.

Multiple regression models were applied to adjust for differences in Cts concentrations depending on potential confounding variables including gender, age, BMI, smoking status, comorbidities (HT, DMt2, MetS), and medications (ACEI/ARB, CCB, BB, diuretics, statins, and PPIs). An exhaustive search method was used to select factors that had the strongest relationship with Cts, i.e.: 1) gender, presence of 2) MetS, 3) HT, therapy with 4) statins, and 5) PPIs. The final multivariate model had a R-squared of 0.1831. Out of five variables included in the model only the presence of MetS ($\beta=-30$, $p=0.005$) was significantly and negatively associated with Cts. For dichotomous dependent variables (Cts halves, HT, DMt2, and MetS) binary logistic regression was used to adjust for gender, age, and BMI. Significance was set at 0.05.

3 Results

3.1 Comparison of examined parameters between AI patients and controls

To assess whether the presence of an AI affected Cts levels, verification of matching between AI patients and controls was undertaken. These groups did not differ in regard to age, sex, BMI, smoking status, and comorbidities (incidence of HT, DM t.2, ASP, and dyslipidemia), see [Table 1](#). Concerning subjects with HT, the number of patients on mono-, dual- and triple-drug therapy (including betablockers) was also comparable ([Supplementary Table 1](#)).

Among AI patients, there were 14 with MACS and 31 classified as NFAI; analyses for AI patients and these two subgroups were performed separately.

Ten-year FHS-ASCVD Risk was comparable between patients with an AI/NFAI/MACS and controls, while in nondiabetic subjects, SCORE2/-OP was significantly higher in patients with MACS than in controls and patients with NFAI: 14% (11-18) vs. 8% (4.5-14), $p = 0.021$, and 8% (6-12), $p = 0.005$, respectively ([Table 1](#)). Still, this CVD risk index was comparable between controls and all AI patients ($p=0.31$), which illustrates effective matching between these groups.

Cts distribution was bimodal both in AI patients and controls. Unadjusted Cts was slightly higher in AI patients: 6.45 (4.9-37) vs.

4.5 (3.5-28) ng/ml, $p=0.047$ ([Figure 1](#)). However, after adjusting for potential confounding variables (gender, age, and BMI), solely BMI and male gender were significantly (negatively) associated with Cts ($\beta=-28.3$, $p=0.01$ and $\beta=-2.3$, $p=0.04$, respectively) but not the presence of an AI ($\beta=-8.1$, $p=0.44$).

Lipid profile, hs-CRP, as well as UA were comparable between controls and AI patients, be it with a NFAI or MACS. Proteinuria and albuminuria were normal in all study participants (respectively below 150 and 30 mg/24h). ABPM parameters (SBP, DBP, and pulse rate, PR) were comparable between AI patients and controls ([Table 1](#) and [Supplementary Table 1](#)).

Concerning TTE, there were significant differences in IVSd, LVPWd, and LVMI between AI patients and controls (respectively 11 (10-12) vs. 10 (9-10) mm, $p=0.003$; 10 (9-11) vs. 9 (8-9.5) mm, $p=0.007$; 86.4 ± 119.2 vs. 84.7 ± 18.5 g/m², $p=0.001$). Moreover, LVH was more prevalent in MACS patients than controls (42.9% vs. 4.4%, $p=0.007$) and NFAI patients (42.9% vs. 14%, $p=0.028$).

Maximum CIMT was higher in patients with an AI, be it with a NFAI or MACS, than in controls: 1 (0.9-1.1) vs. 0.8 (0.8-0.9) mm, $p<0.01$. However, there were no differences in maximum CIMT between patients with a NFAI and MACS ([Table 1](#)). A trend toward a higher prevalence of an ASP in AI, NFAI, and MACS patients (29.7%, 32%, 21.43%, respectively) than controls (9.5%) could be observed ($p=0.12$) ([Table 1](#)).

3.2 Catestatin in clinically-specified patient groups

Upon comparing Cts levels between controls with normal adrenal morphology and AI patients, peptide's levels were tested in different patient groups. Cts was higher in women than in men: 7 (4.8-100) vs. 4.9 (4-7.4) ng/ml, $p=0.015$, and the difference between sexes was significant in both AI patients (7.3 (5.5-103) vs. 6 (4.26-7.6) ng/ml, $p=0.03$) and controls (5.1 (3.8 - 62.6) vs. 2.8 (1.7 - 3.5) ng/ml, $p = 0.043$), see [Figure 2](#).

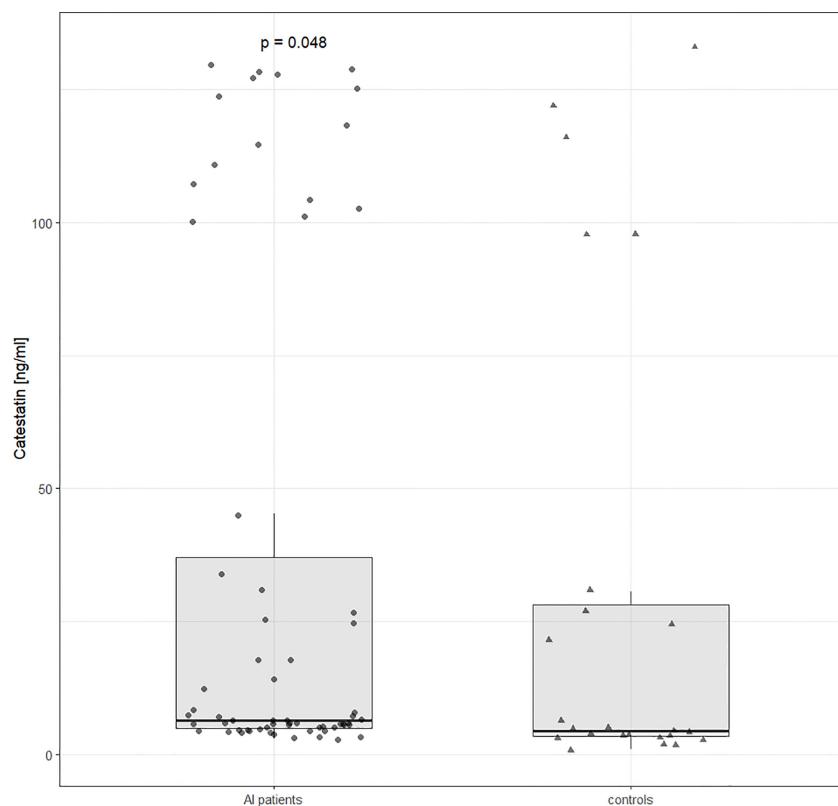
Further, in AI patients and controls analyzed together Cts was lower in hyper- versus normotensive subjects: 5.6 (4-7.1) vs. 15.8 (5.2-103) ng/ml, $p=0.003$, which was also found for AI patients alone: 5.6 (4.36-6.82) vs. 21.7 (6.85-107), $p < 0.001$ ([Figure 3](#)). Cts was also significantly lower in subjects with MetS than in those without it: 25.7 (5.8-115) vs. 5.2 (3.9- 6.9) ng/ml, $p<0.01$ ([Figure 4](#)), regardless of potential confounders (gender, age, BMI, presence of an AI and/or HT, statin and PPI use). We confirmed these differences (normo- versus hypertensive subjects as well as those without and with MetS) in women but not men (probably due to their low number). Cts in hypertensive AI females was lower than in normotensive ones: 5.6 (4.7 – 11.6) vs. 45.2 (8.2 – 118) ng/ml, $p <0.01$, and also lower in those with MetS than without it, both among AI patients: 5.6 (4.8-6.9) vs. 34.3 (7.7 - 121) ng/ml, $p= <0.01$ and controls: 3.8 (3.7-5.1) vs. 61.2 (8.4-112) ng/ml, $p = 0.025$ ([Supplementary Figures 1, 2](#)).

There were no differences in Cts between obese and non-obese subjects, smokers and non-smokers, or, among subjects with HT, 'dippers' and 'non-dippers'.

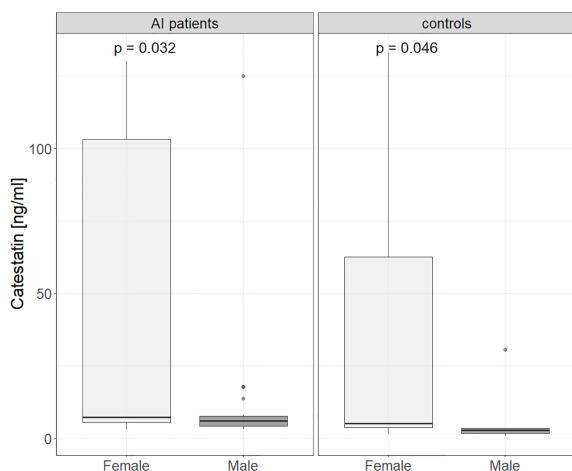
TABLE 1 Clinical, laboratory, ABPM, echocardiographic, and CCA sonography parameters in AI patients and controls.

	Controls	AI patients			p	adjusted p		
		all	NFAI	MACS		Cont. vs. AI	Cont. vs. NFAI	Cont. vs. MACS
n	24	64	50	14	–	–	–	–
F:M ratio	19:5	45:19	35:15	10:4	0.592	1	1	1
Age [years]	62.2 ± 7.4	60.9 ± 8.8	60.1 ± 8.8	63.7 ± 9.3	0.487	0.575	0.854	0.334
BMI [kg/m ²]	27.9 ± 4.6	28.6 ± 4.1	28.8 ± 4.2	28.3 ± 3.9	0.48	0.685	0.968	0.911
Obesity [n (%)]	8 (33.33%)	24 (37.5%)	20 (40%)	4 (28.6%)	0.807	0.928	1	0.928
HT [n (%)]	10 (41.7%)	36 (56.3%)	27 (54%)	9 (64.3%)	0.327	0.555	0.555	0.555
DMt2 [n (%)]	1 (4.17%)	12 (18.8%)	11 (22%)	1 (7.1%)	0.103	0.266	1	0.411
MetS [n (%)]	11 (45.8%)	36 (56.3%)	27 (54%)	9 (64.3%)	0.527	0.621	0.621	0.621
Smokers [n (%)]	24 (33.3%)	28 (43.8%)	20 (40%)	8 (57.1%)	0.521	0.619	0.543	0.543
SCORE2/-OP [%] *	8 (4.5-14)	9 (7-13)	8 (6-12)	14 (11-18)	0.31	0.978	0.021	0.005
FHS-ASCVD score [%]	5.9 (2.8-12)	10.1 (4.8-16.4)	9.3 (4.6-16.4)	12.5 (8.4-15.6)	0.085	0.338	0.253	0.813
Catestatin [ng/ml]	4.5 (3.5-28)	6.5 (4.9-37)	7.2 (5-101)	6.1 (5-7.8)	0.048	0.71	0.7	0.274
HDL-C [mg/dl]	58.7 ± 12.1	54.0 ± 14.7	52.7 ± 14	58.7 ± 17	0.13	0.196	1	0.33
LDL-C [mg/dl]	120 ± 36.6	129 ± 48.2	130 ± 49.1	129 ± 46.8	0.32	0.663	0.828	0.998
TC [mg/dl]	202 ± 42.4	211 ± 53.1	210 ± 53.4	213 ± 53.8	0.45	0.813	0.803	0.978
TGL [<150 mg/dl]	106 (96.8-126)	130 (87-162)	130 (88.5-160)	129 (86.2-162)	0.25	0.414	0.965	0.727
UA [2.5-7 mg/dl]	5.3 (4.8-5.8)	5.1 (4.2-6.1)	5.1 (4.4 -6.1)	4.8 (4.2-5.8)	0.97	0.87	0.983	0.817
hs-CRP [<5 mg/l]	1.4 (1.1-2.5)	1.2 (0.7 - 1.7)	1.2 (0.7-1.6)	1.3 (0.6-2)	0.14	0.811	0.443	0.155
DST cortisol [<50 nmol/L]	–	26.9 ± 34	11.3 ± 17	79.1 ± 22.3	–	–	–	<0.001
24h SBP [mmHg]	118 ± 8.4	121 ± 9.7	120 ± 9.3	118 ± 7.7	0.5	0.62	0.982	0.612
24h DBP [mmHg]	70.6 ± 5.9	71.4 ± 7.8	71.7 ± 8.1	70.3 ± 6.5	0.61	0.818	0.994	0.82
Non-dipper status [n (%)]†	6 (40%)	9 (28.1%)	9 (37.5%)	0	0.442	0.61	0.447	0.46
IVSd [mm]	10 (9-10)	11 (10-12)	11 (10-12)	12 (10.2-12)	0.003	0.045	0.006	0.295
LVIDd [mm]	46.1 ± 4.1	44.8 ± 4.5	45.2 ± 4.7	43.5 ± 3.7	0.193	0.651	0.183	0.427
LVIDs [mm]	29.9 ± 3.3	27.3 ± 3.1	27.3 ± 3.2	27.2 ± 3	0.002	0.005	0.036	0.991
LVPWd [mm]	9 (8-9.5)	10 (9-11)	10 (9-11)	11 (10-11.8)	0.007	0.08	0.009	0.268
LVM [g]	149 ± 27.2	165 ± 41.8	164 ± 42	169 ± 42.6	0.047	0.307	0.289	0.887
LVMI [g/m ²]	86.4 ± 19.2	84.7 ± 18.5	92.7 ± 21	77.9 ± 8.7	0.006	0.037	0.023	0.21
LVH [n (%)] #	1 (4.4%)	13 (20.3%)	7 (14%)	6 (42.9%)	0.101	0.421	0.007	0.028
LAVI [ml/m ²]	24.6 (15.1-31.9)	24.9 (15.9-31.6)	23.8 (13.1-31.8)	25.5 (20.5-30)	0.29	0.639	0.498	0.864
CIMT max [mm]	0.8 (0.7 -0.8)	1 (0.9 - 1.1)	1 (0.9 - 1.1)	0.9 (0.9 - 1)	< 0.01	<0.01	0.007	0.996
ASP [n (%)]	2 (9.5%)	19 (29.7%)	16 (32%)	3 (21.4%)	0.117	0.215	0.526	0.526

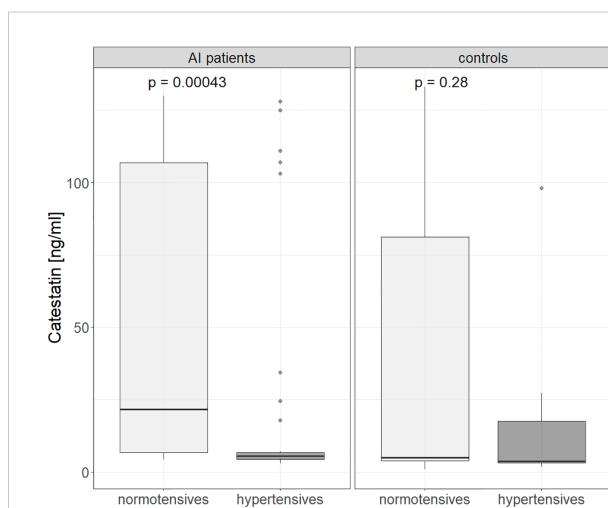
Data are presented as number, n, (percentage, %), mean ± standard deviation or median (interquartile range) depending on distribution; p values were adjusted for multiple comparisons with Benjamini-Hochberg adjustment (for qualitative variables) and TukeyHSD test (for quantitative variables); bold font denotes significant (<0.05) p values; *only nondiabetic patients were included in SCORE2/-OP risk estimation (n= 23, 52, 39, and 13, respectively for Cont., AI, NFAI and MACS patients); † LVH was defined as values of LVMI exceeding 95 or 115 g/m² in females and males respectively; † dipper status was considered only in patients with HT. ASP, atherosclerotic plaques; BMI, body mass index; CIMT, carotid intima media thickness; con., controls; DBP, diastolic blood pressure; DMt2, diabetes mellitus t2; DST, dexamethasone suppression test; FHS-ASCVD Risk – 10-year atherosclerotic cardiovascular disease risk calculated based on the Framingham Heart Study; HDL-C, high density lipoprotein cholesterol; HT, hypertension; hs-CRP, high sensitivity C-reactive protein; IVSd, interventricular septal end diastole; LAVI, left atrial volume index; LDL-C, low density lipoprotein cholesterol; LVH, left ventricular hypertrophy; LVIDd, left ventricular internal diameter end-diastole; LVIDs, left ventricular internal diameter end-systole; LVM, left ventricular mass; LVMI, LVM index; LVPWd, left ventricular posterior wall end diastole; MetS, metabolic syndrome; SBP, systolic blood pressure; SCORE2/-OP, Systematic Coronary Risk Estimation 2/Older People; TC, total cholesterol; TGL, triglycerides, UA, uric acid.

**FIGURE 1**

Catestatin distribution in controls and AI patients. Boxplot and data distribution with dots (AI patients) and triangles (controls) indicating individual datapoints. Unadjusted p value was determined using the Mann Whitney U test. AI, adrenal incidentaloma.

**FIGURE 2**

Catestatin in male and female controls and AI patients. Boxplot chart. P -value was determined using the Mann-Whitney U test. AI, adrenal incidentaloma.

**FIGURE 3**

Catestatin in normotensive and hypertensive in controls and AI patients. Boxplot chart. P -values were determined using the Mann-Whitney U test. AI, adrenal incidentaloma.

3.3 Correlations between catestatin and laboratory, TTE, and CCA USG parameters

To further investigate associations between Cts and CVD risk, correlations were tested between peptide's levels and other

parameters. In AI patients, weak correlations were found between Cts and: BMI ($r=-0.31$) (Figure 5A), FHS-ASCVD Risk ($r=-0.42$) (Figure 5B), and HDL-C ($r=0.32$) regardless of statin therapy (Figure 5C). Interestingly, among participants without it, there were also positive correlations between Cts and: TC and LDL-C

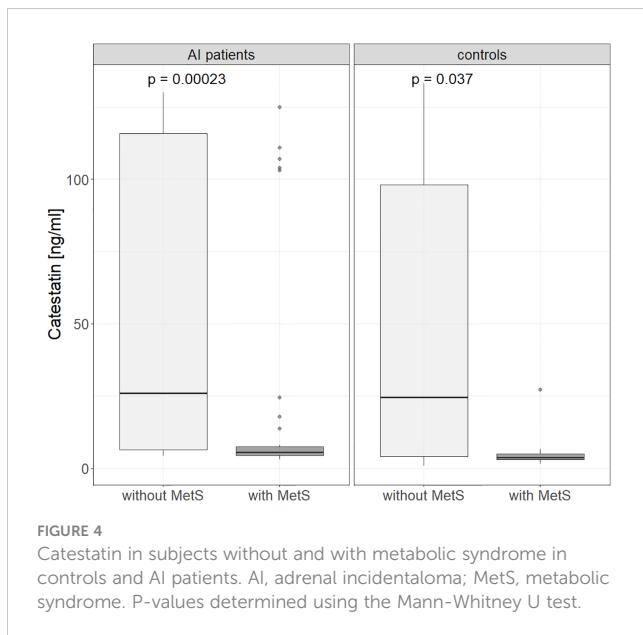


FIGURE 4

Catestatin in subjects without and with metabolic syndrome in controls and AI patients. AI, adrenal incidentaloma; MetS, metabolic syndrome. P-values determined using the Mann-Whitney U test.

($r=0.36$ for both) (Table 2 and Supplementary Figures 3, 4). In AI patients and controls analyzed as a whole a negative correlation between Cts and UA was also observed ($r=-0.27$, $p=0.01$), while for each group analyzed separately, significance has not been reached probably due to sample size.

Analyses in subjects of each sex revealed Cts correlated with HDL ($r=0.31$, $p=0.014$), BMI ($r=-0.29$, $p=0.019$), and UA ($r=-0.27$, $p=0.031$) in women, but not in men (respectively $r=0.13$, $p=0.53$; $r=-0.06$, $p=0.78$; $r=-0.12$, $p=0.56$). A negative correlation between Cts and FHS-ASCVD Risk was also recorded in women with an AI ($r=-0.3$, $p=0.049$) but not in female controls ($r=0.025$, $p=0.92$), nor men with an AI ($n=19$, $r=-0.37$, $p=0.12$). Correlations for male controls were not tested due to a low number of these subjects ($n=5$).

Concerning hormonal tests, only a weak correlation between Cts and DRC was observed, but not with aldosterone, nor ADRR (Supplementary Table 2). Cts did not correlate with ABPM, CCA USG, nor TTE parameters (LVPWd, IVSd, LVMI, LAVI) (Table 2 and Supplementary Table 2).

3.4 Clinical, laboratory, and TTE parameters according to catestatin categories

Further analyses were performed among AI patients based on Cts categories. First, adjustment for gender, age and BMI in binary logistic regression analysis revealed AI patients in the lower half of Cts concentrations (median 6.5, IQR 4.9-37 ng/ml) compared to those in the upper had a higher prevalence of HT (OR 0.17, CI 0.05-5.37, $p=0.003$), and MetS (OR 0.21, CI 0.06-7.51, $p=0.018$). Moreover, BMI, 24-SBP, and FHS-ASCVD Risk were also higher in the former (respectively 30.1 ± 4 vs. $27.2 \pm 3.6 \text{ kg/m}^2$, $p=0.004$; 123 ± 7.4 vs. $117 \pm 9.5 \text{ mmHg}$, $p=0.022$; $13.2(8.9-19.2)$ vs. $6.3(4.2-10.8)$, $p=0.002$), as summarized in Table 3.

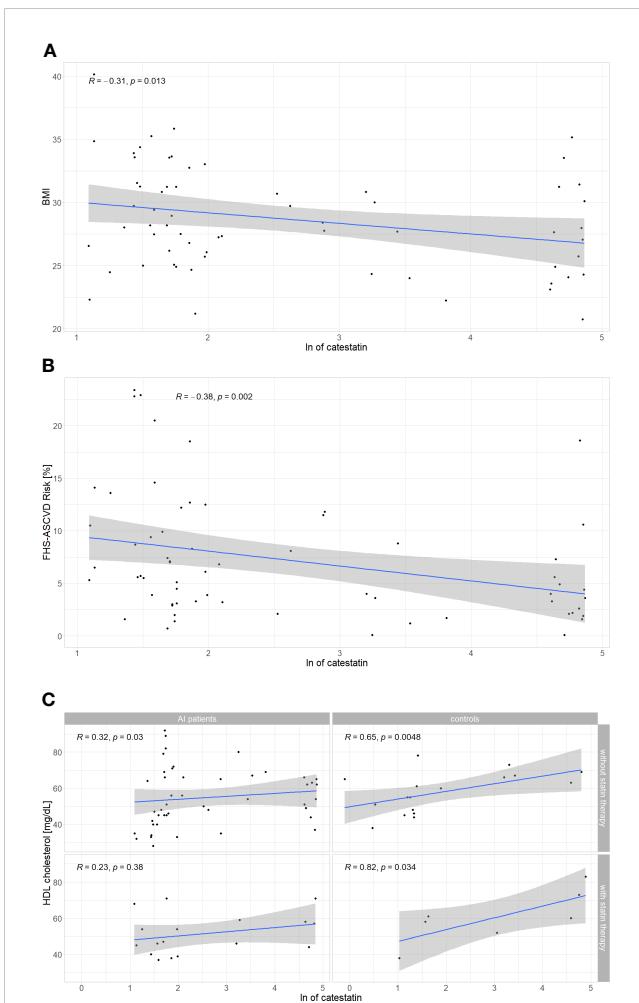


FIGURE 5

Correlations between catestatin and: (A) BMI in AI patients; (B) FHS-ASCVD Risk in AI patients; (C) HDL-C in AI patients and controls. Correlations were computed by the Spearman rank-order method. For HDL-C, correlations were tested separately in subjects with and without statin therapy. AI, adrenal incidentaloma, BMI, body mass index, FHS-ASCVD Risk - 10-year atherosclerotic cardiovascular disease risk calculated based on the Framingham Heart Study; HDL-C, high density lipoprotein cholesterol, ln, natural logarithm.

Second, based on Cts distribution, we divided AI participants into four subgroups, i.e. with: ‘very low’ (Cts <4.9 ng/ml, $n=17$), ‘low’ (≥ 4.9 and <6.5 ng/ml, $n=15$), ‘intermediate’ (≥ 6.5 and ≤ 45.2 ng/ml, $n=17$), and ‘high’ (≥ 100 ng/ml, $n=15$) Cts levels (Table 3). The first two comprised subjects from two lower quarters, while the ‘high Cts’ subgroup corresponded to almost all patients in the fourth quarter (15 instead of 16 patients were included since there were none in the 45.2 - 100 ng/ml range, see Figure 1).

These four Cts subgroups differed significantly in male-to-female ratio, prevalence of HT and MetS, mean/median BMI, HDL-C, 24h SBP, and FHS-ASCVD Risk (Table 3). Post hoc analysis revealed male gender was more prevalent in the ‘very low’ versus ‘high’ Cts subgroup (53% vs. 6.7%), while HT and MetS in the ‘very low’ versus ‘intermediate’ (82.4% vs. 35.3%, $p=0.04$ for both) and ‘high’ Cts subgroups (82.4% vs. 33.3%, $p=0.04$ for both). HDL-C was lower in the ‘very low’ than in the three remaining Cts subgroups (42.9 ± 42.9 vs. 61.6 ± 17.8 , $56.7 \pm$

TABLE 2 Correlations between catestatin and examined parameters.

Correlation between Cts and	Control group (n = 24)		AI patients (n = 64)		Both groups (n = 88)	
	r	p	r	p	r	p
Age	0.246	0.247	-0.142	0.262	-0.016	0.7
BMI	-0.293	0.164	-0.308	0.013	-0.27	0.009
HDL-C	0.704	< 0.001	0.306	0.014	0.344	0.001
HDL-C *	0.649	0.005	0.317	0.03	0.317	0.011
LDL-C	0.102	0.634	0.15	0.236	0.153	0.15
LDL-C *	0.573	0.016	0.361	0.003	0.361	0.003
TC	0.215	0.313	0.118	0.353	0.16	0.137
TC *	0.568	0.017	0.295	0.044	0.364	0.003
TGL	-0.216	0.145	-0.19	0.131	-0.143	0.183
TGL *	-0.085	0.746	0.215	0.313	-0.142	0.263
UA \$	-0.37	0.07	-0.209	0.1	-0.27	0.01
hs-CRP	0.191	0.407	0.005	0.97	-0.045	0.68
LVMI	0.149	0.488	-0.009	0.942	0.05	0.645
LAVI	0.13	0.545	-0.202	0.109	-0.121	0.263
Maximum CIMT	-0.09	0.697	-0.136	0.286	-0.03	0.774
SCORE2/-OP #	0.127	0.563	-0.064	0.652	0.05	0.651
FHS-ASCVD Risk [%]	-0.237	0.265	-0.42	< 0.001	-0.24	0.022

Correlations were computed by the Spearman rank-order method; bold font denotes statistically significant correlations; *denotes correlations in participants without statin therapy: n=17 and 47, respectively for controls and AI patients; \$ patients with and without medications that could lower uric acid (allopurinol, n = 1) were analyzed separately and the results were the same; # denotes correlations in nondiabetics only: n= 23 and 52, respectively for controls and AI patients; BMI, body mass index; CIMT, carotid intima media thickness; FHS-CVD, 10-year atherosclerotic cardiovascular disease risk calculated based on the Framingham Heart Study; HDL-C, high density lipoprotein cholesterol; hs-CRP, high sensitivity C-reactive protein; LAVI, left atrial volume index; LDL-C, low density lipoprotein cholesterol; LVMI, left ventricular mass index; r, correlation coefficient; SCORE2/-OP, Systematic Coronary Risk Estimation 2/-Older People; TC, total cholesterol; TGL, triglycerides; UA, uric acid.

13.7, and 55.8 ± 9.5 mg/dL, adjusted p=0.001, 0.01, and 0.03, respectively). What is more, 24h SBP in the ‘low’ Cts subgroup was higher than in the ‘intermediate’ (124.7 ± 6.7 vs. 114.5 ± 8.5 mmHg, adj. p=0.008).

Ten-year FHS-ASCVD Risk in the ‘very low’ Cts subgroup was higher than in the ‘intermediate’ and ‘high’: 14.3% (19.2-24.5) vs. 7.1% (4.7-14.2) and 5.6% (3.8-9.8), respective adjusted p=0.014 and 0.005, in line with differences in gender proportions, prevalence of metabolic disorders and HT between the subgroups.

Fisher’s exact test revealed differences in LVH prevalence between four Cts subgroups, yet, without significance in pairwise comparisons with correction for multiple testing (Holm-Bonferroni method). No other significant differences were recorded between Cts halves and subgroups in TTE and CCA USG parameters (Table 3). Finally, clinical, laboratory and TTE parameters were also analyzed in the same Cts subgroups for females only (Supplementary Table 3). Significant differences were recorded for: HT and MetS prevalence, HDL-C and hs-CRP concentrations, mean 24h DBP, and FHS-ASCVD Risk. Calculations for male AI subjects were not performed due to their low number.

4 Discussion

Clinical research on Cts is scarce, even though it deserves attention due to its protective effects on the CV system demonstrated *in vitro* and *in vivo*. To our knowledge, our study is the first to examine Cts in patients with an AI, and to show lower Cts in adult patients with MetS than those without it, as well as correlations between Cts and FHS ASCVD risk index.

A thorough assessment was undertaken to investigate associations between Cts and CV risk factors. It must be highlighted that Cts changes dynamically in response to sympathetic nervous system activation in a negative feedback mechanism (25). Also, multiple diseases and drugs lead to CgA secretion, which may affect the concentration of its derivatives, including Cts. For these reasons, we excluded patients with established CVD, stage 3-5 chronic kidney disease, cancer, etc., and controlled the use of PPIs (10, 12, 26, 27). Limitations of our study include a small, heterogeneous patient sample, lack of CgA determination (Cts : CgA ratios may have provided further insights) and hormonal work-up in controls.

Since CgA is not expressed in the adrenocortical adenoma tissue, Cts levels are unlikely to differ between subjects with and

TABLE 3 Clinical, laboratory, ABPM, echocardiographic, and CCA sonography parameters in AI patients according to catestatin category.

Cts half [ng/ml]	Lower (Cts < 6.5)		Upper (Cts ≥ 6.5)		p subgroups	p halves
Cts subgroup [ng/ml]	Very low (< 5)	Low (5 ≤ Cts < 6.5)	Intermediate (6.5 ≤ Cts ≤ 45.2)	High (Cts ≥ 100)		
n	17	15	17	15	-	-
F:M ratio	8:9	12:3	11:6	14:1 *	0.027	0.274
Age [years]	61.9 ± 9.4	61.7 ± 5.9	60.7 ± 11.5	59.2 ± 7.2	0.827	0.412
BMI [kg/m ²]	30.4 ± 4.6	29.9 ± 3.4	27.1 ± 3.2	27.4 ± 4.2	0.034	0.004
Obesity [n (%)]	8 (47.1%)	8 (53.3%)	3 (17.7%)	5 (33.3%)	0.158	0.07
Smokers [n (%)]	7 (41.2%)	9 (40%)	5 (29.4%)	7 (46.7%)	0.378	0.45
PPI therapy [n (%)]	4 (23.5%)	4 (26.7%)	2 (11.8%)	3 (20%)	0.793	0.536
HT [n (%)]	14 (82.4%)	11 (73.3%)	6 (35.3%) *	5 (33.3%) *	0.005	< 0.001
>1 hypotensive drug [n(%)] †	7 (50%)	7 (63.6%)	3 (50%)	1 (20%)	0.495	0.470
DMt2 [n (%)]	6 (35.3%)	4 (26.7%)	1 (5.9%)	1 (6.7%)	0.09	0.022
MetS [n (%)]	14 (82.4%)	11 (73.3%)	6 (35.3%) *	5 (33.3%) *	0.005	< 0.001
Statin use [n (%)]	6 (35.3%)	3 (20%)	4 (23.5%)	4 (26.7%)	0.837	1
HDL-C [mg/dL]	42.9 ± 10.8	61.6 ± 17.8 *	56.7 ± 13.7 *	55.8 ± 9.5 *	0.001	0.076
LDL-C [mg/dL]	124 ± 49.4	121 ± 41.6	127 ± 36.3	146 ± 63.6	0.491	0.27
TC [mg/dL]	202 ± 52.6	206 ± 51	211 ± 41	225 ± 68.4	0.677	0.31
TGL [mg/dL]	143 (98 - 198)	133 (118 - 141)	121 (87 - 168)	105 (87.5 - 142)	0.159	0.36
Uric acid [mg/dL]	5.8 (4.9 - 6.8)	4.8 (4.1 - 5.8)	5.1 (4.2 - 6.1)	4.9 (4.3 - 5.8)	0.112	0.36
MACS [n (%)]	4 (23.5%)	4 (26.7%)	5 (29.4%)	1 (6.7%)	0.417	0.763
Hs-CRP [mg/L]	1.2 (0.8 - 3.9)	1.5 (1 - 2.1)	0.8 (0.6 - 1.5)	1.2 (0.8 - 2)	0.243	0.16
24h SBP [mmHg]	120.8 ± 7.8	124.7 ± 6.7	114.5 ± 8.5 #	119.2 ± 10.5	0.01	0.009
24h DBP [mmHg]	71.1 ± 9.6	74.9 ± 5.9	68.2 ± 6.1	72 ± 7.9	0.126	0.13
24h PR [bpm]	71.2 ± 9.8	72.9 ± 7.5	71.2 ± 6.5	72 ± 8.3	0.929	0.84
Non-dipper status [n (%)] †	4 (28.6%)	2 (18.2%)	1 (16.7%)	2 (40%)	0.723	0.685
LVMI [g/m ²]	86.1 ± 18.1	84.2 ± 17	86.6 ± 19	88.9 (± 24.0)	0.931	0.61
LVH [n (%)]	1 (5.9%)	3 (20%)	2 (11.76%)	7 (46.7%)	0.036	0.213
LAVI [ml/m ²]	26.6 ± 9.6	24 ± 8.6	24 ± 9.44	20.3 ± 7.1	0.258	0.2
Maximum CIMT [mm]	1 (0.9 - 1.2)	1 (0.9 - 1.1)	1 (0.9 - 1.1)	0.9 (0.9 - 1.1)	0.685	0.393
ASP [n(%)]	7 (41.2%)	4 (26.7%)	4 (23.5%)	4 (26.7%)	0.709	0.585
SCORE2/-OP [%] ‡	8 (7 - 12)	12 (8 - 15.5)	7 (5 - 14.5)	8.5 (8 - 11.5)	0.572	0.29
FHS-ASCVD Risk [%]	14.3 (9.2 - 24.5)	11.2 (7.8 - 16.2)	7.1 (4.7 - 14.2)*	5.6 (3.8 - 10)*	0.003	0.002

AI patients were categorized based on catestatin concentrations into those in the lower and upper half, and further into four subgroups (two lowest were identical with first and second quartiles). Data are presented as number (percentage), mean ± standard deviation or median(interquartile range) depending on distribution; p-values were calculated with one-way ANOVA and Tukey's HSD tests (quantitative variables) or Fisher's exact test (categorical variables) with Benjamini-Hochberg correction for multiple testing. Bold font denotes significant (<0.05) p values; * denotes datapoints significantly different versus those for the 'very low' Cts subgroup in post hoc test; # denotes datapoints significantly different versus those for the 'low Cts' subgroup in post hoc test; † dipper status was considered only in patients with HT. ‡ Only nondiabetic patients were included in SCORE2/-OP risk estimation (n=11, 11, 16, and 14 for consecutive subgroups). ABPM, ambulatory blood pressure monitoring; ASP, atherosclerotic plaques; BMI, body mass index; bpm, beats per minute; CCA, common carotid artery; CIMT, carotid intima media thickness; DBP, diastolic blood pressure; DMt2, diabetes mellitus type 2; FHS-ASCVD Risk, 10-year atherosclerotic cardiovascular disease risk calculated based on data from the Framingham Heart Study; HDL-C, high density lipoprotein cholesterol; HT, hypertension; hs-CRP, high sensitivity C-reactive protein; LDL-C, low density lipoprotein cholesterol; LVH, left-ventricular hypertrophy; LVMI, left ventricular mass index; max, maximum; MACS, mild autonomous cortisol secretion; MetS, metabolic syndrome; PPI, proton pump inhibitor; PR, pulse rate; SCORE2/-OP, Systematic Coronary Risk Estimation 2/-Older People; SBP, systolic blood pressure; TC, Total cholesterol; TGL, triglycerides.

without an adrenal adenoma (28, 29), which is what we found here for AI patients compared to age-, sex-, and BMI-matched controls after adjusting for confounding factors. Other researchers showed higher plasma CgA in patients with an adrenal adenoma than in subjects without one, which may underlie a slightly higher unadjusted Cts in our AI patients than controls (24, 25). What should be noticed is a similar distribution of Cts levels in controls and AI patients, including the proportion of subjects with high (>97 ng/ml) Cts: 21% in the former and 23% in the latter. This further suggests that the presence of an AI does not affect Cts. Regarding hormonal activity, we found no differences in Cts between patients with MACS and NFAI, nor correlations between Cts and UFC or 1-mg DST cortisol, possibly due to small sample size.

Despite comparable age, BMI, male-to-female ratio, smoking status and comorbidities, IVSd, LVPWd, LVM, and LVMI were higher in AI patients than controls, which was driven by values recorded in subjects with MACS. Iacobellis et al. reported similar results (higher LVM in AI subjects versus controls, the difference depended on patients with MACS) (30). In our study, SCORE2-OP was higher in MACS compared to NFAI patients and controls, which indicates increased CV risk associated with subclinical hypercortisolism. The data support the hypothesis that chronic mild elevation of cortisol levels in AI patients adversely affect the CV system rather than the presence of an adrenal adenoma *per se*. Low Cts was associated here with a higher prevalence of male gender, HT, MetS, as well as BMI, 24h SBP, UA and lower HDL-C. Consequently, an association between Cts and ASCVD risk was recorded (Figure 6). In line with our results, Cts was lower in obese children with MetS than in those without it, and in normal-weight controls (13); O'Connor et al. showed Cts correlated negatively with BMI (12), and Durakoglugil et al. reported a positive correlation

between Cts and HDL-C (14). In the latter study, a negative correlation between plasma Cts and TGL concentration was also observed (14), which was not confirmed here. Surprisingly, among participants without statins, we recorded weak positive correlations between Cts and TC as well as LDL-C. The former may be connected with a positive correlation between Cts and HDL-C, however, the latter is difficult to explain, and requires further clarification.

To date, there are controversies regarding Cts in HT; lower Cts levels have been associated with this disease (12, 14, 31–35) (Figure 6). Here, Cts in AI subjects with HT was lower than in normotensive ones, however, the difference was not significant in the controls and we recorded no correlations between Cts and ABPM results. Our data add important facts to the discussion: low Cts levels are more common in HT, however, some patients do exhibit intermediate and high concentrations of the peptide. This was observed in individuals with effective hypotensive treatment revealed by 24-h monitoring.

Further, no significant associations were recorded here between Cts and TTE as well as CCA USG parameters. Small sample size clearly limits conclusions that can be drawn from these data. More sensitive methods (e.g. global longitudinal strain and microvasculature assessment) may have yielded different results.

Possibly, the most intriguing question is the clinical significance of high versus very low/low Cts in individuals with similar established CV risk factors. For instance, non-smoking females aged ca. 60, with overweight and HT (FHS-ASCVD Risk between 10 and 20%) were recorded both in the first half and highest quarter of Cts levels among AI patients. Longitudinal assessment of much larger populations is required to determine whether Cts provides protection against CVD. If so, determining therapeutic strategies that stimulate Cts would be beneficial.

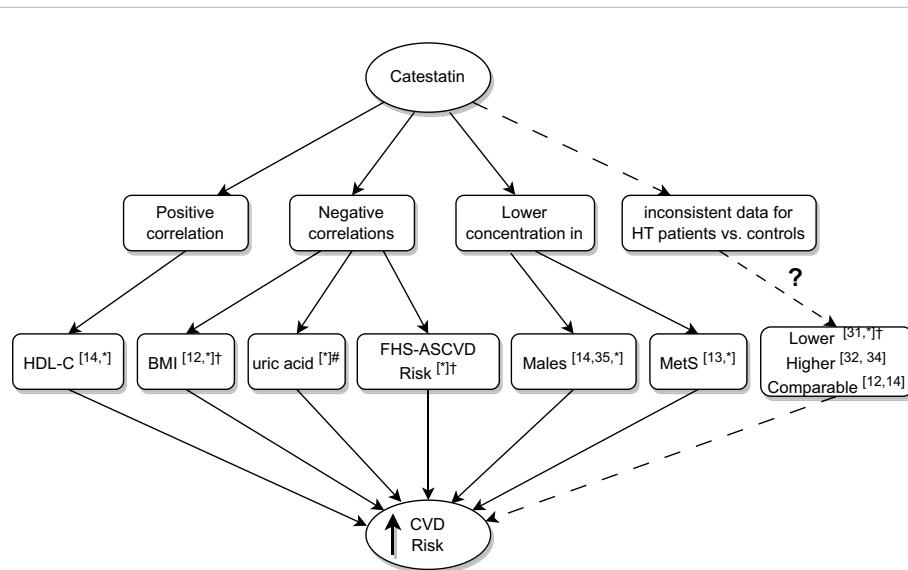


FIGURE 6

FIGURE 3
 Summary of research on associations between low catestatin and cardiovascular risk. Superscript numbers indicate references to previous studies; *indicate results of the current study; #correlation between uric acid and Cts did not reach significance in AI patients analyzed here, it did upon analyzing AI patients and controls together; [†]significance achieved in AI patients. AI, adrenal incidentaloma, BMI, body mass index; Cts, catestatin; CVD, cardiovascular disease; FHS-ASCVD Risk, 10-year atherosclerotic cardiovascular disease risk calculated based on the Framingham Heart Study; HDL-C, high density lipoprotein cholesterol; HT, hypertension; MetS, Metabolic Syndrome.

5 Conclusions

We are the first to report that among persons without overt CVD other than primary HT, plasma Cts concentrations in patients with an AI are comparable to those of matched controls with normal adrenal morphology. Correlations between Cts and: HDL-C (positive) as well as BMI, UA and FHS-ASCVD Risk (negative) point at cardioprotective effects of the peptide. Data from ABPM, TTE and CCA intima-media assessment did not yield associations between Cts and BP or HT-mediated organ damage. It must be highlighted that many factors influence Cts, and further research is necessary to apply it as a biomarker.

Data availability statement

The original contributions presented in the study are included in the article/[Supplementary Material](#). Further inquiries can be directed to the corresponding author.

Ethics statement

The studies involving human participants were reviewed and approved by the Independent Bioethics Committee for Scientific Research at Medical University of Gdańsk (NKBB/659/2019). The patients/participants provided their written informed consent to participate in this study.

Author contributions

EZ secured ethical approval for the study, analyzed the data, and reviewed the literature. EZ and PK collected the data and wrote the manuscript. JS and AK performed echocardiography and ultrasound examination of the common carotid artery. PK and KS carried out critical interpretations. All authors contributed to the article, approved the submitted version, and are accountable for the content of the work.

Funding

The study was supported by the project POWR.03.05.00-00-z082/18 co-financed by the European Union through the European Social Fund under the Operational Programme Knowledge Education Development 2014–2020 and by the Medical University of Gdańsk (grant MB 2022: 01-10022 and MN 01-350/08/126).

References

1. Yu W, Cheng JD. Uric acid and cardiovascular disease: an update from molecular mechanism to clinical perspective. *Front Pharmacol* (2020) 11:582680. doi: 10.3389/fphar.2020.582680
2. Koosha P, Roohafza H, Sarrafzadeh N, Vakhshoori M, Talaei M, Sheikhbahaei E, et al. High sensitivity c-reactive protein predictive value for cardiovascular disease: a nested case control from isfahan cohort study (ICS). *Glob Heart* (2020) 15(1):1–13. doi: 10.5334/GH.367

Acknowledgments

The authors would like to thank Anna Koelmer and Peter Greśner from Center for Biostatistics and Bioinformatics of Medical University of Gdańsk and Piotr Wiśniewski for support in data analysis.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2023.1198911/full#supplementary-material>

SUPPLEMENTARY FIGURE 1

Catestatin in normotensive and hypertensive AI patients and controls – analyses in subjects of each sex. Boxplot chart. P-values determined using the Mann-Whitney U test. AI – adrenal incidentaloma.

SUPPLEMENTARY FIGURE 2

Catestatin in subjects without and with metabolic syndrome in controls and AI patients – analyses in subjects of each sex. Boxplot chart. P-values determined using the Mann-Whitney U test. AI – adrenal incidentaloma; MetS – metabolic syndrome.

SUPPLEMENTARY FIGURE 3

Correlations between catestatin and LDL-C level in AI patients and controls. Correlations were computed by the Spearman rank-order method and were tested separately in subjects with and without statin therapy. AI – adrenal incidentaloma, LDL-C – low density lipoprotein cholesterol, ln – natural logarithm.

SUPPLEMENTARY FIGURE 4

Correlations between catestatin and total cholesterol level in AI patients and controls. Correlations were computed by the Spearman rank-order method and were tested separately in subjects with and without statin therapy. AI – adrenal incidentaloma, ln – natural logarithm.

3. Mozaffarian D, Wilson PWF, Kannel WB. Beyond established and novel risk factors lifestyle risk factors for cardiovascular disease. *Circulation* (2008) 117(23):3031–8. doi: 10.1161/CIRCULATIONAHA.107.738732
4. Mahata SK, Corti A. Chromogranin a and its fragments in cardiovascular, immunometabolic, and cancer regulation. *Ann N Y Acad Sci* (2019) 1455(1):34–58. doi: 10.1111/nyas.14249
5. Zalewska E, Kmiec P, Sworczak K. Role of cathestatin in the cardiovascular system and metabolic disorders. *Front Cardiovasc Med* (2022) 9:909480. doi: 10.3389/fcvm.2022.909480
6. Mahata SK, Mahapatra NR, Mahata M, Wang TC, Kennedy BP, Ziegler MG, et al. Catecholamine secretory vesicle stimulus-transcription coupling *in vivo*. demonstration by a novel transgenic promoter/photoprotein reporter and inhibition of secretion and transcription by the chromogranin a fragment cathestatin. *J Biol Chem* (2003) 278(34):32058–67. doi: 10.1074/jbc.M305545200
7. Angelone T, Quintieri AM, Brar BK, Limchaiyawat PT, Tota B, Mahata SK, et al. The antihypertensive chromogranin a peptide cathestatin acts as a novel endocrine/paracrine modulator of cardiac inotropism and lusitropism. *Endocrinology* (2008) 149(10):4780–93. doi: 10.1210/en.2008-0318
8. Bandyopadhyay GK, Vu CU, Gentile S, Lee H, Biswas N, Chi NW, et al. Cathestatin (Chromogranin A352–372) and novel effects on mobilization of fat from adipose tissue through regulation of adrenergic and leptin signaling. *J Biol Chem* (2012) 287(27):23141–51. doi: 10.1074/jbc.M111.335877
9. Bandyopadhyay G, Tang K, Webster NJG, van den Bogaart G, Mahata SK. Cathestatin induces glycogenesis by stimulating the phosphoinositide 3-kinase-AKT pathway. *Acta Physiol (Oxf)* (2022) 235(1):e13775. doi: 10.1111/apha.13775
10. Chen Y, Wang X, Yang C, Su X, Yang W, Dai Y, et al. Decreased circulating cathestatin levels are associated with coronary artery disease: the emerging anti-inflammatory role. *Atherosclerosis* (2019) 281:78–88. doi: 10.1016/j.atherosclerosis.2018.12.025
11. Wang X, Xu S, Liang Y, Zhu D, Mi L, Wang G, et al. Dramatic changes in cathestatin are associated with hemodynamics in acute myocardial infarction. *Biomarkers* (2011) 16(4):372–7. doi: 10.3109/1354750X.2011.578260
12. O'Connor DT, Kailasam MT, Kennedy BP, Ziegler MG, Yanaihara N, Parmer RJ. Early decline in the catecholamine release-inhibitory peptide cathestatin in humans at genetic risk of hypertension. *J Hypertens* (2002) 20(7):1335–45. doi: 10.1097/0004872-200207000-00020
13. Simunovic M, Supe-Domic D, Karin Z, Degoricija M, Paradzik M, Bozic J, et al. Serum cathestatin concentrations are decreased in obese children and adolescents. *Pediatr Diabet* (2019) 20(5):549–55. doi: 10.1111/pedi.12825
14. Durakoglugil ME, Ayaz T, Kocaman SA, Kirbaş A, Durakoglugil T, Erdogan T, et al. The relationship of plasma cathestatin concentrations with metabolic and vascular parameters in untreated hypertensive patients: influence on high-density lipoprotein cholesterol. *Anatol J Cardiol* (2015) 15(7):577–85. doi: 10.5152/akd.2014.5536
15. Williams B, Mancia G, Spiering W, Agabiti Rosei E, Azizi M, Burnier M, et al. ESC/ESH guidelines for the management of arterial hypertension: the task force for the management of arterial hypertension of the European society of cardiology (ESC) and the European society of hypertension (ESH). *Eur Heart J* (2018) 39(33):3021–104. doi: 10.1093/eurheartj/ehy339
16. Yağız B, Akalın A, Yorulmaz G, Macunluoğlu AC, Yağız O. Can non-functional adrenal incidentaloma be ranked among cardiovascular risk factors? *Eur Res J* (2022) 8(6):747–54. doi: 10.18621/eurj.872835
17. Fassnacht M, Arlt W, Bancos I, Dralle H, Newell-Price J, Sahdev A, et al. Management of adrenal incidentalomas: European society of endocrinology clinical practice guideline in collaboration with the European network for the study of adrenal tumors. *Eur J Endocrinol* (2016) 175(2):G1–34. doi: 10.1530/EJE-16-0467
18. Alberti KGMM, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JL, Donato KA, et al. Harmonizing the metabolic syndrome: a joint interim statement of the international diabetes federation task force on epidemiology and prevention; national heart, lung, and blood institute; American heart association; world heart federation; international. *Circulation* (2009) 120(16):1640–5. doi: 10.1161/CIRCULATIONAHA.109.192644
19. D'Agostino RB, Vasan RS, Pencina MJ, Wolf PA, Cobain M, Massaro JM, et al. General cardiovascular risk profile for use in primary care: the framingham heart study. *Circulation* (2008) 117(6):743–53. doi: 10.1161/CIRCULATIONAHA.107.699579
20. American Heart Association. (n.d.). 2018 Prevention Guidelines Tool CV Risk Calculator. Available at: <https://static.heart.org/riskcalc/app/index.html#/baseline-risk/>.
21. Visseren FLJ, Mach F, Smulders YM, Carballo D, Koskinas KC, Bäck M, et al. ESC Guidelines on cardiovascular disease prevention in clinical practice. *Eur Heart J* (2021) 42(34):3227–337. doi: 10.1093/eurheartj/ehab484
22. Williams B, Mancia G, Spiering W, Agabiti Rosei E, Azizi M, Burnier M, et al. ESC/ESH guidelines for the management of arterial hypertension. *Eur Heart J* (2018) 39(33):3021–104. doi: 10.1093/eurheartj/ehy339
23. Lang RM, Badano LP, Mor-Avi V, Afilalo J, Armstrong A, Ernande L, et al. Recommendations for cardiac chamber quantification by echocardiography in adults: an update from the American society of echocardiography and the European association of cardiovascular imaging. *Eur Heart J Cardiovasc Imaging* (2015) 16(3):233–71. doi: 10.1093/ejhci/jev014
24. Starling MR, Walsh RA. Accuracy of biplane axial oblique and oblique cineangiographic left ventricular cast volume determinations using a modification of simpson's rule algorithm. *Am Heart J* (1985) 110(6):1219–25. doi: 10.1016/0002-8703(85)90016-x
25. Mahata SK, Mahata M, Yoo SH, Taupenot L, Wu H, Aroda VR, et al. A novel, catecholamine release-inhibitory peptide from chromogranin a: autocrine control of nicotinic cholinergic-stimulated exocytosis. *Adv Pharmacol* (1998) 42(1):260–4. doi: 10.1016/S1054-3589(08)60743-7
26. Zhu D, Wang F, Yu H, Mi L, Gao W. Cathestatin is useful in detecting patients with stage b heart failure. *Biomarkers* (2011) 16(8):691–7. doi: 10.3109/1354750X.2011.629058
27. Glinicki P, Jeske W. Chromogranin a (CgA) - the influence of various factors *in vivo* and *in vitro*, and existing disorders on its concentration in blood. *Endokrynol Pol* (2010) 61(4):384–7.
28. Bernini G, Moretti A, Fontana V, Orlandini C, Miccoli P, Berti P, et al. Plasma chromogranin a in incidental non-functioning, benign, solid adrenocortical tumors. *Eur J Endocrinol* (2004) 151(2):215–22. doi: 10.1530/eje.0.1510215
29. Bernini GP, Moretti A, Borgioli M, Bardini M, Miccoli P, Berti P, et al. Plasma and tissue chromogranin in patients with adrenocortical adenomas. *J Endocrinol Invest* (2004) 27(9):821–5. doi: 10.1007/BF03346275
30. Iacobellis G, Petramala L, Barbaro G, Kargi AY, Serra V, Zinnamosca L, et al. Epicardial fat thickness and left ventricular mass in subjects with adrenal incidentaloma. *Endocrine* (2013) 44(2):532–6. doi: 10.1007/s12020-013-9902-5
31. O'Connor DT, Zhu G, Rao F, Taupenot L, Fung MM, Das M, et al. Heritability and genome-wide linkage in US and australian twins identify novel genomic regions controlling chromogranin a: implications for secretion and blood pressure. *Circulation* (2008) 118(3):247–57. doi: 10.1161/CIRCULATIONAHA.107.709105
32. Meng L, Ye XJ, Ding WH, Yang Y, Di BB, Liu L, et al. Plasma catecholamine release-inhibitory peptide cathestatin in patients with essential hypertension. *J Cardiovasc Med* (2011) 12(9):643–7. doi: 10.2459/JCM.0b013e328346c142
33. Tütün N, Güralp O, Gökk K, Hamzaoglu K, Oner YO, Makul M, et al. Serum cathestatin level is increased in women with preeclampsia. *J Obstet Gynaecol (Lahore)* (2022) 42(1):55–60. doi: 10.1080/01443615.2021.1873922
34. Kumric M, Vrdoljak J, Dujic G, Supe-Domic D, Ticinovic Kurir T, Dujic Z, et al. Serum cathestatin levels correlate with ambulatory blood pressure and indices of arterial stiffness in patients with primary hypertension. *Biomolecules* (2022) 12(9). doi: 10.3390/biom12091204
35. Fung MM, Salem RM, Mehtani P, Thomas B, Lu CF, Perez B, et al. Direct vasoactive effects of the chromogranin a (CHGA) peptide cathestatin in humans *In vivo*. *Clin Exp Hypertens* (2011) 32(5):278–87. doi: 10.1161/CIRCULATIONAHA.110.956839



Role of Catestatin in the Cardiovascular System and Metabolic Disorders

Ewa Zalewska, Piotr Kmiec* and Krzysztof Sworczak

Department of Endocrinology and Internal Medicine, Medical University of Gdańsk, Gdańsk, Poland

Catestatin is a multifunctional peptide that is involved in the regulation of the cardiovascular and immune systems as well as metabolic homeostasis. It mitigates detrimental, excessive activity of the sympathetic nervous system by inhibiting catecholamine secretion. Based on *in vitro* and *in vivo* studies, catestatin was shown to reduce adipose tissue, inhibit inflammatory response, prevent macrophage-driven atherosclerosis, and regulate cytokine production and release. Clinical studies indicate that catestatin may influence the processes leading to hypertension, affect the course of coronary artery diseases and heart failure. This review presents up-to-date research on catestatin with a particular emphasis on cardiovascular diseases based on a literature search.

OPEN ACCESS

Edited by:

Jian Shi,

University of Leeds, United Kingdom

Reviewed by:

Teresa Pasqua,

University of Calabria, Italy

Geert Van Den Bogaart,

University of Groningen, Netherlands

***Correspondence:**

Piotr Kmiec

piotr.kmiec@gumed.edu.pl

Specialty section:

This article was submitted to
Cardiovascular Metabolism,
a section of the journal

Frontiers in Cardiovascular Medicine

Received: 31 March 2022

Accepted: 25 April 2022

Published: 19 May 2022

Citation:

Zalewska E, Kmiec P and Sworczak K
(2022) Role of Catestatin in the
Cardiovascular System and Metabolic
Disorders.

Front. Cardiovasc. Med. 9:909480.
doi: 10.3389/fcvm.2022.909480

INTRODUCTION

The sympathetic nervous system is crucial in preserving homeostasis in humans. However, its excessive activity has been recognized to underlie pathologic processes of many cardiovascular diseases (CVDs), which are the leading cause of death globally (1). Among others, upregulated sympathetic nervous system activity has been associated with hypertension (HT), adverse myocardial remodeling, arrhythmias, sudden cardiac death, and overall poor prognosis in patients with heart failure (HF) (2).

Basal and reflex control of the sympathetic activity associated with cardiovascular function occurs in the rostral ventrolateral medulla (3). It sends catecholaminergic projections to the sympathetic preganglionic neurons of the spinal cord. In turn, preganglionic neurons release acetylcholine to activate postganglionic neurons and chromaffin cells of the adrenal medulla, which synthesize and secrete catecholamines (CAs): norepinephrine and epinephrine (stored in sufficient quantities within the cells, which is the reason for a positive chromaffin reaction, and, hence, their name) (3, 4). CAs target α - and β -adrenergic receptors, a family of G protein-coupled receptors. Adrenoceptors are divided into α_1 , α_2 , β_1 , β_2 , and β_3 subtypes. In the smooth muscle cells of blood vessels, there are mainly α_1 -adrenergic receptors, coupled to stimulatory Gq proteins, which induce constriction by activating phospholipase C. In the heart, the predominant receptor subtype β_1 activates the Gs-adenylyl cyclase – adenosine monophosphate – protein kinase A signaling cascade to induce positive inotropic and chronotropic effects (5). α_2 receptors are coupled to inhibitory Gi proteins that inactivate adenylyl cyclase and are mainly found in the central nervous system, where their activation lowers arterial blood pressure (BP) (5).

In 1965, Banks and Helle reported that CA secretion from chromaffin granules of the adrenal medulla is associated with the release of soluble proteins (6). In July 1967, Blaschko's group coined the term "chromogranin A" (CgA) for the major component of these proteins (7, 8). In 1988, Simon et al. reported for the first time that proteolytic hydrolysis of CgA, obtained from cultured bovine adrenal medullary chromaffin cells, generated a peptide product that is capable of inhibiting CAs release, a promising and novel mechanism for counteracting the sympathetic outflow (9, 10). However, the exact identity of this functional peptide remained elusive until 1997, when Mahata et al. synthesized 15 peptides spanning ~80% of the entire CgA molecule and demonstrated that only one, bovine CgA_{344–364} [RSMRLSFRARGYGFGRGPGQLQ], inhibited CA secretion induced by nicotine (11). They named it "catestatin" (Cts) due to its high capacity to suppress the release of CAs (11).

Cts was initially considered a regulatory peptide that inhibits CA secretion by acting as a noncompetitive mediator of the nicotinic cholinergic stimulation of chromaffin cells and sympathetic neurite outgrowth (11, 12). Further studies showed that Cts inhibits CA release also due to adenylate cyclase-activating polypeptide stimulation and regulates dense core vesicle quanta (13) (**Figure 1**). Moreover, Cts has emerged as a pleiotropic peptide, which – among others – is involved in the regulation of the cardiovascular and immune system, as well as metabolic homeostasis (14, 17, 25–27).

This review presents up-to-date research on the role of Cts in the cardiovascular system and metabolic disorders contributing to CVDs, and Cts as a putative clinical biomarker. It is based on an electronic literature search of PubMed database performed April 20, 2022, using the key term 'catestatin', which yielded 235 results. Papers were included based on screening of titles and abstracts.

CATESTATIN BIOLOGY

Cts (human CgA_{352–372}, bovine CgA_{344–364}, and rat CgA_{367–387}) is a neuroendocrine peptide that is derived from the proteolytic cleavage of its precursor compound CgA. Human chromogranin A (hCgA) consists of nine dibasic sites and is cleaved by prohormone convertases, namely, cathepsin L, plasmin, and kallikrein, which generates: 1) Cts, which is a 21-amino acid, hydrophobic peptide derived from hCgA's C terminal fragment, 2) a dysglycemic peptide pancreastatin (hCgA_{250–301}), 3) a vasodilating, antiadrenergic, and antiangiogenic peptide vasostatin-1 (hCgA_{1–76}), 4) a peptide that acts as an antigen for diabetogenic CD4+ T-cell clones WE14 (hCgA_{324–337}), and, 5) a poadrenergic peptide serpinin (hCgA_{411–436}) (15).

Abbreviations: Akt, Protein kinase B; AMI, acute myocardial infarction; BP, blood pressure; Ca²⁺, calcium ions; CAD, coronary artery disease; CA, catecholamine; CgA, Chromogranin A; Cts-KO, catestatin knockout; CVD, cardiovascular disease; Chga-KO, Chromogranin knockout; Cts, catestatin; hCgA, human CgA; HF, heart failure; HT, hypertension; I/R injury, ischemia/reperfusion injury; nAChR, nicotinic acetylcholine receptor; NO, nitric oxide; nNOS, neuronal nitric oxide synthase; NYHA, New York Heart Association; OSA, obstructive sleep apnea; PMNs, Polymorphonuclear neutrophils; WT, wild type.

Cts includes a sequence [SSMKLSFRARAYGFRGPGPQL] that is highly conserved across various species and is flanked by proteolytic cleavage sites (11, 28). Five single-nucleotide polymorphisms have been identified in the hCgA gene (*CHGA*) region expressing Cts: Gly364Ser (rs9658667), Pro370Leu (rs965868), Arg374Gln (rs9658669) (29), Tyr363Tyr (rs9658666), and Gly367Val (rs200576557) (15, 30, 31). Intriguingly, a genome-wide association study identified two loci that affect Cts concentrations in regions that include genes encoding kallikrein and Factor XII; these enzymes participating in a proteolytic cascade (Factor XII activates prokallikrein to kallikrein, which activates FXII) were shown to generate active compounds from chromogranin A and B cleavage *in vivo* and *in vitro* (32). In particular, kallikrein produced a 12-amino-acid CgA-fragment (CgA_{361–372}) with preserved biological activity of 21-amino-acid Cts (32).

Apart from limiting CA secretion, Cts is a potent inhibitor of the release of other chromaffin cell cotransmitters, including neuropeptide Y, adenosine triphosphate, and CgA; it is widely distributed in secretory granules of the chromaffin cells, diffuse neuroendocrine system, neuronal cells, bone-marrow derived cells, the auditory system, and the heart (20, 24, 33, 34) (**Figure 1**). In the latter, processing of CgA can be carried out by extracellular proteases on both cardiomyocyte cell membrane and in the extracellular matrix (33, 35). In a rat model, it was shown that the heart generates intracardiac CgA fragments, including locally derived Cts, in response to hemodynamic and excitatory challenges (36). Moreover, recently, a CgA_{1–373} fragment that encompasses the Cts domain was found to elicit direct cardiac effects both *in vitro* and *ex vivo* (37).

CATESTATIN AND REGULATION OF BLOOD PRESSURE

In vitro and *in vivo* Animal Studies

In vitro, it was shown Cts binds to α, β, δ, and γ subunits of nicotinic acetylcholine receptors (nAChR) with a high affinity binding site on the β subunit near the membrane surface, which blocks sodium ions' uptake, thus inhibiting membrane depolarization, and blocking the influx of calcium ions (Ca²⁺) through voltage-gated calcium channels (38, 39). Inhibition of Ca²⁺ influx suppresses both CA release by exocytosis (all-or-none secretion) and *CHGA* transcription through a pathway involving the activation of protein kinase C and mitogen-activated protein kinase (40) (**Figure 1**).

Human Cts polymorphic variants were shown to exhibit varying potency of nAChR inhibition *in vitro* using a rat pheochromocytoma cell line and the human receptor (30, 41, 42). One should bear in mind that lower potency of a Cts variant in this respect does not necessarily translate to higher plasma CAs, since in such a case lower desensitization to CA secretion occurs (i.e., an effect due to repeated exposure of the nAChR to Cts). Further, since Cts blood-lowering effect depends partly on the release of nitric oxide (NO), human Cts Gly364Ser variant (see Clinical Studies) was shown to display lower NO-triggering and

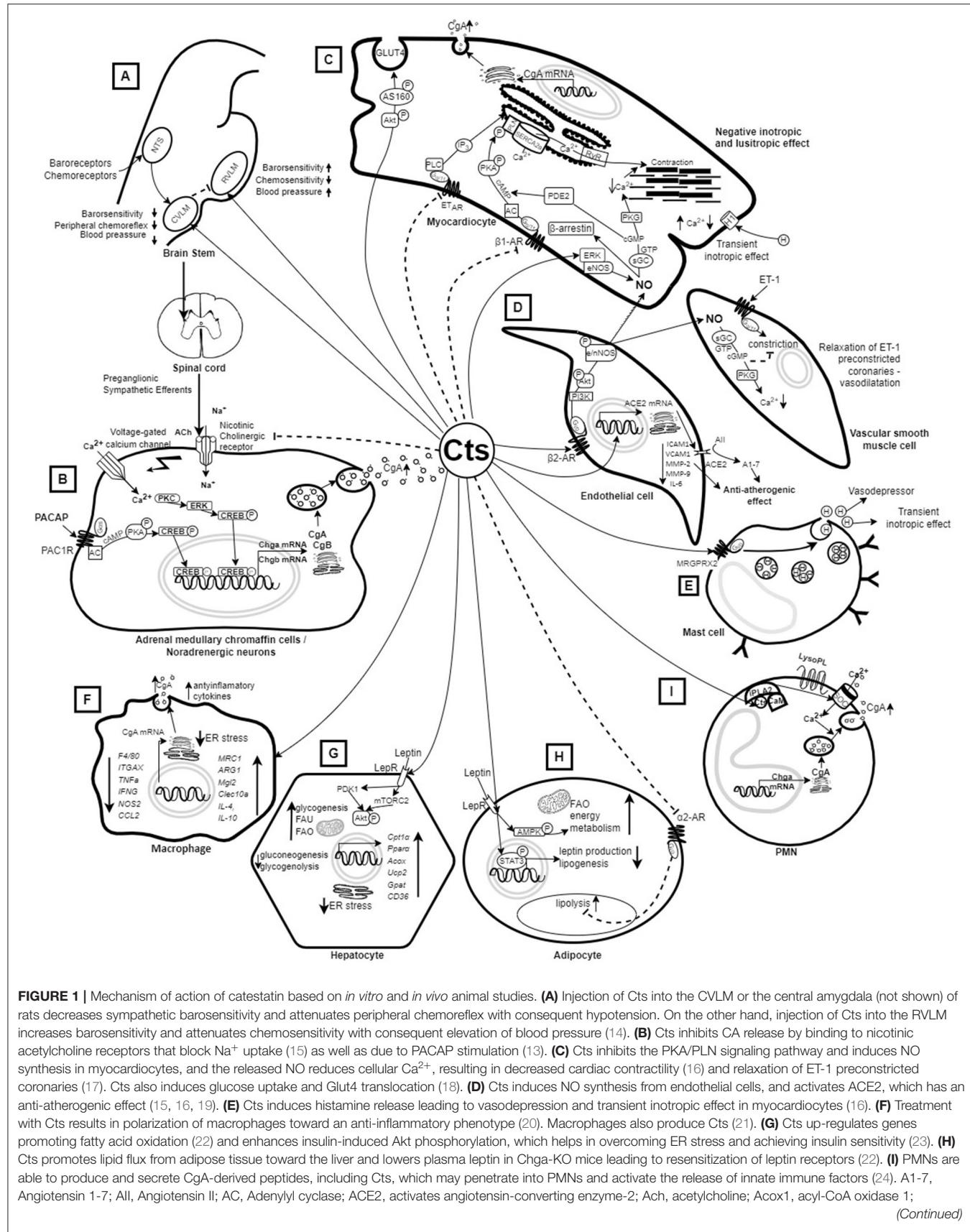


FIGURE 1 | Akt, Protein kinase B; AMPK, AMP-activated protein kinase; ARG1, Arginase 1 gene; β 1AR, β 1 adrenergic receptors; β 2AR, β 2 adrenergic receptors; Ca^{2+} , calcium ions; cAMP, adenosine monophosphate; CAs, catecholamines; CCL2, C-C Motif Chemokine Ligand 2; CD36, cluster of differentiation 36; CgA, Chromogranin A; CgB, Chromogranin B; Chga-KO, Chromogranin knockout; cGMP, cyclic guanosine monophosphate; Cpt1 α , Carnitine palmitoyltransferase 1 α ; CREB, cAMP response element-binding protein; Cts, catestatin; CVLM, caudal ventrolateral medulla; DNL, *de novo* lipogenesis; eNOS, endothelial nitric oxide synthase; ER, endoplasmic reticulum; ERK, extracellular signal-regulated kinases; ET-1, endothelin 1; ETAR, Endothelin receptor type A; ETBR, Endothelin receptor type B; FAO, Fatty acid oxidation; FAU, Fatty acid uptake; Gpat4, lipogenic gene glycerol-3-phosphate acyltransferase; GTP, guanosine-5' triphosphate; H, histamine; ICAM1, Intercellular Adhesion Molecule 1; IFNG, Interferon Gamma gene; IL-4, interleukin 4; IL-6, interleukin 6; IL-10, interleukin 10; iPLA2, calcium-independent phospholipase A2; ITGAX, Integrin Subunit Alpha X; LepR, Leptin receptor; LysoPL, lysophospholipids; MMP-2 Matrix Metallopeptidase 2; MMP-9, Matrix metallopeptidase 9; MRC1, Mannose Receptor C-Type 1 gene; MRGPRX2, Mas-Related G Protein-Coupled Receptor-X2; Na+, sodium; NO, nitric oxide; NOS2, Nitric Oxide Synthase 2; nNOS, neuronal nitric oxide synthase; NTS, Nucleus tractus solitarius; P, phosphor; PACAP, Pituitary adenylate cyclase-activating polypeptide; PAC1R, Pituitary adenylate cyclase-activating polypeptide receptor; PDE2, Phosphodiesterase 2; PI3K, Phosphoinositide 3-kinase; PKA, Protein kinase A; PKC, protein kinase C; PKG, protein kinase G; PLN, phospholamban; PMNs, Polymorphonuclear neutrophils; Ppar α , Peroxisome proliferator-activated receptor- α ; RVLM, rostral ventrolateral medulla; RyR, Ryanodine receptor; sGC, soluble guanylyl cyclase; SERCA, Sarcoplasmic reticulum Ca-ATPase; SOC, Store-Operated Calcium Channels; STAT3, Signal Transducer And Activator Of Transcription 3; TAG, Triacylglycerols; TNFa, TNF alpha gene; Ucp2, uncoupling protein 2; VCAM1, vascular cell adhesion molecule 1. Dashed arrow – inhibition; continuous arrow – stimulation.

absent anti-adrenergic activity compared to wild-type peptide (WT-Cts) *in vitro* (31, 43).

This *in vitro* research concerning CA release suppression by Cts is supported by *in vivo* studies with genetically modified rodents. First, knockout of the CgA gene region encoding Cts (Cts-KO) in mice resulted in HT, left ventricular hypertrophy, and elevated CAs, yet, high BP was abated by intraperitoneal injection of exogenous Cts (21). Moreover, this strain exhibited marked cardiac and adrenal macrophage infiltration as well as increased proinflammatory cytokine levels, which may trigger CA release and escalate HT (21). Chlodronate depletion of macrophages, and bone marrow transfer between Cts-KO and WT mice demonstrated that immunosuppression of macrophages by Cts partly underlies its antihypertensive and anti-inflammatory effects (21).

Second, ablation of the CgA gene in another mouse model (Chga-KO) led to – among others – high baseline BP and elevated plasma CAs as well as exaggerated pressor and depressor responses to phenylephrine and sodium nitroprusside (44–46). Exogenous Cts replacement selectively diminished stress-induced increments in BP and HR (45) and restored the sensitivity of high-pressure baroreceptors, i.e., attenuation of both reflex tachycardia (due to sodium nitroprusside-induced hypotension) and reflex bradycardia (following phenylephrine-induced HT) occurred (45). This indicates Cts functions efficiently as an antihypertensive peptide even under stressful conditions.

Third, in a genetically humanized mice strain expressing insufficient hCgA amounts, increased plasma CA levels as well as elevated systolic and diastolic BP were recorded in comparison to WT controls and a strain with sufficient hCgA expression (47).

However, Kennedy et al. showed Cts did not affect plasma norepinephrine levels and actually significantly (11-fold) increased those of epinephrine following electrical stimulation of male Sprague-Dawley rats, although it did reduce BP increases (even with anti-adrenergic pretreatment) (48). The possible explanation of these results, which contrast with findings from *in vitro* and Cst- and CgA-knockout studies, is that epinephrine release was compensatory to effects triggered by Cts in acute injury by electric stimulation, which could not have been observed *in vitro* (due to study limitations) and in KO mice owing

to significant genetic alterations. Mitigation of pressor responses by Cts in the study by Kennedy et al. was attributed to histamine release: it increased 21-fold within 2 min of Cts injection, and the BP lowering effect was abolished by hydroxyzine pretreatment, which was also the case for epinephrine elevation (48). Krüger et al. confirmed this hypothesis *in vitro*: Cts lead to histamine release from rat mast cells *via* a peptidergic pathway – by activating the G protein, because this effect was suppressed by the pertussis toxin, a Gi/Go inactivator (49) (**Figure 1**).

Furthermore, although plasma Cts concentrations gradually increased with the progression of HT in spontaneously hypertensive rats, exogenous Cts reduced HR (indicating anti-sympathetic activity) (50), as well as ameliorated vascular, renal and cardiac proliferation (51). *In vitro*, Cts was also reported to act as a potent inhibitor of isoproterenol- and endothelin-1-mediated activities in the frog (52) and rat heart (53). Angelone et al. showed that WT-Cts increased heart rate and decreased left ventricular pressure, rate-pressure product, and, both positive and negative left ventricular contractility. The authors suggested that these negative inotropic and lusitropic effects of WT-Cts may contribute to its hypotensive action (53) (**Figure 1**).

Concerning the central nervous system, Cts plays an important role in cardiorespiratory control (14). Gaede et al. showed that Cts antagonizes both nAChR and β -adrenergic receptors involved in cardiovascular regulation using urethane-anesthetized, vagotomized rats. Cts mitigated the hypertensive effect of nicotine and prevented increased splanchnic sympathetic activity caused by isoproterenol, a non-selective β adrenergic receptor agonist (54). The results of the study indicate Cts may affect BP levels in HT (54). Further, depending on the region of the medulla, Cts exerted sympathoexcitatory or cholinergic effects (3, 55, 56). Injection of Cts into the rostral ventrolateral medulla (a key site for BP control in the brain stem) resulted in increased barosensitivity and attenuation of chemosensitivity with consequent BP elevation (3). On the other hand, injection of Cts into the caudal ventrolateral medulla (55) and the central amygdala (56) (both contain inhibitory neurons of the rostral ventrolateral medulla) of rats resulted in decreased sympathetic barosensitivity and attenuation of the peripheral chemoreflex with consequent hypotension (**Figure 1**).

Effects of Cts based on *in vitro* and *in vivo* experimental studies are presented in **Figure 1**. In summary, Cts may inhibit CA release from chromaffin cells and noradrenergic neurons (11) and induces desensitization of CAs (57); it also exhibits a potent vasodilatory effect mediated – at least in part – by histamine release (48, 49). Moreover, negative inotropic and lusitropic effects of Cts may lower BP (53). Finally, Cts plays a role in central cardiorespiratory control (3, 56). Taken together, experimental studies point to Cts as a novel regulator of BP.

Clinical Studies

On the one hand, similarly to *in vivo* studies on rodents, it was shown that infusion of Cts into the dorsal hand veins of normotensive individuals, after pharmacologic venoconstriction with phenylephrine, resulted in a dose-dependent vasodilation, which was predominantly observed in female subjects (58). On the other, clinical studies generated controversies regarding the association between Cts levels and primary HT.

O'Connor et al. showed that normotensive offspring of patients with HT had significantly lower Cts concentrations compared to normotensive participants with a negative family history of HT (1.32 ± 0.038 vs. 1.5 ± 0.076 ng/mL, $p = 0.024$). Low plasma Cts levels predicted enhanced pressor response to a sympathoadrenal stressor (59). Therefore, reduced Cts levels were postulated to predispose to the development of HT, although categorization by BP status (normotensive vs. hypertensive) did not reveal differences in plasma Cts (1.36 ± 0.03 vs. 1.26 ± 0.06 ng/mL, $p = 0.27$) (59). In line with the result, Durakoglugil et al. showed that the difference in Cts concentrations between previously untreated hypertensive patients and healthy controls was insignificant after adjusting for age, gender, height, and weight (60). However, in another study by the O'Connor group, in a larger cohort (452 normotensives, 215 primary hypertensives), Cts was significantly reduced in HT patients (1.47 ± 0.06 vs. 1.26 ± 0.08 ; $p = 0.036$) (61). In contrast, Meng et al. showed that Cts was significantly higher in patients with essential HT than normal controls (1.19 ± 0.74 vs. 1.53 ± 0.72 ng/mL, $p < 0.01$) but found no correlation between Cts and the degree of HT (1.56 ± 0.59 vs. 1.42 ± 0.59 vs. 1.57 ± 0.76 ng/mL, $p > 0.05$) (62). They also suggested Cts level may serve as a prognostic factor for complications of HT, as patients with left ventricular hypertrophy had lower Cts-to-norepinephrine ratios than those without (3.63 ± 1.62 vs. 2.76 ± 0.86 , $p < 0.01$) (62). In order to explain the contradictory results concerning Cts levels in normo- and HT, it was hypothesized that Cts decreases in prehypertension, however, sympathetic nervous system activity increases as HT progresses, which contributes to compensatory Cts elevation. Consequently, HT develops when elevated Cts no longer inhibits CA oversecretion (62).

Cts genetic variants exert varying effects on BP (16). The Gly364Ser variant was associated with lower diastolic BP than the Gly364Gly variant (wild type) in two Caucasian hypertensive groups of European ancestry and ca. 1.8% variance of population diastolic BP was attributable to Cts single nucleotide polymorphism (63). Conversely, in Asian populations, Ser-364 allele carriers displayed elevated systolic (up to ≈ 8 mm Hg; $p = 0.004$) and diastolic (up to ≈ 6 mm Hg; $p = 0.001$) BP

(Indian) (31) as well as systolic BP, pulse pressure and arterial stiffness (Japanese) (64). In line with these, in a normotensive Indian population, Ser-364 allele carriers exhibited much lower plasma CA - by about 40% (consistent with its diminished nAChR desensitization-blocking effect *in vitro*) - than Gly364Gly carriers (30), yet, BP was similar in subjects with different alleles. These contradictory results underscore the necessity of genetic association studies for ethnically different populations.

Processing of CgA to Cts by endoproteolytic enzymes may be involved in the pathogenesis of HT. O'Connor et al. showed that CgA was increased by 117% ($p < 0.001$) in HT patients, whereas Cts reduced by 15% ($p = 0.036$), which suggests diminished conversion of CgA to Cts in HT (normotensives' CgA/Cts ratio of 4.5 ± 0.2 vs. 5.9 ± 0.4 in HT patients, $p = 0.005$) (61). Fung et al. demonstrated that in women compared to men higher plasma Cts (1.30 ± 0.033 vs. 1.14 ± 0.27 nM, $p < 0.001$) combined with lower CgA precursor concentrations (3.89 ± 0.15 vs. 4.65 ± 0.33 nM, $p = 0.006$) may be associated with decreased processing of CgA to Cts in the latter (female vs. male Cts/CgA ratio: 26.3 ± 0.006 vs. 23.1 ± 0.006), which predisposes to HT (58).

Moreover, Biswas et al. showed that CHGA variants undergo differential processing to produce Cts in the presence of the endoproteolytic enzyme plasmin (65). Their study indicates that less efficient processing of the Pro370Leu protein (product of one of CHGA variants) can contribute to the prevalence of CVDs (65).

CgA variants may also impact target organ damage in HT: black people suffering from end stage renal disease exhibit a common CHGA variant (3'-UTR C+87T), which decreased reporter gene expression and subsequently lead to lower Cts levels in this population (2.10 ± 0.88 vs. 3.23 ± 0.29 ng/mL, $p = 0.01$) (66).

Clinical studies concerning Cts in HT are presented in **Table 1**. To summarize, Cts was shown to decrease during prehypertension (59), however, the progression of elevated BP was associated with a compensatory increase in its concentration (62). Moreover, the development of HT may well be connected with diminished conversion of CgA to Cts (58, 61, 65) and depends on different variants of Cts exerting varying effects on BP (16, 63).

CARDIAC FUNCTION OF CATESTATIN

In vitro and *in vivo* Animal Studies

Cts induces NO synthesis from endothelial cells and cardiomyocytes (43, 53). In endothelial cells, NO is acquired from both: (1) the endothelin receptor B – endothelial nitric oxide synthase (eNOS) – NO, and (2) the protein kinase B (Akt) – eNOS/neuronal nitric oxide synthase (nNOS) – NO pathways. In cardiomyocytes, NO release results from the extracellular signal-regulated kinase – eNOS – NO pathway (53) (**Figure 1**). NO reduces inotropism and lusitropism of cardiomyocytes through various pathways (52, 53, 88, 89) (**Figure 1**). It was demonstrated *in vitro* using frog (52), eel (89), and rat (90) heart preparations that higher NO generation due to Cts ameliorates the Frank-Starling response, which is a compensatory mechanism to maintain adequate heart

TABLE 1 | Clinical studies concerning catestatin.

References	Study participants	Main results
Catestatin and hypertension		
O'Connor et al. (59)	40 normotensives with positive HT family history, 176 normotensives without HT family history, 61 patients with HT	Offspring of HT patients had lower Cts than normotensives without HT family history: 1.32 ± 0.038 vs. 1.5 ± 0.076 ($p = 0.024$) Plasma Cts was not different in normotensives vs. hypertensives (1.36 ± 0.03 vs. 1.26 ± 0.06 ($p = 0.27$))
O'Connor et al. (61)	452 normotensives, 215 patients with HT	Cts was reduced by 15% in hypertensives patients ($p = 0.036$), whereas CgA was increased by 117% ($p < 0.001$); the ratio of CgA/Cts was thus increased by 31% ($p = 0.005$), implying decreased conversion of CgA to Cts in PH.
Salem et al. (66)	Black patients with HT and ESRD ($n = 150$) and black controls ($n = 58$)	ESRD patients had lower Cts: 2.10 ± 0.88 vs. 3.23 ± 0.29 ($p = 0.01$)
Meng et al. (62)	136 HT patients (109 with and 27 without LVH 27) and 61 healthy controls	Cts was higher in hypertensives vs. controls: 1.19 ± 0.74 vs. 1.53 ± 0.72 ($p < 0.01$) There was a non-significant trend toward lower Cts in HT patients with LVH than those without: 1.55 ± 0.7 vs. 1.4 ± 0.53 ($p > 0.05$).
Catestatin and coronary artery disease		
Wang et al. (67)	50 STEMI patients and 25 non-CAD control patients	STEMI patients had lower plasma Cts on admission than controls: 16.5 ± 5.4 vs. 21.4 ± 6.4 ($p < 0.01$), increased on day 3 to 30.7 ± 12.2 ($p < 0.01$), and on day 7 decreased to levels below than admission (13.8 ± 5.3 , $p < 0.01$) in MI
Meng et al. (68)	52 healthy controls 58 STEMI patients, 31 of whom were assessed by echocardiography 3 months later to reveal 7 cases with LVR and 24 without LVR	Plasma Cts on admission higher in patients with AMI than in controls: 1.00 (0.66–1.50) vs. 0.84 (0.56–1.17) ($p < 0.05$) Mean Cts higher on day 3: 1.12 (0.76–1.70) vs. 1.00 (0.66–1.50) ($p < 0.01$) and on day 7: 1.32 (0.81–1.73) ng/ml vs. 1.00 (0.66–1.50) ($p < 0.01$) Patients with LVR 3 months after AMI vs. those without had higher Cts: -on admission: 2.02 (1.14–5.87) vs. 0.94 (0.39–1.66) ($p = 0.001$), -on day 3: 2.47 (1.10–5.95) vs. 1.16 (0.31–1.95) ($p = 0.006$), and, -on day 7 after STEMI: 3.08 (0.41–6.77) vs. 1.20 (0.30–3.73) ($p = 0.021$)
Liu et al. (69)	30 healthy controls; 15 SAP, 47 UAP, 22 NSTEMI, and 36 STEMI patients	Plasma Cts was higher in CAD patients than controls: 0.41 ± 0.14 vs. SAP patients: 0.72 ± 0.50 ($p < 0.05$) UAP patients 0.88 ± 0.58 ($p < 0.05$) NSTEMI patients 1.05 ± 0.48 ($p < 0.05$) STEMI patients 1.31 ± 0.91 ($p < 0.05$)
Pei et al. (70)	STEMI patients with MA ($n = 61$) STEMI patients without MA ($n = 64$)	Plasma Cts was higher in patients with STEMI complicated by MA compared with those without MA: 0.083 ± 0.011 vs. 0.076 ± 0.007 ($p < 0.001$)
Zhu et al. (71)	30 non-CAD controls ($n = 30$) 100 AMI patients including 74 with adverse events on follow-up and 26 without	Cts lower in MI patients on admission vs. controls: 16.7 ± 5.4 vs. 21.8 ± 6.3 ($p < 0.0001$), higher on day 3 at 30.9 ± 12.1 ($p < 0.0001$ vs. admission), and again lower on day 7: 13.9 ± 5.2 ($p = 0.0003$ vs. admission) Cts on admission and day 3 in the adverse events group (19.4 ± 6.7 ng/ml, 44.6 ± 13.0 , respectively) higher than in the non-adverse events group (15.8 ± 4.5 ($p = 0.003$), 26.1 ± 7.5 ($p < 0.0001$), respectively).
Xu et al. (72)	38 patients with CTO and 38 controls	Cts higher in CTO patients than in controls 1.97 ± 1.01 vs. 1.36 ± 0.97 ($p = 0.009$)
Zhu et al. (73)	72 STEMI patients and 30 control patients without CAD on imaging	Patients with Cts level above median at day 3 (28.71 ng/ml) developed worse ventricular function during the 65 months follow-up ($p < 0.0001$).
Xu et al. (74)	46 STEMI patients, 89, 35 control patients without CAD on imaging	Cts in patients with STEMI (0.80 ± 0.62) and UAP (0.99 ± 0.63) lower than in controls (1.38 ± 0.98 ; $p = 0.001$).
Kojima et al. (75)	25 CAD patients: 20 with AMI and 5 with UAP; controls: 20 non-CAD patients with mild hypertension and 13 healthy volunteers	Plasma Cts levels were lower in CAD patients (2^*) than in non-CAD patients (4^* ; $p \leq 0.05$).
Chen et al. (19)	204 healthy volunteers 224 CAD patients	CAD patients had lower serum Cts than controls: 1.14 (1.05–1.24) vs. 2.15 (1.92–2.39); $p < 0.001$, and the levels decreased in a stepwise manner with increasing number of diseased vessels: 1.95 (1.83–2.07) vs. 1.57 (1.42–1.73) vs. 1.13 (1.00–1.27), $p < 0.001$ (for 1, 2, and 3 vessels, respectively)
Catestatin and heart failure		
Zhu et al. (76)	300 moderate to severe HF patients: –108 in stage A; 76 in Stage, 116 - Stage C	Cts decreased with higher HF stages and there was a significant difference between stage A and B: 21.29 ± 7.10 vs. 14.61 ± 4.69 ($p < 0.05$).
Liu et al. (77)	172 controls 228 HF patients in NYHA class I – IV	Plasma Cts increased with higher classes, NYHA class III and class IV patients had higher Cts levels than controls: 0.848 (0.664–1.260); 1.54 (0.856–2.432), respectively vs. 0.696 (0.504–0.883) ($p < 0.05$). NYHA class I and class II patients had similar Cts to controls: 0.612 (0.52–0.844); 0.722 (0.532–1.112), respectively vs. 0.696 (0.504–0.883) ($p > 0.05$)

(Continued)

TABLE 1 | Continued

References	Study participants	Main results
Peng et al. (78)	Cohort of 202 HF patients followed-up for a median of 52.5 months: 143 survived, 59 died – 49 for cardiac causes	Plasma Cts was higher in non-survivors both for all and cardiac causes 1.06 (0.66–1.82) and 1.18 (0.69–1.83), respectively vs. 0.75 (0.58–1.12) in survivors ($p \leq 0.005$)
Wolowiec et al. (79)	Upon a follow-up of 24 months out of 52 HFrEF patients 11 reached the composite endpoint (CE) of unplanned hospitalization and all-cause death 24 healthy volunteers served as controls	Cts lower in HFrEF patients who reached a CE than in those who did not – both before and after exertion: 14.23 (11.05–15.82) vs. 16.86 (14.25–19.46) ($p = 0.03$) and 4.81 (2.20–6.25) vs. 7.82 (5.81–63.48) ($p = 0.002$), respectively; Cts in patients similar to controls: before exertion 15.95 (13.89–18.81) vs. 16.6 (14.75–22.20) ($p = 0.12$), after: 7.04 (4.97–11.08) vs. 9.26 (6.11–140.23) ($p = 0.13$)
Borovac et al. (80)	96 HF patients hospitalized due to an acute worsening of HF, 6 did not survive	Cts was significantly higher among non-survivors than survivors: 19.8 (9.9–28) vs. 5.6 (3.4–9.8) ($p < 0.001$)
Catestatin and other diseases affecting the cardiovascular system		
Sun et al. (81)	330 controls; 329 hemodialysis patients followed-up for 36 months – 29 died for cardiac and 28 for non-cardiac causes	Cts higher in hemodialysis patients (1.9 ± 0.3) vs. controls (1.2 ± 0.2), $p < 0.001$ Cts higher (2.2 ± 0.1) in patients who died for cardiac causes than in survivors (1.8 ± 0.2) and non-cardiac death non-survivors 1.8 ± 0.3 , $p < 0.001$
Izci et al. (82)	97 controls 160 APE patients: 72 with sPESI ≥ 1 and 88 < 1	Plasma Cts higher in APE patients than controls: 27.3 ± 5.7 vs. 17.5 ± 6.1 ($p < 0.001$); Cts higher in patients with sPESI ≥ 1 than with sPESI < 1: 37.3 ± 6.1 vs. 24.2 ± 5.3 ($p < 0.001$)
Tütün et al. (83)	100 women with preeclampsia 100 women with uncomplicated pregnancy as controls.	Plasma Cts was significantly increased in the preeclampsia patients compared to the controls: 0.29 ± 0.096 vs. 0.183 ± 0.072 ($p < 0.001$)
Liu et al. (84)	260 healthy workers	Plasma CgA-to-catestatin ratio correlated with effort, reward (negatively), overcommitment, and efford-reward imbalance: $r = 0.218, -0.249, 0.275$, and 0.279 , respectively, $p < 0.001$ for all
Catestatin and metabolic syndrome		
Simunovic et al. (85)	92 obese subjects (BMI Z score >2), age 10–18; 39 controls	Lower plasma Cts concentrations in obese subjects compared to controls: 10.03 ± 5.05 vs. 13.13 ± 6.25 ($p = 0.004$); lower Cts in the subgroup of obese patients with MS: 9.02 ± 4.3 vs. 10.54 ± 5.36 vs. 13.13 ± 6.25 ($p = 0.008$). Cts negatively correlated with DBP ($r = -0.253, p = 0.014$), HOMA-IR ($r = -0.215, p = 0.037$) and hsCRP ($r = -0.208, p = 0.044$).
Kim et al. (86)	85 subjects with mild OSA, 26 with moderate-to-severe OSA, 102 were controls, mean age 7.7 ± 1.4 years	Children with OSA have reduced plasma Cts levels (Log Cts in moderate-to-severe OSA: 0.12 ± 0.22 vs. mild OSA: 0.23 ± 0.20 vs. controls: 0.28 ± 0.19 ; differences among three groups: $p < 0.01$). Cts levels were inversely correlated with AHI ($r = -0.226, p < 0.01$) and with mean arterial BP level ($r = -0.184, p < 0.05$).
Borovac et al. (87)	78 OSA patients; 51 controls	Plasma Cts higher in OSA patients compared to controls: 2.9 ± 1.2 vs. 1.5 ± 1.1 ($p < 0.001$). In OSA patients Cts correlated with neck circumference ($r = 0.318, p < 0.001; \beta = 0.384, p < 0.001$) and HDL cholesterol ($r = -0.320, p < 0.001; \beta = -0.344, p < 0.001$).

Catestatin concentrations are given in ng/mL and the results are shown as median (interquartile range) or mean \pm standard deviation; AHA, American Heart Association; AMI, acute myocardial infarction; APE, Acute pulmonary embolism; AWHF, acute worsening of heart failure; BP, blood pressure; CAD, coronary artery disease; CD, Crohn's disease; CE, composite endpoint including unplanned hospitalization and death for all causes; CPET, Cardiopulmonary Exercise Testing (6-minut walk test); Cts, catestatin; CTO, chronic total occlusions; DBP, diastolic blood pressure; ERI, effort, reward imbalance; ESRD, end stage renal disease; HDL, high-density lipoprotein; HF, heart failure; HFrEF, Heart Failure with Reduced Ejection Fraction; HOMA-IR, homeostatic model assessment of insulin resistance; hsCRP, high sensitivity C-reactive protein; HT, hypertension; IBD, inflammatory bowel diseases; LVH, left ventricular hypertrophy; LVR, left ventricular remodeling; MA, malignant arrhythmia; MS, metabolic syndrome; NSTEMI, non-ST segment elevation myocardial infarction; NYHA, New York Heart Association; OSA, obstructive sleep apnea; SAP, stable angina pectoris; STEMI, acute ST- segment elevation myocardial infarction; sPESI, simplified PESI; UAP, unstable angina pectoris; UC, ulcerative colitis. *Mean value (standard deviation was not provided).

function in response to changes in venous return (90). Moreover, Cts exerts counterregulatory action against β -adrenergic and endothelin-1 stimulation pointing to Cts as a novel, beneficial cardiac modulator (53) (Figure 1).

In line with this research, *in vitro*, Cts attenuated norepinephrine-mediated hypertrophic responses in H9c2 cardiac myoblasts and at 10–25 nM signaling was moderated primarily by $\beta 1/2$ -adrenoceptors (91). Interestingly, Bassino et al. observed that a low (5 nM) WT-Cts concentration reduced β -adrenergic stimulation in bovine aortic endothelial cells, although it exhibited no significant effect on myocardial contractility. Higher concentrations (10–50 nM) of Cts induced

a transient positive inotropic effect followed by a negative antiadrenergic effect (43). The transient positive effect was probably related to histamine release from mast cells induced by Cts. Histamine exerts a positive inotropic effect on rat ventricular myocardium and H1 histamine receptor antagonist mepyramine blocked the positive effect induced by higher doses of WT-Cts (10 nM). In turn, WT-Cts at a minimal concentration (5 nM) reduced isoproterenol-induced enhancement of papillary muscle contractility via H1 receptor activation without altering basal contractile ability (43).

Several studies point at a cardioprotective role of Cts in ischemia/reperfusion (I/R) injury of the heart in rodent models.

It was apparent for Cts pretreatment (25 nM, 50 nM, and 100 nM infused for 15 min before ischemia) (92) and chronic administration of Cts after MI (0.25 mg/kg/12 h injection intraperitoneally 24 h after AMI for 28 days) (67). Further, infusion of a Cts dose of 75 nM for 20 min at the beginning of reperfusion significantly reduced infarct size, limited post-ischemic contracture, and improved recovery of developed left ventricular pressure (93–96). Protection due to Cts in I/R-injured myocardium was related to salvage of oxidative-stress-induced apoptosis by activation of the β 2-adrenergic receptor, and PKB/Akt pathway (96). It must be highlighted that infusion of a higher dose WT-Cts (100 nM) for 120 min during reperfusion caused deleterious effects and did not activate Akt (97). Interestingly, the Gly364Ser variant of Cts at a higher dose (100 nM for 120 min) lowered infarct size, which is probably associated with lower inhibitory activity on the nicotinic cholinergic receptor of the Cts-Gly364Ser variant (97). It is worth underlining that cardioprotection achieved by ischemic preconditioning in wild-type mice was absent in Cts-KOs (21).

Modulation of cardiac glucose metabolism is another cardioprotective mechanism of Cts, in particular, improvement of glucose uptake early during post-ischemic reperfusion (18). In physiological conditions, cardiomyocyte metabolism depends mainly on fatty acid oxidation, although the heart shifts toward glucose metabolism in response to ischemia to ameliorate myocardial contractile efficiency. In isolated rat cardiomyocytes, Cts induced Akt and AS160 phosphorylation and significantly enhanced glucose uptake (18), which may be crucial for recovery of contractile function during post-ischemic reperfusion (98).

A recent study involving an *in vivo* and *in vitro* approach investigated Cts in acute pulmonary embolism (99). Its administration in mice with this pathology increased survival as well as augmented thrombus resolution by attenuating endothelial inflammation (99).

In summary, based on *in vitro* and *in vivo* animal models, it was demonstrated that Cts exhibits a potential cardioprotective effect by acting directly as a cardiodepressing peptide through multiple signaling pathways (52, 53, 88–90), it may also reduce apoptosis of cardiomyocytes induced by oxidative stress (67, 92–96), and is beneficial in acute pulmonary embolism (99). However, more studies are needed in these areas, in particular, regarding Cts in I/R injury of the heart (67, 92–97).

Clinical Studies

Coronary Artery Disease

Wang et al. (100) were the first to investigate Cts in patients with coronary artery disease (CAD) by measuring its plasma concentration in 58 acute myocardial infarction (AMI) patients on admission, day 3 and day 7, and in 25 control subjects who were admitted to the same hospital for atypical chest pain but with normal coronary arteries confirmed by coronary angiography. Compared with controls (21.4 ± 6.4 ng/ml), plasma Cts concentrations were significantly lower on admission due to AMI (16.5 ± 5.4 ng/ml, $p < 0.01$), higher on the third day (30.7 ± 12.2 ng/ml, $p < 0.01$) and again lower on day 7 (13.8 ± 5.3 ng/ml, $p < 0.01$).

Zhu et al. (71) and Xu et al. (74) obtained results consistent with the above: compared to controls, Cts was lower on admission due to AMI, it increased significantly on the third day of hospitalization, but decreased 1 week following AMI. In line with these studies, Kojima et al. showed in small samples of patients that Cts was significantly lower in patients with AMI or unstable angina pectoris than in non-CAD patients (75).

However, Meng et al. (68) and Liu et al. (69) showed that plasma Cts levels at the time of admission were significantly higher in patients with AMI compared to healthy controls. The discrepancy between studies may result from different control groups, which included patients with chest pain without CAD on imaging in studies by Wang et al. (100), Zhu et al. (71) and Xu et al. (74) vs. healthy volunteers in studies by Meng et al. (68) and Liu et al. (69).

In addition, Chen et al. demonstrated serum Cts levels were lower in stable angina pectoris (SAP) patients compared to healthy controls (median (inter-quartile range) 1.14 (1.05–1.24) ng/mL vs. 2.15 (1.92–2.39) ng/mL, $p < 0.001$). Moreover, a stepwise decrease in serum Cts was found when classifying CAD patients according to the number of diseased vessels (19). However, Liu et al. showed that SAP patients ($n = 15$) had significantly higher Cts levels compared to controls (0.41 ± 0.14 ng/mL vs. 0.72 ± 0.50 ng/mL, $p < 0.05$) (69). Divergent findings are difficult to account for, perhaps these depend on disease symptoms in patients of the latter study (Cts increases as CAs are released when pain occurs); small SAP patient sample size in the study by Liu et al. could have biased the results (69).

Further, Xu et al. demonstrated that mean plasma Cts in coronary artery chronic total occlusion patients, who underwent coronary angiography or percutaneous coronary intervention for the first time, were significantly higher than in patients with chest pain but with normal coronary arteries (1.97 ± 1.01 vs. 1.36 ± 0.97 ng/ml, $p = 0.009$) (72).

Of the 58 AMI patients mentioned above, Meng et al. studied 31 by echocardiography 3 months after AMI onset, and diagnosed 7 with left ventricular remodeling (LVR). Interestingly, these patients had significantly higher plasma Cts levels on admission, day 3, and day 7 than those without LVR (68). In line with Meng et al., Dan Zhu et al. found that AMI patients, whose Cts level exceeded the median at day 3 of hospitalization (28.71 ng/ml), exhibited worse left ventricular function in echocardiography 65 months after AMI (73).

In another study performed by Dan Zhu et al., adverse events occurred more frequently in the 65-month follow-up in patients whose Cts level on day 3 exceeded the median as well as in those whose ratios of Cts on day 7 to day 3 were below the median (71). Moreover, plasma Cts level turned out to be an independent prognostic factor for malignant arrhythmia (MA) after AMI, and was significantly higher in patients with AMI complicated by MA (70). However, Xu et al. showed no significant differences in major adverse cardiovascular events, including death from cardiovascular causes, recurrent AMI, or hospital admission for HF, or revascularization between patients with high and low Cts concentrations (72).

Heart Failure

Concerning HF, Dan Zhu et al. showed that the higher the severity stage of HF (according to the American Heart Association, AHA), the lower the Cts level, and that Cts might be a better predictive factor for stage B HF than brain natriuretic peptide, a marker commonly used in clinical practice for HF diagnosis and severity assessment (76). However, according to a study by Liu et al., plasma Cts levels were increasingly higher in patients with growing severity of the New York Heart Association (NYHA) HF classes I to IV, and NYHA class III and IV patients exhibited significantly higher plasma Cts levels than controls, NYHA class I, and class II subjects (77). Seemingly contradictory results may be a consequence of patient enrollment criteria: Zhu et al. recruited asymptomatic chronic patients with stage C HF (76), while NYHA IV patients had resting dyspnea (77). As mentioned before, symptoms that trigger CA release can cause a compensatory increase in Cts levels.

Peng et al. demonstrated that Cts was an independent risk factor for all-cause death [hazard ratio 1.84 (95% CI: 1.02–3.32, $p = 0.042$)] and cardiac death [hazard ratio 2.41 (95% CI: 1.26–4.62, $p = 0.008$)] during a median 52.5-month follow-up (78). In line with their results, in the CATSTAT-HF Study, Cts was found to be an independent and significant predictor of in-hospital death, and its level was significantly higher among non-survivors than survivors (80). However, in the study by Wołowiec et al., patients who reached the composite endpoint of unplanned hospitalization and death for all causes during a 24-month follow-up had lower Cts levels – assessed both before and after physical exertion. Firth coefficient was 6.58 (penalized 95% CI 1.66–21.78, $p = 0.003$) (79). Again, the divergent data could result from different study populations: Wołowiec et al. enrolled only hemodynamically stable patients (79), while in the CATSTAT-HF Study patients with acute decompensation of chronic HF were included (80), and more than half of the patients (55.9%) in the study by Peng et al. study were classified as NYHA III-IV (78).

Catestatin in Other Diseases Affecting the Cardiovascular System

In the scope of CVD, apart from studies on the role of Cts in HT, CAD, and HF, few other have been published.

Sun et al. studied hemodialyzed patients and demonstrated that plasma Cts levels equal to or greater than the mean (1.9 ng/ml) were associated with higher cardiac death risk (RR 6.13, 95% CI 2.54, 18.45) during a 36-month follow-up. Moreover, there was no such association for non-cardiac death (RR 1.29, 95% CI 0.70, 2.85) (81).

Izci et al. showed that plasma Cts levels were higher in patients with acute pulmonary embolism than in control subjects. Also, there was a positive correlation between Cts and right ventricular dysfunction, and between Cts and Simplified Pulmonary Embolism Severity Index (± 0.581 , $p < 0.001$), a score used to estimate 30-day mortality in patients diagnosed with non-high-risk acute pulmonary embolism. Furthermore, a Cts cut-off level of 31.2 ng/ml predicted mortality with a sensitivity of 100% and specificity of 52.6% (AUC = 0.883, 95% CI: 0.689–0.921) (82).

In preeclampsia patients, plasma Cts was significantly elevated compared to controls: 0.29 ± 0.096 vs. 0.183 ± 0.072 ($p < 0.001$) (83), and correlated positively with systolic and diastolic BP, urea, creatinine and uric acid levels (83).

Stress may account for at least in part-, the extent of sympathetic nervous activation, deleteriously affects the immune and coagulation systems, increasing the risk of CVDs. Interestingly, it was demonstrated recently that plasma CgA correlated positively with effort, overcommitment, and effort-reward imbalance ($r = 0.267$, 0.319, and 0.304, respectively, $p < 0.001$ for all three), and negatively with reward ($r = -0.237$, $p < 0.001$). Plasma CgA-to-Cts ratio was also associated with work stress in a manner similar to CgA (84).

Summary of Clinical Studies

Clinical studies regarding Cts in CAD and HF are summarized in **Table 1**. It must be highlighted that CAs release causes an increase in Cts, which changes dynamically. Low levels of Cts may play a pathogenic role in cardiac ischemia (74). Research indicates that Cts is involved in the course of CAD as well as HF, and, possibly, its concentration may be applied in monitoring.

ROLE OF CATESTATIN IN METABOLIC DISORDERS AND ATHEROSCLEROSIS

Metabolic disorders such as obesity, insulin resistance and type 2 diabetes are associated with CVDs. Obesity with an abnormal lipid profile may lead to insulin resistance and is associated with higher cardiovascular risk. In turn, insulin resistance correlates strongly with cardiovascular pathology and is a powerful predictor of future development of type 2 diabetes, an independent risk factor for CVDs (101).

Dysregulated immune system is involved in the pathogenesis of these widely prevalent metabolic disorders. The interdependence of adverse systemic metabolic conditions and immune responses gave rise to the term “immunometabolism,” which is currently also used to describe pathologic reprogramming of immune cells not only in the spectrum of metabolic syndrome, but also other diseases (e.g., neoplasms and autoimmunity). Here, the former meaning of immunometabolism was adopted.

In vitro and *in vivo* Animal Studies

Based on *in vitro* and *in vivo* studies with rodent models, Cts plays a role in the crosstalk between the immune and metabolic systems (15), in particular, in the development of atherosclerosis (19). Cts may act as an anti-atherogenic and anti-inflammatory peptide that reduces leukocyte-endothelium interaction by activating angiotensin-converting enzyme-2 and suppressing tumor necrosis factor- α -elicited expression of inflammatory cytokines and adhesion molecules (19); development of atherosclerosis was attenuated by Cts treatment in apolipoprotein E knockout mice fed a high-fat diet (a mouse model of atherosclerosis) (19) (**Figure 1**). Further, after vascular injury, Cts increased the expression of *Mrc1*, a gene encoding an anti-inflammatory peptide, and

prevented macrophage-driven atherosclerosis (75). Presumably, anti-inflammatory actions of Cts partly depend on the regulation of chemotaxis (102). *In vitro* and *in vivo* (rodent models), Cts counteracted chemoattraction of monocytes and neutrophils by inflammatory chemokines (102), likely contributing to reduced immune infiltration in the heart (21, 75). Regulation of chemotaxis by Cts is complex and naturally occurring Cts variants differ in their chemotactic properties (103, 104), which may influence the propensity for CVD.

The liver plays both a critical role in metabolism, and serves as an important site of immune regulation as it contains resident immune cells as well as synthesizes a number of inflammatory proteins (105). Both tissue-resident (Kupffer cells) and recruited macrophages contribute to an inflammatory state of the liver, e.g., in obesity-induced insulin resistance and type 2 diabetes. Using a rodent model, Ying et al. showed that Cts may inhibit the function and infiltration of macrophages in the liver, which suppresses hepatic gluconeogenesis and improves insulin sensitivity (20). Moreover, the authors demonstrated that treatment of diet-induced obese mice with Cts elicited beneficial changes, including decreased plasma lipids and insulin as well as hepatic lipid content, attenuated expression of gluconeogenic and proinflammatory genes, and increased expression of anti-inflammatory genes in both Kupffer cells and recruited monocyte-derived macrophages in the liver (20) (**Figure 1**). Furter *in vivo* research on both diet-induced obese and normal chow diet mice showed that Cts decreased obesity-induced endoplasmic reticulum dilation in hepatocytes and macrophages, and enhanced insulin sensitivity in mammalian cells (106).

Latest research *in vivo* on a rodent Cts-KO model has shown that Cts directly promotes hepatic glycogen synthesis, reduces gluconeogenesis and glycogenolysis, as well as enhances downstream insulin signaling (23). Positive effects of Cts were observed in Chga-KO mice too, which – despite regular chow diet – were obese, and had increased plasma CAs, leptin, adiponectin, and ketone bodies at baseline (22). Moreover, compared to WT controls, Chga-KO mice exhibited higher glucose-stimulated insulin secretion, which was likely caused by changes in secretory vesicles and mitochondria observed in CgA-deficient β -cells (107). In this model, treatment with Cts lowered circulating CAs and leptin as well as reduced adipose tissue by about 25% resulting in a lean phenotype, increased lipolysis, enhanced fatty acid oxidation and assimilation into lipids in the liver (**Figure 1**). These beneficial metabolic effects of exogenous Cts were presumed to result from reduction of CA and adiponectin resistance; the effect was mediated by the inhibition of α 2 adrenergic signaling (22) (**Figure 1**). In addition, Cts improved leptin signaling (determined by phosphorylation of AMPK and Stat3 in Chga-KO mice) and peripheral leptin sensitivity in both diet-induced obese mice and in leptin-deficient *ob/ob* mice (22) (**Figure 1**). In another model with genetically modified rodents, Cts was found to restore sodium-glucose transporter 1 (SGLT1) expression and abundance as well as intestinal turnover in double knock-out leptin receptor b mice, an experimental model of

obesity, type 2 diabetes with hyperleptinemia. The effect was possibly mediated by antagonistic binding of Cts to the leptin receptor a (108).

To sum up, it is increasingly clear that Cts is vital for maintaining metabolic homeostasis, which is partly achieved by affecting the immune system. Based on *in vitro* and *in vivo* experimental studies, Cts may inhibit inflammatory response and leukocyte-endothelial cell interactions (19, 21, 75, 102, 103), prevent macrophage-driven atherosclerosis (19, 75), regulate monocyte migration (103), and cytokine production and release (102–104). As a novel regulator of metabolism, in rodents, Cts was shown to help in achieving insulin sensitivity, overcoming endoplasmic reticulum stress (106), reducing adipose tissue by increasing lipolysis, enhancing oxidation of fatty acids, and their assimilation into lipids (22).

Clinical Studies

So far, only few studies have been conducted in humans on the metabolic role of Cts. Clearly, more research is required in this area.

In a study reported above, O'Connor et al. showed that plasma Cts levels correlated negatively with BMI and plasma leptin concentrations in hypertensive and normotensive subjects (59). Durakoglugil et al. reported that plasma Cts was an independent predictor of high-density lipoprotein cholesterol, and correlated inversely with plasma triglycerides (60). Interestingly, the Ser-364 allele was strongly associated with elevated plasma triglyceride and glucose levels (30).

Simunovic et al. compared plasma Cts levels in 92 obese children and adolescent with those of 39 healthy (normal weight) controls: it levels were significantly lower in the former (10.03 ± 5.05 vs. 13.13 ± 6.25 ng/mL, $p = 0.004$) (85), and, in addition, lower in the subgroup of obese patients with metabolic syndrome vs. those without and controls (9.02 ± 4.3 , 10.54 ± 5.36 , and 13.13 ± 6.25 ng/ml, respectively, $p = 0.008$). Moreover, Cts negatively correlated with diastolic BP ($r = -0.253$, $p = 0.014$), homeostatic model assessment of insulin resistance index ($r = -0.215$, $p = 0.037$), and high sensitivity C-reactive protein ($r = -0.208$, $p = 0.044$) (85) (**Table 1**). In another study, children with obstructive sleep apnea (OSA) had reduced plasma Cts levels (Log Cts in moderate-to-severe OSA: 0.12 ± 0.22 vs. mild OSA: 0.23 ± 0.20 vs. controls: 0.28 ± 0.19 ; $p < 0.01$) and Cts correlated negatively with apnea-hypopnea index ($r = -0.226$; $p < 0.01$) as well as mean arterial BP ($r = -0.184$; $p < 0.05$) (86) (**Table 1**).

Borovac et al. measured plasma Cts levels in 78 male OSA patients aged 50.3 ± 8.8 years and 51 age-, sex- and BMI-matched control subjects. They demonstrated that Cts serum levels are higher in the former (2.9 ± 1.2 vs. 1.5 ± 1.1 ng/mL, $p < 0.001$) (87). Cts significantly correlated with neck circumference ($r = 0.318$, $p < 0.001$; $\beta = 0.384$, $p < 0.001$) and high-density lipoprotein cholesterol ($r = -0.320$, $p < 0.001$; $\beta = -0.344$, $p < 0.001$), as well as apnea-hypopnea index among non-obese obstructive sleep apnea subjects ($r = 0.466$, $p = 0.016$; $\beta = 0.448$, $p = 0.026$) (87) (**Table 1**).

Evidently, reduced Cts levels seem to be associated with an adverse metabolic profile, including obesity and metabolic syndrome, abnormal lipid concentrations and insulin resistance.

CONCLUSIONS AND PERSPECTIVES

In conclusion, current data indicate that the endogenous bioactive peptide Cts is a vital regulatory factor of cardiovascular and immunometabolic homeostasis. Possibly, in future, Cts can be used in the diagnosis of CVDs and metabolic disorders as a novel biomarker, which may aid in clinical decision-making. Application of Cts in therapy might potentially mitigate detrimental sympathoexcitatory effects, which underlie cardiovascular and metabolic diseases. It should be highlighted

that, so far, studies on Cts have been carried out mainly on animal models. More research is required to take advantage of beneficial effects of Cts in clinical practice.

AUTHOR CONTRIBUTIONS

EZ and PK review the literature, and EZ wrote the first draft of the manuscript. PK and EZ wrote sections of the manuscript. KS carried out critical interpretations. All authors contributed to manuscript revision, read, and approved the submitted version.

REFERENCES

- Gerberding JL. Measuring pandemic impact: vital signs from vital statistics. *Ann Intern Med.* (2020) 173:1022–3. doi: 10.7326/M20-6348
- Valensi P. Autonomic nervous system activity changes in patients with hypertension and overweight: role and therapeutic implications. *Cardiovasc Diabetol.* (2021) 20:1–12. doi: 10.1186/s12933-021-01356-w
- Gaede AH, Pilowsky PM. Catestatin in rat RVLM is sympathoexcitatory, increases barosensitivity, and attenuates chemosensitivity and the somatosympathetic reflex. *Am J Physiol Regul Integr Comp Physiol.* (2010) 299:1538–45. doi: 10.1152/ajpregu.00335.2010
- Callingham BA. The chromaffin cell. *Nature.* (1965) 208:6. doi: 10.1038/208006a0
- Motiejunaite J, Amar L, Vidal-Petiot E. Adrenergic receptors and cardiovascular effects of catecholamines. *Ann Endocrinol (Paris).* (2021) 82:193–7. doi: 10.1016/j.ando.2020.03.012
- Banks P, Helle K. The release of protein from the stimulated adrenal medulla. *Biochem J.* (1965) 97:40–1. doi: 10.1042/bj0970040C
- Blaschko H, Comline RS, Schneider FH, Silver M, Smith AD. Secretion of a chromaffin granule protein, chromogranin, from the adrenal gland after splanchnic stimulation. *Nature.* (1967) 215:58–9. doi: 10.1038/215058a0
- Schneider FH, Smith AD, Winkler H. Secretion from the adrenal medulla: biochemical evidence for exocytosis. *Br J Pharmacol Chemother.* (1967) 31:94–104. doi: 10.1111/j.1476-5381.1967.tb01980.x
- Pasqua T, Angelone T, Spena A, Cerra MC. Biological roles of the ectopic chromogranin-a-derived peptide catestatin. *Curr Med Chem.* (2017) 24:3356–72. doi: 10.2174/0929867324666170616104759
- Simon JP, Bader MF, Aunis D. Secretion from chromaffin cells is controlled by chromogranin A-derived peptides. *Proc Natl Acad Sci U S A.* (1988) 85:1712–6. doi: 10.1073/pnas.85.5.1712
- Mahata SK, O'Connor DT, Mahata M, Yoo SH, Taupenot L, Wu H, et al. Novel autocrine feedback control of catecholamine release: a discrete chromogranin A fragment is a noncompetitive nicotinic cholinergic antagonist. *J Clin Invest.* (1997) 100:1623–33. doi: 10.1172/JCI119686
- Biswas N, Gayen J, Mahata M, Su Y, Mahata SK, O'Connor DT. Novel peptide isomer strategy for stable inhibition of catecholamine release: application to hypertension. *Hypertension.* (2012) 60:1552–9. doi: 10.1161/HYPERTENSIONAHA.112.202127
- Sahu BS, Mahata S, Bandyopadhyay K, Mahata M, Avolio E, Pasqua T, et al. Catestatin regulates vesicular quanta through modulation of cholinergic and peptidergic (PACAPergic) stimulation in PC12 cells. *Cell Tissue Res.* (2019) 376:51–70. doi: 10.1007/s00441-018-2956-1
- Bozic J, Kumric M, Ticinovic Kurir T, Urlic H, Martinovic D, Vilovic M, et al. Catestatin as a biomarker of cardiovascular diseases: a clinical perspective. *Biomedicines.* (2021) 9:1757. doi: 10.3390/biomedicines9121757
- Mahata SK, Corti A. Chromogranin a and its fragments in cardiovascular, immunometabolic, and cancer regulation. *Ann N Y Acad Sci.* (2019) 1455:34–58. doi: 10.1111/nyas.14249
- Zhao Y, Zhu D. Potential applications of catestatin in cardiovascular diseases. *Biomark Med.* (2016) 10:877–88. doi: 10.2217/bmm-2016-0086
- Pasqua T, Rocca C, Spena A, Angelone T, Cerra MC. Modulation of the coronary tone in the expanding scenario of Chromogranin-A and its derived peptides. *Fut Med Chem.* (2019) 11:1501–11. doi: 10.4155/fmc-2018-0585
- Gallo MP, Femmino S, Antoniotti S, Querio G, Alloatti G, Levi R. Catestatin induces glucose uptake and GLUT4 trafficking in adult rat cardiomyocytes. *Biomed Res Int.* (2018) 2018:1–7. doi: 10.1155/2018/2086109
- Chen Y, Wang X, Yang C, Su X, Yang W, Dai Y, et al. Decreased circulating catestatin levels are associated with coronary artery disease: The emerging anti-inflammatory role. *Atherosclerosis.* (2019) 281:78–88. doi: 10.1016/j.atherosclerosis.2018.12.025
- Ying W, Mahata S, Bandyopadhyay GK, Zhou Z, Wollam J, Vu J, et al. Catestatin inhibits obesity-induced macrophage infiltration and inflammation in the liver and suppresses hepatic glucose production, leading to improved insulin sensitivity. *Diabetes.* (2018) 67:841–8. doi: 10.2337/db17-0788
- Ying W, Tang K, Avolio E, Schilling JM, Pasqua T, Liu MA, et al. The immunosuppression of macrophages underlies the cardioprotective effects of catestatin (CST). *bioRxiv.* (2021). doi: 10.1101/2020.05.12.092254
- Bandyopadhyay GK, Vu CU, Gentile S, Lee H, Biswas N, Chi NW, et al. Catestatin (Chromogranin A352-372) and novel effects on mobilization of fat from adipose tissue through regulation of adrenergic and leptin signaling. *J Biol Chem.* (2012) 287:23141–51. doi: 10.1074/jbc.M111.335877
- Bandyopadhyay G, Tang K, Webster NJG, van den Bogaart G, Mahata SK. Catestatin induces glycogenesis by stimulating the phosphoinositide 3-kinase-AKT pathway. *Acta Physiol (Oxf).* (2022) 235:e13775. doi: 10.1111/apha.13775
- Zhang D, Shooshtarizadeh P, Laventie BJ, Colin DA, Chich JE, Vidic J, et al. Two chromogranin a-derived peptides induce calcium entry in human neutrophils by calmodulin-regulated calcium independent phospholipase A2. *PLoS ONE.* (2009) 4:e4501. doi: 10.1371/journal.pone.0004501
- Muntjewerff EM, Dunkel G, Nicolaisen MJT, Mahata SK, Van Den Bogaart G. Catestatin as a target for treatment of inflammatory diseases. *Front Immunol.* (2018) 9:1–10. doi: 10.3389/fimmu.2018.02199
- Bourebara Y, Mularczyk M, Marycz K, Bourebaba L. Catestatin peptide of chromogranin A as a potential new target for several risk factors management in the course of metabolic syndrome. *Biomed Pharmacother.* (2021) 134:111113. doi: 10.1016/j.bioph.2020.111113
- Mahata SK, Kiranmayi M, Mahapatra NR. Catestatin: a master regulator of cardiovascular functions. *Curr Med Chem.* (2018) 25:1352–74. doi: 10.2174/0929867324666170425100416
- Taylor C V, Taupenot L, Mahata SK, Mahata M, Wu H, Yasothornsriku S, et al. Formation of the catecholamine release-inhibitory peptide catestatin from chromogranin A: Determination of proteolytic cleavage sites in hormone storage granules. *J Biol Chem.* (2000) 275:22905–15. doi: 10.1074/jbc.M001232200
- Wen G, Mahata SK, Cadman P, Mahata M, Ghosh S, Mahapatra NR, et al. Both rare and common polymorphisms contribute functional variation at CHGA, a regulator of catecholamine physiology. *Am J Hum Genet.* (2004) 74:197–207. doi: 10.1086/381399
- Sahu BS, Obbineni JM, Sahu G, Allu PKR, Subramanian L, Sonawane PJ, et al. Functional genetic variants of the catecholamine-release-inhibitory peptide catestatin in an Indian population: allele-specific effects on metabolic traits. *J Biol Chem.* (2012) 287:43840–52. doi: 10.1074/jbc.M112.407916
- Kiranmayi M, Chirasani VR, Allu PKR, Subramanian L, Martelli EE, Sahu BS, et al. Catestatin Gly364Ser variant alters systemic blood pressure and the risk for hypertension in human populations

- via endothelial nitric oxide pathway. *Hypertension*. (2016) 68:334–47. doi: 10.1161/HYPERTENSIONAHA.116.06568
32. Benyamin B, Maihofer AX, Schork AJ, Hamilton BA, Rao F, Schmid-Schönbein GW, et al. Identification of novel loci affecting circulating chromogranins and related peptides. *Hum Mol Genet*. (2017) 26:233–42. doi: 10.1093/hmg/ddw380
 33. Biswas N, Curello E, O'Connor DT, Mahata SK. Chromogranin/secretogranin proteins in murine heart: Myocardial production of chromogranin A fragment catestatin (Chga364–384). *Cell Tissue Res*. (2010) 342:353–61. doi: 10.1007/st00441-010-1059-4
 34. Mahapatra NR, Mahata M, Mahata SK, O'Connor DT. The chromogranin A fragment catestatin: Specificity, potency and mechanism to inhibit exocytotic secretion of multiple catecholamine storage vesicle co-transmitters. *J Hypertens*. (2006) 24:895–904. doi: 10.1097/01.hjh.0000222760.99852.e0
 35. Sugawara M, Resende JM, Marquette A, Chich J, Bechinger B. Membrane structure and interactions of human catestatin by multidimensional solution and solid-state NMR spectroscopy. *FASEB J*. (2010) 24:1737–46. doi: 10.1096/fj.09-142554
 36. Pasqua T, Corti A, Gentile S, Pochini L, Bianco M, Metz-Boutigue MH, et al. Full-length human chromogranin-A cardioactivity: myocardial, coronary, and stimulus-induced processing evidence in normotensive and hypertensive male rat hearts. *Endocrinology*. (2013) 154:3353–65. doi: 10.1210/en.2012-2210
 37. Rocca C, Grande F, Granieri MC, Colombo B, De Bartolo A, Giordano F, et al. The chromogranin A1-373 fragment reveals how a single change in the protein sequence exerts strong cardioregulatory effects by engaging neuropilin-1. *Acta Physiol*. (2021) 231:e13570. doi: 10.1111/apha.13570
 38. Taupenot L, Mahata SK, Mahata M, Parmer RJ, O'Connor DT. Interaction of the catecholamine release-inhibitory peptide catestatin (human chromogranin A352–372) with the chromaffin cell surface and Torpedo electrophax: Implications for nicotinic cholinergic antagonism. *Regul Pept*. (2000) 95:9–17. doi: 10.1016/S0167-0115(00)00135-X
 39. Herrero CJ, Alés E, Pintado AJ, López MG, García-Palomero E, Mahata SK, et al. Modulatory mechanism of the endogenous peptide catestatin on neuronal nicotinic acetylcholine receptors and exocytosis. *J Neurosci*. (2002) 22:377–88. doi: 10.1523/JNEUROSCI.22-02-00377.2002
 40. Mahata SK, Mahapatra NR, Mahata M, Wang TC, Kennedy BP, Ziegler MG, et al. Catecholamine secretory vesicle stimulus-transcription coupling *in vivo*. Demonstration by a novel transgenic promoter/photoprotein reporter and inhibition of secretion and transcription by the chromogranin A fragment catestatin. *J Biol Chem*. (2003) 278:32058–67. doi: 10.1074/jbc.M305545200
 41. Mahata SK, Mahata M, Wen G, Wong WB, Mahapatra NR, Hamilton BA, et al. The catecholamine release-inhibitory “catestatin” fragment of chromogranin A: Naturally occurring human variants with different potencies for multiple chromaffin cell nicotinic cholinergic responses. *Mol Pharmacol*. (2004) 66:1180–91. doi: 10.1124/mol.104.002139
 42. Sahu BS, Mohan J, Sahu G, Singh PK, Sonawane PJ, Sasi BK, et al. Molecular interactions of the physiological anti-hypertensive peptide catestatin with the neuronal nicotinic acetylcholine receptor. *J Cell Sci*. (2012) 125:2787. doi: 10.1242/jcs.114389
 43. Bassino E, Fornero S, Gallo MP, Ramella R, Mahata SK, Tota B, et al. A novel catestatin-induced antiadrenergic mechanism triggered by the endothelial PI3KeNOS pathway in the myocardium. *Cardiovasc Res*. (2011) 91:617–24. doi: 10.1093/cvr/cvr129
 44. Mahapatra NR, O'Connor DT, Vaingankar SM, Sinha Hikim AP, Mahata M, Ray S, et al. Hypertension from targeted ablation of chromogranin A can be rescued by the human ortholog. *J Clin Invest*. (2005) 115:1942–52. doi: 10.1172/JCI24354
 45. Gayen JR, Gu Y, O'Connor DT, Mahata SK. Global disturbances in autonomic function yield cardiovascular instability and hypertension in the chromogranin A null mouse. *Endocrinology*. (2009) 150:5027–35. doi: 10.1210/en.2009-0429
 46. Gayen JR, Zhang K, RamachandraRao SP, Mahata M, Chen Y, Kim HS, et al. Role of reactive oxygen species in hyperadrenergic hypertension: biochemical, physiological, and pharmacological evidence from targeted ablation of the chromogranin A (Chga) gene. *Circ Cardiovasc Genet*. (2010) 3:414–25. doi: 10.1161/CIRCGENETICS.109.924050
 47. Dev NB, Mir SA, Gayen JR, Siddiqui JA, Mustapic M, Vaingankar SM. Cardiac electrical activity in a genetically “humanized” chromogranin A monogenic mouse model with hyperadrenergic hypertension. *J Cardiovasc Transl Res*. (2014) 7:483–93. doi: 10.1007/s12265-014-9563-7
 48. Kennedy BP, Mahata SK, O'Connor DT, Ziegler MG. Mechanism of cardiovascular actions of the chromogranin A fragment catestatin *in vivo*. *Peptides*. (1998) 19:1241–8. doi: 10.1016/S0196-9781(98)00086-2
 49. Krüger PG, Mahata SK, Helle KB. Catestatin (CgA344–364) stimulates rat mast cell release of histamine in a manner comparable to mastoparan and other cationic charged neuropeptides. *Regul Pept*. (2003) 114:29–35. doi: 10.1016/S0167-0115(03)00069-7
 50. Jianqiang G, Li S, Guo J. Effect of catecholamine release-inhibitory peptide catestatin on sympathetic activity of hypertension. *J Am Coll Cardiol*. (2016) 68:C33–4. doi: 10.1016/j.jacc.2016.07.123
 51. Liu R, Sun NL, Yang SN, Guo JQ. Catestatin could ameliorate proliferating changes of target organs in spontaneously hypertensive rats. *Chin Med J (Engl)*. (2013) 126:2157–62. doi: 10.3760/cma.j.issn.0366-6999.20120757
 52. Mazza R, Pasqua T, Gattuso A. Cardiac heterometric response: The interplay between catestatin and nitric oxide deciphered by the frog heart. *Nitric Oxide*. (2012) 27:40–9. doi: 10.1016/j.niox.2012.04.003
 53. Angelone T, Quintieri AM, Brat BK, Limchaiyawat PT, Tota B, Mahata SK, et al. The antihypertensive chromogranin A peptide catestatin acts as a novel endocrine/paracrine modulator of cardiac inotropism and lusitropism. *Endocrinology*. (2008) 149:4780–93. doi: 10.1210/en.2008-0318
 54. Gaede AH, Lung MSY, Pilowsky PM. Catestatin attenuates the effects of intrathecal nicotine and isoproterenol. *Brain Res*. (2009) 1305:86–95. doi: 10.1016/j.brainres.2009.09.088
 55. Gaede AH, Pilowsky PM. Catestatin, a chromogranin A-derived peptide, is sympathoinhibitory and attenuates sympathetic barosensitivity and the chemoreflex in rat CVLM. *Am J Physiol Regul Integr Comp Physiol*. (2012) 302:365–72. doi: 10.1152/ajpregu.00409.2011
 56. Avolio E, Mahata SK, Mantuano E, Mele M, Alò R, Facciolo RM, et al. Antihypertensive and neuroprotective effects of catestatin in spontaneously hypertensive rats: Interaction with GABAergic transmission in amygdala and brainstem. *Neuroscience*. (2014) 270:48–57. doi: 10.1016/j.neuroscience.2014.04.001
 57. Mahata SK, Mahata M, Parmer RJ, O'Connor DT. Desensitization of Catecholamine Release. *J Biol Chem*. (1999) 274:2920–8. doi: 10.1074/jbc.274.5.2920
 58. Fung MM, Salem RM, Mehtani P, Thomas B, Lu CF, Perez B, et al. Direct vasoactive effects of the chromogranin A (CHGA) peptide catestatin in humans *in vivo*. *Clin Exp Hypertens*. (2011) 32:278–87. doi: 10.3109/10641960903265246
 59. O'Connor DT, Kailasam MT, Kennedy BP, Ziegler MG, Yanaihara N, Parmer RJ. Early decline in the catecholamine release-inhibitory peptide catestatin in humans at genetic risk of hypertension. *J Hypertens*. (2002) 20:1335–45. doi: 10.1097/00004872-200207000-00020
 60. Durakoglugil ME, Ayaz T, Kocaman SA, Kirbaş A, Durakoglugil T, Erdogan T, et al. The relationship of plasma catestatin concentrations with metabolic and vascular parameters in untreated hypertensive patients: Influence on high-density lipoprotein cholesterol. *Anatol J Cardiol*. (2015) 15:577–85. doi: 10.5152/akd.2014.5536
 61. O'Connor DT, Zhu G, Rao F, Taupenot L, Fung MM, Das M, et al. Heritability and genome-wide linkage in US and Australian twins identify novel genomic regions controlling chromogranin a: implications for secretion and blood pressure. *Circulation*. (2008) 118:247–57. doi: 10.1161/CIRCULATIONAHA.107.709105
 62. Meng L, Ye XJ, Ding WH, Yang Y, Di BB, Liu L, et al. Plasma catecholamine release-inhibitory peptide catestatin in patients with essential hypertension. *J Cardiovasc Med*. (2011) 12:643–7. doi: 10.2459/JCM.0b013e328346c142
 63. Rao F, Wen G, Gayen JR, Das M, Vaingankar SM, Rana BK, et al. Catecholamine release-inhibitory peptide catestatin (chromogranin A352–372): Naturally occurring amino acid variant Gly364Ser causes profound changes in human autonomic activity and alters risk for hypertension. *Circulation*. (2007) 115:2271–81. doi: 10.1161/CIRCULATIONAHA.106.628859
 64. Choi Y, Miura M, Nakata Y, Sugawara T, Nissato S, Otsuki T, et al. A common genetic variant of the chromogranin A-derived peptide catestatin

- is associated with atherogenesis and hypertension in a Japanese population. *Endocr J.* (2015) 62:797–804. doi: 10.1507/endocrj.EJ14-0471
65. Biswas N, Vaingankar SM, Mahata M, Das M, Gayen JR, Taupenot L, et al. Proteolytic cleavage of human chromogranin A containing naturally occurring catestatin variants: Differential processing at catestatin region by plasmin. *Endocrinology.* (2008) 149:749–57. doi: 10.1210/en.2007-0838
 66. Salem RM, Cadman PE, Chen Y, Rao F, Wen G, Hamilton BA, et al. Chromogranin A polymorphisms are associated with hypertensive renal disease. *J Am Soc Nephrol.* (2008) 19:600–14. doi: 10.1681/ASN.2007070754
 67. Wang D, Liu T, Shi S, Li R, Shan Y, Huang Y, et al. Chronic administration of catestatin improves autonomic function and exerts cardioprotective effects in myocardial infarction rats. *J Cardiovasc Pharmacol Ther.* (2016) 21:526–35. doi: 10.1177/1074248416628676
 68. Meng L, Wang J, Ding WH, Han P, Yang Y, Qi LT, et al. Plasma catestatin level in patients with acute myocardial infarction and its correlation with ventricular remodelling. *Postgrad Med J.* (2013) 89:193–6. doi: 10.1136/postgradmedj.2012-131060
 69. Liu L, Ding W, Zhao F, Shi L, Pang Y, Tang C. Plasma levels and potential roles of catestatin in patients with coronary heart disease. *Scand Cardiovasc J.* (2013) 47:217–24. doi: 10.3109/14017431.2013.794951
 70. Pei Z, Ma D, Ji L, Zhang J, Su J, Xue W, et al. Usefulness of catestatin to predict malignant arrhythmia in patients with acute myocardial infarction. *Peptides.* (2014) 55:131–5. doi: 10.1016/j.peptides.2014.02.016
 71. Zhu D, Xie H, Wang X, Liang Y, Yu H, Gao W. Correlation of plasma catestatin level and the prognosis of patients with acute myocardial infarction. *PLoS ONE.* (2015) 10:1–14. doi: 10.1371/journal.pone.0122993
 72. Xu W, Yu H, Li W, Gao W, Guo L, Wang G. Plasma catestatin: a useful biomarker for coronary collateral development with chronic myocardial ischemia. *PLoS ONE.* (2016) 11:1–11. doi: 10.1371/journal.pone.0149062
 73. Zhu D, Xie H, Wang X, Liang Y, Yu H, Gao W. Catestatin-A novel predictor of left ventricular remodeling after acute myocardial infarction. *Sci Rep.* (2017) 7:44168. doi: 10.1038/srep44168
 74. Xu W, Yu H, Wu H, Li S, Chen B, Gao W. Plasma Catestatin in Patients with acute coronary syndrome. *Cardiol.* (2017) 136:164–9. doi: 10.1159/000448987
 75. Kojima M, Ozawa N, Mori Y, Takahashi Y, Watanabe-Kominato K, Shirai R, et al. Catestatin prevents macrophage-driven atherosclerosis but not arterial injury-induced neointimal hyperplasia. *Thromb Haemost.* (2018) 118:182–94. doi: 10.1160/TH17-05-0349
 76. Zhu D, Wang F, Yu H, Mi L, Gao W. Catestatin is useful in detecting patients with stage B heart failure. *Biomarkers.* (2011) 16:691–7. doi: 10.3109/1354750X.2011.629058
 77. Liu L, Ding W, Li R, Ye X, Zhao J, Jiang J, et al. Plasma levels and diagnostic value of catestatin in patients with heart failure. *Peptides.* (2013) 46:20–5. doi: 10.1016/j.peptides.2013.05.003
 78. Peng F, Chu S, Ding W, Liu L, Zhao J, Cui X, et al. The predictive value of plasma catestatin for all-cause and cardiac deaths in chronic heart failure patients. *Peptides.* (2016) 86:112–7. doi: 10.1016/j.peptides.2016.10.007
 79. Wołowiec Ł, Rogowicz D, Banach J, Gilewski W, Sinkiewicz W, Grzesk G. Catestatin as a new prognostic marker in stable patients with heart failure with reduced ejection fraction in two-year follow-up. *Dis Markers.* (2020) 2020:8847211. doi: 10.1155/2020/8847211
 80. Borovac JA, Glavas D, Susilovic Grabovac Z, Supe Domic D, Stanisic L, D'Amario D, et al. Circulating sST2 and catestatin levels in patients with acute worsening of heart failure: a report from the CATSTAT-HF study. *ESC Hear Fail.* (2020) 7:2818–28. doi: 10.1002/eihf.12882
 81. Sun H, Xian W, Geng L, Li E, Peng Z, Tian J. Increased plasma level of catestatin might be associated with poor prognosis in hemodialysis patients. *Int Urol Nephrol.* (2017) 49:1063–9. doi: 10.1007/s11255-017-1528-8
 82. Izci S, Acar E, Inanir M. Plasma catestatin level predicts sPESI score and mortality in acute pulmonary embolism. *Arch Med Sci Atheroscler Dis.* (2020) 5:49–56. doi: 10.5114/amsad.2020.95562
 83. Tütün N, Güralp O, Gök K, Hamzaoglu K, Oner YO, Makul M, et al. Serum catestatin level is increased in women with preeclampsia. *J Obstet Gynaecol.* (2022) 42:55–60. doi: 10.1080/01443615.2021.1873922
 84. Liu X, Dang W, Liu H, Song Y, Li Y, Xu W. Associations between chronic work stress and plasma chromogranin A/catestatin among healthy workers. *J Occup Health.* (2022) 64:e12321. doi: 10.1002/1348-9585.12321
 85. Simunovic M, Supe-Domic D, Karin Z, Degoricija M, Paradzik M, Bozic J, et al. Serum catestatin concentrations are decreased in obese children and adolescents. *Pediatr Diabetes.* (2019) 20:549–55. doi: 10.1111/pedi.12825
 86. Kim J, Lee S, Bhattacharjee R, Khalyfa A, Kheirandish-Gozal L, Gozal D. Leukocyte telomere length and plasma catestatin and myeloid-related protein 8/14 concentrations in children with obstructive sleep apnea. *Chest.* (2010) 138:91–9. doi: 10.1378/chest.09-2832
 87. Borovac JA, Dogas Z, Supe-Domic D, Galic T, Bozic J. Catestatin serum levels are increased in male patients with obstructive sleep apnea. *Sleep Breath.* (2019) 23:473–81. doi: 10.1007/s11325-018-1703-x
 88. Angelone T, Quintieri AM, Pasqua T, Gentile S, Tota B, Mahata SK, et al. Phosphodiesterase type-2 and NO-dependent S-nitrosylation mediate the cardioinhibition of the antihypertensive catestatin. *Am J Physiol Circ Physiol.* (2011) 302:H431–H44. doi: 10.1152/ajpheart.00491.2011
 89. Imbrogno S, Garofalo F, Cerra MC, Mahata SK, Tota B. The catecholamine release-inhibitory peptide catestatin (chromogranin A344–364) modulates myocardial function in fish. *J Exp Biol.* (2010) 213:3636–43. doi: 10.1242/jeb.045567
 90. Angelone T, Quintieri AM, Pasqua T, Filice E, Cantafio P, Scavello F, et al. The NO stimulator, Catestatin, improves the Frank-Starling response in normotensive and hypertensive rat hearts. *Nitric Oxide.* (2015) 50:10–9. doi: 10.1016/j.niox.2015.07.004
 91. Alam MJ, Gupta R, Mahapatra NR, Goswami SK. Catestatin reverses the hypertrophic effects of norepinephrine in H9c2 cardiac myoblasts by modulating the adrenergic signaling. *Mol Cell Biochem.* (2019) 464:205–19. doi: 10.1007/s11010-019-03661-1
 92. Liao F, Zheng Y, Cai J, Fan J, Wang J, Yang J, et al. Catestatin attenuates endoplasmic reticulum induced cell apoptosis by activation type 2 muscarinic acetylcholine receptor in cardiac ischemia/reperfusion. *Sci Rep.* (2015) 5:16590. doi: 10.1038/srep16590
 93. Penna C, Alloatti G, Gallo MP, Cerra MC, Levi R, Tullio F, et al. Catestatin improves post-ischemic left ventricular function and decreases ischemia/reperfusion injury in heart. *Cell Mol Neurobiol.* (2010) 30:1171–9. doi: 10.1007/s10571-010-9598-5
 94. Perrelli MG, Tullio F, Angotti C, Cerra MC, Angelone T, Tota B, et al. Catestatin reduces myocardial ischaemia/reperfusion injury: Involvement of PI3K/Akt, PKCs, mitochondrial KATP channels and ROS signalling. *Pflugers Arch.* (2013) 465:1031–40. doi: 10.1007/s00424-013-1217-0
 95. Penna C, Pasqua T, Amelio D, Perrelli MG, Angotti C, Tullio F, et al. Catestatin increases the expression of anti-apoptotic and pro-angiogenetic factors in the post-ischemic hypertrophied heart of SHR. *PLoS ONE.* (2014) 9:e102536. doi: 10.1371/journal.pone.0102536
 96. Chu SY, Peng F, Wang J, Liu L, Meng L, Zhao J, et al. Catestatin in defense of oxidative-stress-induced apoptosis: a novel mechanism by activating the beta2 adrenergic receptor and PKB/Akt pathway in ischemic-reperfused myocardium. *Peptides.* (2020) 123:170200. doi: 10.1016/j.peptides.2019.170200
 97. Brar BK, Helgeland E, Mahata SK, Zhang K, O'Connor DT, Helle KB, et al. Human catestatin peptides differentially regulate infarct size in the ischemic-reperfused rat heart. *Regul Pept.* (2010) 165:63–70. doi: 10.1016/j.regpep.2010.07.153
 98. Tamm C, Benzi R, Papageorgiou I, Tardy I, Lerch R. Substrate competition in postischemic myocardium: effect of substrate availability during reperfusion on metabolic and contractile recovery in isolated rat hearts. *Circ Res.* (1994) 75:1103–12. doi: 10.1161/01.RES.75.6.1103
 99. Chen H, Liu D, Ge L, Wang T, Ma Z, Han Y, et al. Catestatin prevents endothelial inflammation and promotes thrombus resolution in acute pulmonary embolism in mice. *Biosci Rep.* (2019) 39:1–10. doi: 10.1042/BSR20192236
 100. Wang X, Xu S, Liang Y, Zhu D, Mi L, Wang G, et al. Dramatic changes in catestatin are associated with hemodynamics in acute myocardial infarction. *Biomarkers.* (2011) 16:372–7. doi: 10.3109/1354750X.2011.578260
 101. Ormazabal V, Nair S, Elfeky O, Aguayo C, Salomon C, Zuñiga FA. Association between insulin resistance and the

- development of cardiovascular disease. *Cardiovasc Diabetol*. (2018) 17:1–14. doi: 10.1186/s12933-018-0762-4
102. Muntjewerff EM, Parv K, Mahata SK, van Riessen NK, Phillipson M, Christoffersson G, et al. The anti-inflammatory peptide Catestatin blocks chemotaxis. *J Leukoc Biol*. (2021) 1–6. doi: 10.1002/JLB.3CRA1220-790RR
103. Egger M, Beer AGE, Theurl M, Schgoer W, Hotter B, Tatarczyk T, et al. Monocyte migration: a novel effect and signaling pathways of catestatin. *Eur J Pharmacol*. (2008) 598:104–11. doi: 10.1016/j.ejphar.2008.09.016
104. Aung G, Niyonsaba F, Ushio H, Kajiwara N, Saito H, Ikeda S, et al. Catestatin, a neuroendocrine antimicrobial peptide, induces human mast cell migration, degranulation and production of cytokines and chemokines. *Immunology*. (2011) 132:527–39. doi: 10.1111/j.1365-2567.2010.03395.x
105. Crispe IN. The liver as a lymphoid organ. *Annu Rev Immunol*. (2009) 27:147–63. doi: 10.1146/annurev.immunol.021908.132629
106. Dasgupta A, Bandyopadhyay GK, Ray I, Bandyopadhyay K, Chowdhury N, De RK, et al. Catestatin improves insulin sensitivity by attenuating endoplasmic reticulum stress: In vivo and in silico validation. *Comput Struct Biotechnol J*. (2020) 18:464–81. doi: 10.1016/j.csbj.2020.02.005
107. Wollam J, Mahata S, Riopel M, Hernandez-Carretero A, Biswas A, Bandyopadhyay GK, et al. Chromogranin A regulates vesicle storage and mitochondrial dynamics to influence insulin secretion. *Cell Tissue Res*. (2017) 368:487–501. doi: 10.1007/s00441-017-2580-5
108. Dominguez Rieg JA, Chirasani VR, Koepsell H, Senapati S, Mahata SK, Rieg T. Regulation of intestinal SGLT1 by catestatin in hyperleptinemic type 2 diabetic mice. *Lab Investig*. (2016) 96:98–111. doi: 10.1038/labinvest.2015.129

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zalewska, Kmiec and Sworczak. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Gdańsk, dnia 05.09.2023

lekierz, Ewa Zalewska
(tytuł zawodowy, imię i nazwisko)

OŚWIADCZENIE

Jako współautor pracy pt."Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas" oświadczam, iż mój własny wkład merytoryczny w przygotowanie, przeprowadzenie i opracowanie badań oraz przedstawienie pracy w formie publikacji to:

opracowanie metodyki pracy, uzyskanie zgody Komisji Bioetycznej, rekrutacja grupy badanej i kontrolnej, w tym przeprowadzenie badania podmiotowego i przedmiotowego, elektrokardiografii, całodobowego pomiaru ciśnienia tętniczego metodą holter oraz zabezpieczenie materiału biologicznego celem diagnostyki laboratoryjnej; uzyskane dane opracowałem statystycznie i napisałem pierwszą wersję manuskryptu.

Oświadczam, iż samodzielna i możliwa do wyodrębnienia część ww. pracy wykazuje na mój indywidualny wkład przy opracowywaniu koncepcji, wykonywaniu części eksperymentalnej, opracowaniu i interpretacji wyników tej pracy, która wchodzi w skład rozprawy doktorskiej.


Ewa Zalewska
LEKARZ 319052

(podpis współautora)

Gdańsk, dnia 04.09.2023

dr Piotr Kmiec
(tytuł zawodowy, imię i nazwisko)

OŚWIADCZENIE

Jako współautor pracy pt. "Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas" oświadczam, iż mój własny wkład merytoryczny w przygotowanie, przeprowadzenie i opracowanie badań oraz przedstawienie pracy w formie publikacji to:
udział w planowaniu metodyki pracy, rekrutacji grupy badanej i opracowaniu statystycznym danych oraz korekta manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie w/w pracy przez lek Ewę Zalewską jako część rozprawy doktorskiej w formie spójnego tematycznie zbioru artykułów opublikowanych w czasopismach naukowych.

Oświadczam, iż samodzielna i możliwa do wyodrębnienia część ww. pracy wykazuje indywidualny wkład lek Ewy Zalewskiej przy opracowywaniu koncepcji, wykonywaniu części eksperymentalnej, opracowaniu i interpretacji wyników tej pracy.



(podpis współautora)

Gdańsk, dnia 05.09.2023

lekierz, Jakub Sobolewski
(tytuł zawodowy, imię i nazwisko)

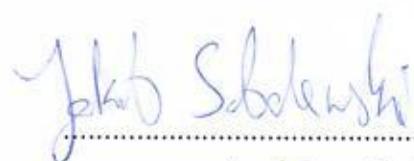
OŚWIADCZENIE

Jako współautor pracy pt. "Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas" oświadczam, iż mój własny wkład merytoryczny w przygotowanie, przeprowadzenie i opracowanie badań oraz przedstawienie pracy w formie publikacji to:

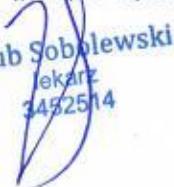
wykonanie u uczestników badania echokardiografii i oceny ultrasonograficznej kompleksu intima - media tętnicy szyjnej wspólnej oraz weryfikację ostatecznej wersji manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie w/w pracy przez lek Ewę Zalewską jako część rozprawy doktorskiej w formie spójnego tematycznie zbioru artykułów opublikowanych w czasopismach naukowych.

Oświadczam, iż samodzielna i możliwa do wyodrębnienia część ww. pracy wykazuje indywidualny wkład lek Ewy Zalewskiej przy opracowywaniu koncepcji, wykonywaniu części eksperymentalnej, opracowaniu i interpretacji wyników tej pracy.



(podpis współautora)


Jakub Sobolewski
lekierz
3482514

Gdańsk, dnia 05.09.2023

dr Andrzej Koprowski
(tytuł zawodowy, imię i nazwisko)

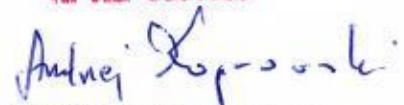
OŚWIADCZENIE

Jako współautor pracy pt. "Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas" oświadczam, iż mój własny wkład merytoryczny w przygotowanie, przeprowadzenie i opracowanie badań oraz przedstawienie pracy w formie publikacji to:
wykonanie u uczestników badania echokardiografii i oceny ultrasonograficznej kompleksu intima - media tętnicy szyjnej wspólnej oraz weryfikację ostatecznej wersji manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie w/w pracy przez lek Ewę Zalewską jako część rozprawy doktorskiej w formie spójnego tematycznie zbioru artykułów opublikowanych w czasopismach naukowych.

Oświadczam, iż samodzielna i możliwa do wyodrębnienia część ww. pracy wykazuje indywidualny wkład lek Ewy Zalewskiej przy opracowywaniu koncepcji, wykonywaniu części eksperymentalnej, opracowaniu i interpretacji wyników tej pracy.

dr med. Andrzej Koprowski
SPECJALISTA KARDIOLOG
SPECJALISTA CHORÓB WEWNĘTRZNYCH
NPWZ 9955380



.....
(podpis współautora)

Gdańsk, dnia...04.09.2023...

prof. dr hab. n. med. Krzysztof Sworczak
(tytuł zawodowy, imię i nazwisko)

OŚWIADCZENIE

Jako współautor pracy pt. "Low catestatin as a risk factor for cardiovascular disease – assessment in patients with adrenal incidentalomas" oświadczam, iż mój własny wkład merytoryczny w przygotowanie, przeprowadzenie i opracowanie badań oraz przedstawienie pracy w formie publikacji to:
nadzór merytoryczny oraz korekta manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie w/w pracy przez lek Ewę Zalewską jako część rozprawy doktorskiej w formie spójnego tematycznie zbioru artykułów opublikowanych w czasopismach naukowych.

Oświadczam, iż samodzielna i możliwa do wyodrębnienia część ww. pracy wykazuje indywidualny wkład lek Ewy Zalewskiej przy opracowywaniu koncepcji, wykonywaniu części eksperymentalnej, opracowaniu i interpretacji wyników tej pracy.

Kierownik
Katedry i Kliniki Endokrynologii
i Chorób Wewnętrznych
prof. dr hab. Krzysztof Sworczak

.....
(podpis współautora)

Gdańsk, dnia 04.09.2023

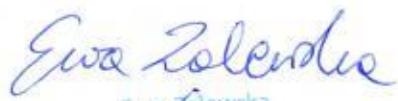
lekierz, Ewa Zalewska
(tytuł zawodowy, imię i nazwisko)

OŚWIADCZENIE

Jako współautor pracy pt. "Role of Catestatin in the Cardiovascular System and Metabolic Disorders" oświadczam, iż mój własny wkład merytoryczny w przygotowanie, przeprowadzenie i opracowanie badań oraz przedstawienie pracy w formie publikacji to:

przegląd literatury, napisanie pierwszej wersji manuskryptu oraz opracowanie rycin i tabel.

Oświadczam, iż samodzielna i możliwa do wyodrębnienia część ww. pracy wykazuje na mój indywidualny wkład w powstanie artykułu poglądowego wchodzącego w skład rozprawy doktorskiej.


Ewa Zalewska
LEKARZ 3319052

.....
(podpis współautora)

Gdańsk, dnia 04.09.2023

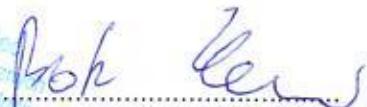
dr Piotr Kmiec
(tytuł zawodowy, imię i nazwisko)

OŚWIADCZENIE

Jako współautor pracy pt. "Role of Catestatin in the Cardiovascular System and Metabolic Disorders" oświadczam, iż mój własny wkład merytoryczny w przygotowanie, przeprowadzenie i opracowanie badań oraz przedstawienie pracy w formie publikacji to: przegląd literatury i korekta merytoryczna manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie w/w pracy przez lek Ewę Zalewską jako część rozprawy doktorskiej w formie spójnego tematycznie zbioru artykułów opublikowanych w czasopismach naukowych.

Oświadczam, iż samodzielna i możliwa do wyodrębnienia część ww. pracy wykazuje indywidualny wkład lek Ewy Zalewskiej przy opracowywaniu koncepcji, wykonywaniu części eksperymentalnej, opracowaniu i interpretacji wyników tej pracy.



(podpis współautora)

Gdańsk, dnia 04.09.2023

prof. dr hab. n. med. Krzysztof Sworczak
(tytuł zawodowy, imię i nazwisko)

OŚWIADCZENIE

Jako współautor pracy pt. "Role of Catestatin in the Cardiovascular System and Metabolic Disorders" oświadczam, iż mój własny wkład merytoryczny w przygotowanie, przeprowadzenie i opracowanie badań oraz przedstawienie pracy w formie publikacji to: nadzór merytoryczny oraz korekta manuskryptu.

Jednocześnie wyrażam zgodę na przedłożenie w/w pracy przez lek Ewę Zalewską jako część rozprawy doktorskiej w formie spójnego tematycznie zbioru artykułów opublikowanych w czasopismach naukowych.

Oświadczam, iż samodzielna i możliwa do wyodrębnienia część ww. pracy wykazuje indywidualny wkład lek Ewy Zalewskiej przy opracowywaniu koncepcji, wykonywaniu części eksperymentalnej, opracowaniu i interpretacji wyników tej pracy.

Kierownik
Katedry i Kliniki Endokrynologii
i Chorób Wewnętrznych
prof. dr hab. Krzysztof Sworczak

.....
(podpis współautora)